



Review Article

Controlling vector-borne health issues and *in-vitro* effects of Nanoemulsion of essential Oils: A ReviewAnupam Kumar Pal^{1*}, Nisha Sharma²¹ Research Scholar, University Institute of Pharmacy, C.S.J.M. University, Kanpur, U.P, India² Associate Professor, University Institute of Pharmacy, C.S.J.M. University, Kanpur, U.P, India

ABSTRACT

Controlling the parasitic and infectious diseases is a permanent health issue worldwide, requiring innovative methods for the prevention & treatment of illness triggered by parasites. Controlling vector and its intermediate hosts is an effective method for anticipation of human and animal's diseases. It is essential to have bioactive components that act competently on the agents which produce the illnesses. Synthetic agents have strong surplus effects in humans & other animals, and they cause biological toxicity, affecting animals, plants and disturbing the local environment. Many studies have reported the effect of the Essential Oils (EOs). Essential Oils extracted from the medicinal plants are generally used as insect repellents worldwide. They are very safe and favorable to the environment with minimum ill-effects on animals and public health. It helps to control the vectors and also applicable against pathogens. Commonly Essential Oils easily degrade and cause less environmental pollution. Problems associated to solubility as well as stability lead to the improvement of effective carriers for formulations containing Essential Oils that is nanoemulsion. Nanoemulsion is a colloidal dispersion system, thermodynamically stable, prepared by two different non-miscible liquids assorted with emulsifying agents such as surfactants and co-surfactants, to form a single phase. Nanoemulsion a novel drug delivery system can be formulated by using two different techniques, the persuasion technique and the brute force technique. This review describes some studies accomplished with nanoemulsions as carriers of Essential Oils that have repellent, larvicidal, acaricidal, insecticidal and antiparasitic activities. Thus it can be used as substitutes in the vector control of parasitic and communicable diseases.

Keywords: Nanoemulsion; essential oils; vector control; infectious diseases, insecticidal.

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INTRODUCTION

The Essential Oils (EOs) are complex combinations of unstable organic components which are obtained from the secondary metabolites in plants. They are constituted by two main compounds that are hydrocarbons (such as terpenes & sesquiterpenes) and oxygenated components (such as alcohols, aldehydes, esters, ethers, ketones, lactones, phenols and phenol ethers) [1]. These compounds are commonly responsible for the typical odor of plants. More than three thousands essential Oils are identified, and about 10 to 15% of them have commercial significance in different fields such as cosmetics, nutrition, and pharmaceutical industries [2].

Hence, they are generally known as safe by the United State Food and Drug Administration (USFDA) [3]. Their conformation can vary significantly with plant species and biodiversities [4]. The aromatic plants with essential oils shows several effects such as repellents, low-risk insecticides, antifungals, antivector growth regulators, oviposition deterrents and antifeedants. The use of essential oils for these activities had increased due to its popularity among organic farmers and environmentally conscious consumers [5-8]. The production of essential oils is done generally by distillation techniques (such as hydro, steam & dry) and/or mechanical cold pressing of plants [9]. Mostly the "Clavenger steam distillation apparatus" is used for

production of essential oils globally. It has been developed and extended for large scale production of essential oils in industries. In recent times, contemporary essential oil isolation techniques have been initiated such as microwave-assisted processing (MAP) & supercritical fluid isolation (SFI) techniques^[10]. Essential oils could cover, on usual, among 25 and 90 different compounds. The constituents of essential oils are usually designated to two different phytochemical groups: terpenoids, example monoterpenes and sesquiterpenes of low mol. wt. and to a smaller range, Phenyl-propanoids example volatile phenolic compounds. Terpenoid is the chief component of the essential oils. Monoterpenes are present in the essential oils which may contain terpenes such as hydrocarbons (α -pinene and D-limonene), ethers (1, 8-cineol), aldehydes (cinnamaldehyde & citronellal), lactones (artemisin), alcohols (cadinol) and ketones (thujone). Wide variety structures of sesquiterpenes have more than 100 skeletons and enlargement of the series to fifteen carbons may enhance the quantity of potential cyclizations. Less common aromatic compound includes Phenyl-propanoids like p-allylanisole & phenolic components like thymol. The composition of essential oils may vary in different plant species. For example eucalyptus *Eucalyptus globulus*, belonging to family Myrtaceae with chief component of the essential oil monoterpene 1, 8-cineol, whereas coriander *Coriandrum sativum* belonging to family Umbelliferae, has sesquiterpenes linalool as the main component. The oil isolated from leaves of *Eucalyptus globules* have allelopathic & insecticidal property which acts as a natural pesticides^[11-12]. In various studies, with fumigation effects^[13] and repellency^[14], the effectiveness of these oils had been validated^[15]. The mixture and growth of essential oils in plants are linked with the occurrence of compound and to very specific structures like glandular trichomes (*Lamiaceae*) and secretory cavities (*Myrtaceae*, *Rutaceae*). Depending on the species, the essential oils could be stored in many plant organs such as, in flowers, fruits, leaves, seeds, wood, roots and rhizomes^[16]. Biologically, the essential oils have various effective properties such as insecticidal effects, and reduce growth of insects and repellent activity. They are efficiently significant to regulate phytophagous insects and also regulate local and plot insects. The mechanism of action (MOA) of the components present in essential oils on pests are generally through neurotoxic effects, through the inhibition of acetylcholinesterase (AChE)^[17], functionality disruption of gamma-aminobutyric acid (GABA) receptors [18], and as agonist of octopaminergic system^[19]. Even though the biological effects of specific chemical components of essential oils are commonly known, the toxic effect of their mixtures is a very difficult aspect to evaluate. The most attractive features of essential oils are that they are, in general, low-risk products on animals, and their environmental determination is short. Their toxicity for mammals is low, having values of oral LD₅₀ that varies from 1000 to 1950 mg kg⁻¹ in rats. In addition, due to their use as medicines, some essential oils are relatively well studied experimentally and clinically. The toxic effects of essential oil components are classified into three basic classes based on the toxicological measures^[20]. Class 1 with low functionality and less oral toxicity like

limonene, Class 2 with some functionality & midway toxicity for example menthofuran and Class 3 with sensitive functionality & high possible toxicity for example elemicin. Based on classification and other toxicological measures, a different procedure has industrialized to estimate the protection from essential oils^[21]. The essential oils can be promising antiprotozoal agents. The compounds like terpinen-4-oland/or thymol & camphor the chief component have antiprotozoal activity^[22, 23].

2. NANOEMULSION AND NANOTECHNOLOGY:

An emulsion is a combination of two or more liquid which are generally non-miscible. Macroscopic separation of both liquid phases is measured by accumulation of non-ionic surfactant. When the emulsion size ranges between 10 to 200 nm, it is called as "Nanoemulsion". Nanoemulsion can be formed through dispersion of the oil-in-water (o/w) and/or water-in-oil (w/o) and allow the incorporation of different active substances. They have some individual characteristics, like transparency when observed by the naked eye, with bluish reflection, owing to the diffusion of light b/w the nanoparticles^[24]. The very lesser extent of particles also discusses greater resistance to the effects of incineration and sedimentation, because the effect of gravity is smaller on them^[25]. Surfactants have an essential role in the formulation of Nanoemulsion systems. Interfacial tension produced owing to the immiscibility of mixture, liquid systems is made compact by the use of surfactant. The formulation process of the Nanoemulsion is passed out b/w the lower to higher energy emulsification method. Lower energy method includes the "phase inversion method", but higher energy method includes ultrasound and "high-pressure homogenization". In addition the system displays lower interfacial tension that simplifies the diffusion of other ingredients and the major advantage is when the combination of extremely lipophilic substances like essential oils, into the aqueous systems is compulsory^[26]. The nanoemulsions have various applications in numerous areas like medications, cosmetics, agriculture, etc.^[27]. In recent development, different methods are being accepted in the advancement of Nanoemulsion for repellent activity, larvicidal effects and insecticidal effects using essential oils. Nanotechnology is a trans-disciplinary and promising area of science owing to its miscellaneous potential of application, belonging to the field of electronic and mechanical engineering, telecommunications, industrial chemistry, civil construction, Health, development of nanostructure drugs, biological carriers and devices for diagnosis^[28, 29]. Advancement of technologies in electron microscopy dates back to 1980s, beneficial to observe study, manipulate and develop nanostructure systems, which led to technical revolution in various fields of science^[30]. Nano-metric materials have a size in the range between 0.1-200 nm.^[31] Nanotechnology had provided substitute for the research and development of novel drug delivery systems in the biological area and health sciences. A field that delivers a number of patient benefits like dose reduction, lower number of administrations, improved therapeutic efficacy and reduction in side effects. Nano-systems have an advantage of protecting active materials against mechanisms of inactivation and

degradation, also providing the combination of materials with different polarity in relation to matrix, to promote sustained release and/or targeting of drug action in a definite tissue [32, 33].

3. NANOEMULSION OF ESSENTIAL OIL:

In recent years, various scientific studies have been described for the use of nanoemulsions as appropriate carriers of active essential oils. They are mainly classified into 3 types which are formed depending on the composition such as:

- a) Oil in water Nanoemulsions wherein oil droplets are dispersed in the continuous aqueous phase
- b) Water in oil Nanoemulsions wherein water droplets are dispersed in the continuous oil phase
- c) Bi-continuous Nanoemulsions wherein micro domains of oil and water are interdispersed within the system [34, 35].

Their composition and preparation is very easy with high thermodynamic stability with low production cost and the possibility of production on an industrial level provide a significant advantage to this technology over the biological and pharmacological usage of the essential oils [36, 37]. Nanoemulsions are encouraging tools to fight with vector-borne diseases in public control of communicating mediators. It is probably owing to the characteristic physico-chemical possessions of the Nano-metric emulsion system [38]. The dimension of the nanoemulsion in nanometer increases its specification and distribution target, resulting in more efficiency than marketed pesticides. Moreover, nanoemulsions pesticides has lesser surfactant concentration than microemulsion, and surfactants are significantly more eco-friendly & economically feasible. Essential oils dispersed in various nano-formulations include numerous MOA and de-regulation of the growth-hormone which inclines to stopover the insect flaking that leads to its deaths and also enzymatic inhibition, between others [39-41]. According to the recent studies, the organic effects of nanoemulsions on etiological mediators of parasitic source had reported that the use of nanoemulsions have potentially increased in the control of infectious/parasitic diseases.

4. BIOLOGICAL ACTIVITIES OF ESSENTIAL OIL NANOEMULSIONS

4.1. LARVICIDAL & PUPICIDAL ACTIVITY

The mosquito species, *Culex quinquefasciatus* are substitute as repeated vector for *Wuchereria bancrofti*, a humanoid filarial roundworm which originate filariasis. It is a disease which generally causes lymphedema that leads to patient illness and are potentially lethal [42]. Eucalyptus oil loaded Nanoemulsion developed showed larvicidal activity as well as pupicidal activity against *Culex quinquefasciatus*. The treated larvae/pupa exhibited lower protein levels, as well as significant fall in the stages of AChE and acid/basic phosphatase. The nanoemulsions could be used as a nontoxic and active substitute in restraint of vectors [43]. The nanoemulsions of essential oils (*Ocimum basilicum*) exhibited larvicidal effects in 3rd instar larvae and pupicidal activity on *Aedes aegypti* (dengue-transmitting mosquitoes). Essential Oil compositions contained about 89% of methyl

chavicol (estragole), Phenylpropanoids with insecticidal property [44]. The study reported that about 100 fold dilution of Nanoemulsions initiated larvicidal effects of 60 to 70% after period of 60 to 75 minute, separately. Complete losses of larval at this concentration were observed after a submission period of 90 minute. While, diluted (about ten-fold) Nanoemulsion induced 100% larvicidal effects on *Aedes aegypti* in 15 minute [43]. Brazilian scientists established Nanoemulsions using *Rosmarinus officinalis* essential oil as an active component for 4th instar larvae of *Aedes aegypti*, in 2015. The mortality ratios were found after 24 to 48 hours of interaction with the Nanoemulsion, which induced 85 to 90% mortality, respectively [45]. 1, 8-cineol is the chief component of *Rosmarinus officinalis* which showed potent larvicidal activity [46, 47]. The validated non-emulsified *R. officinalis* essential oil showed a DL₉₅ of 409 ppm after 24 hours of interaction indicates the greater larvicidal efficiency of the Nanoemulsion [48].

The Nanoemulsions of *Cinnamomum zelanycum* essential oil (5% conc.) with chief component cinnamaldehyde caused 70% death of mealworm (*Alphitobius diaperinus*) larvae at the L8 stage after two days, and had three times more stronger effects as compared to un-emulsified essential oil within 3 days. Besides, the nanoemulsions exhibited no harmful effects on existence and imitation tests of springtails (*Folsomia candida*), indicated that nano-encapsulation of cinnamon oil meaningfully reduced its harmful effects without changing the efficiency in controlling *Aedes diaperinus*. Botas et al., (2017) reported larvicidal bioassays against *Aedes aegypti* in a study [49], wherein nanoemulsions based on essential oils of *Baccharis reticulata* was studied. D-limonene the chief component (25%) of *Baccharis reticulata* and the nanoemulsions of these essential oils showed larval mortality against *Aedes aegypti*, with LC₅₀ values of 120.10 µg/mL & 80.95 µg/mL, respectively [50]. Balasubramani et al., in a study reported the Nanoemulsions of *Vitex negundo* L. essential oils with larvicidal activity for 2nd & 3rd instar larvae, after 12 to 24 hour contact times. After 12 hour exposure period, the LC₅₀ values of both 2nd & 3rd instar larvae was 65.54 and 71.12 ppm, respectively when compared to the values of the non-emulsified Essential Oils which was 120.13 and 91.94 ppm, respectively. Similarly, after a 24 hour exposure period, the Nanoemulsion LC₅₀ values were 29.10 and 42.93 ppm, although the non-emulsified essential oil values were 76.65 and 55.87 ppm, respectively. Osanloo et.al., (a) worked on control of anopheles species, which causes malaria worldwide, by the application of Nanoemulsion loaded essential oils from *Artemisia dracunculus* belonging to family *Asteraceae*. The activity of Nanoemulsions on both 3rd & 4th instar larvae were done, with LC₅₀ or LC₉₀ of 12.13 or 16.98 ppm, respectively. P-allylanisole is the main component of *Artemisia dracunculus* [51]. Osanloo et al., (b) evaluated larvicidal activity on *Anopheles stephensi* by the act of nano-emulsified essential oils from *Anethum graveolens* which mainly consists of p-cymene and α-phellandrene as chief components. After one hour of contact with nanoemulsions, larvicidal effects of 50 and 90% were found at 40.1 and 64.9 ppm, individually, against 3rd & 4th instar larvae [52], in

comparison to *Anethum graveolens* non-emulsified essential oils, the Nanoemulsion exhibited more efficiency, since the oil LD₅₀ without nano-emulsification was 100 ppm after the same contact time [53-54]. The nanoemulsions of *Ocimum basilicum* essential oils showed larvicidal effects on 2nd & 3rd

instar larvae of *Culex quinquefasciatus*, trans- β -Guaiene (16.89%) and α -Cadinol (15.66%) the chief components of Essential Oil. Following table enumerates various essential oil Nanoemulsions with larvicidal activity (Table 1):

Table: 1.Essential Oil Nanoemulsion with Larvicidal activity:

S. no.	Common Name	Chief constituent of essential oil	Emulsifying agent	Insect
1.	Tarragon	p-Allylanisole	Tween 20	<i>Anopheles stephensi</i>
2.	True cinnamon Tree	Cinnamaldehyde	Tween 80	<i>Alphitobius diaperinus</i>
3.	Dill	p-Cymene and α -phellandrene	Tween 20	<i>Anopheles stephensi</i>
4.	Eucalyptus	1,8-cineole	Tween 80	<i>Culex quinquefasciatus</i>
5.	Sand-Rosemary	D-limonene	Tween 80	<i>Aedes aegypti</i>
6.	Cumin	Pinene and cymene	Tween 20	<i>Aedes aegypti</i> & <i>Anopheles stephensi</i>
7.	Chinese chaste tree	β -caryophyllene	Tween 80	<i>Aedes aegypti</i>
8.	Basil	Trans- β -Guaiene and α -Cadinol	Tween 80	<i>Culex quinquefasciatus</i>
9.	Rosemary	Caffeic acid and betulinic acid	Polysorbate 20	<i>Aedes aegypti</i>
10.	Basil	Methyl-chavicol	Tween 20	<i>Aedes aegypti</i>
11.	Sucupira	γ -muurolene and biciclogermacrene	Polysorbate 20	<i>Aedes aegypti</i>

4.2. MOSQUITO REPELLENT AND INSECTICIDAL ACTIVITY OF ESSENTIAL OILS

Some monoterpenes (such as α -pinene, cineole, thymol, limonene, camphor, citronellol, terpinolene, citronellal and eugenol) are common components of different essential oils, which showed mosquito repellent activities [55-58]. Among sesquiterpenes like β -caryophyllene showed strong repellent effects on *Aedes aegypti* [59]. However repellent effects of some essential oils commonly appear to be linked with the occurrence of monoterpenoids and sesquiterpenes [60-63]. Further authors have reported that phytol, a linear diterpene alcohol, have greater repellent activity for *Anopheles gambiae*. Also, the oxygenated components such as cinnamyl alcohol, geraniol, phenyl ethyl alcohol, β -citronellol and α -pinene, extracted from the essential oil of *Dianthus caryophyllum*, exhibited strong repellent effects for ticks (*I. ricinus*) [64,65]. The repellent effects of 12 to 16 essential oils from natural and striking Argentine plants and 21-isolated metabolites; 3-alcohols (thymol, benzyl alcohol and menthol) were reported as the most active towards *Pediculus humanus capitis* [66]. The repellent effects of 10 to 12 different Kenyan plants species against *Anopheles gambiae* belonging to family *Diptera*, and about 5 to 10 pure metabolites were isolated from them, the most effective chemicals such as citronellal, geraniol, caryophyllene oxide, carvacrol, perillyl alcohol, cisverbenol, cis-carveol, 4-isopropyl benzene methanol, 3-carene, thymol and myrcene showed strong repellent activity. This constituent belongs to different categories such as diterpenoid, sesquiterpenoid and acyclic, monocyclic & bicyclic monoterpenoids. Many components have shown activity on adult insects, as well as on other insect development stages [67]. The effect of several nanoformulations of *Ocimum sanctum* essential oils on *Culex quinquefasciatus* & *Aedes aegypti* has been reported. By the use of filter paper impregnation technique, the preparation containing essential oils (30%) showed 98% effect on *Aedes aegypti* and 100% effect on *Culex quinquefasciatus* [68]. The Nanoemulsion loaded citronella oil showed the repellent activity against *Aedes aegypti*, using the human-bait method,

based on standard test of World Health Organization (WHO). The Nanoemulsions containing 20% of citronella oil and glycerol-water (1:1) ratio were reported to be more protective (about 2.7 hour), which can be measured high in contrast with essential oils diluted to 10% in olive oil exhibited protection (about 56.30 min.). d-limonene & citronellal were the chief component of the essential oils (40% each) and had greater repellent effects [69]. The repellent activity of Nanoemulsions from different essential oils on adult mosquito (*Aedes aegypti*), 7 nano-formulations were prepared using different concentration of oils from hairy basil (*Ocimum americanum*), Vetiver (*Vetiveria zizanioides*) belonging to family Poaceae & citronella oil (*Cymbopogon nardus*). Using the human-bait method, the preparation with ratio 5:5:10 (vetiver, hairy basil and citronella) showed repellent effects with a protection time of 4.8 hours. Citronellal (citronella) & limonene, vetiveric acid (vetiver) and 3-carene & caryophyllene (hairy basil) were the chief components in the essential oils, and possibly represented the activity through synergic mechanisms (Table 2) [70].

4.3. ACARICIDAL ACTIVITY:

Rhipicephalus microplus is an Ixodidae tick which causes severe financial damages on livestock. It is defined as vector of tick-transmitting agents like *Babesia bigemina*, *Anaplasma marginale* & *Babesia bovis*. Some groups of researchers had described the acaricidal effects of essential oils loaded Nanoemulsion against this species [71]. The study assessed the effects of the Nanoemulsions-based *Cinnamomum verum* (5%) that induced 96% and 64% of oviposition inhibition, *in-vitro* & *in-vivo*, correspondingly. In comparison, additional study presented that the non-emulsified essential oils didn't cause oviposition inhibition. Furthermore, after 25 days, the cows which were previously invaded and treated with the Nanoemulsion were free of the parasites [71,72]. Similarly, the Nanoemulsion of *Eucalyptus globules* essential oils exhibited that the preparations at 1% & 5% inhibited parasite reproduction by 52% and 60.8%, individually [73]. The various essential oils formulated in Nanoemulsion with acaricidal activity are shown in Table 2.

Table: 2. Essential Oil Nanoemulsion-Insecticidal, repellent and acaricidal activity:

S. No.	Common Name	Main Essential Oil Compound	Emulsificant	Parasite/Insect
1.	Eucalyptus	1,8-cineole	Tween 80	<i>Rhipicephalus microplus</i> (Acaricidal activity) <i>Culex quinquefasciatus</i>
2.	Cinnamomum	Cinnamaldehyde	Tween 80	<i>Rhipicephalus microplus</i> (Acaricidal activity) <i>Aedes aegypti</i>
3.	Citronella	D-limonene and Citronellal, D-limonene and citronellal (<i>C. nardus</i>),	Montanov 82	<i>Aedes aegypti</i> (Repellent activity)
4.	Cumin	Cymene	Tween 20	<i>Aedes aegypti</i> / <i>Anopheles stephensi</i> (Repellent activity)
5.	Nutmeg	Pinene and camphene	Montanov 82	<i>Aedes aegypti</i> (Repellent activity)
6.	Holy Basil	Ursolic acid & β -caryophyllene	Tween 20	<i>Culex quinquefasciatus</i> and <i>Aedes aegypti</i> (Insecticide activity)

4.4. ETIOLOGICAL AGENTS FOR ANTIPARASITIC ACTIVITY

Trypanosoma evansi are flagellated parasite & the etiological agent of the disease called “Surra” & “Mal das Cadeiras” mainly affects in Brazilian horses and hardly affects humans [74]. The *in-vitro* trypanocidal effects of the nano-emulsified *Schinus molle* essential oils had been evaluated. The Nanoemulsions (0.5% and 1% concentration) were able to decrease the number of living parasites in 79% and 100%, correspondingly. The study outcomes were compared with the non-emulsified essential oils, showed a minor death ratio, with 64% and 70%, correspondingly [75]. The *in-vitro* effect of essential oils (*Lavandula officinalis*) Nanoemulsions against *Trichomonas vaginalis* were observed. The Nanoemulsion (100 μ g/mL concentration) showed 82% of growth resistance and demonstrated low toxicity & macrophages (about 91% viability) [76]. In an study for antiparasitic effects of nano-emulsified essential oils of *Lavandula* class, the Nanoemulsion of this oil and also *Rosmarinus officinalis* (rosemary), exhibited an antileishmanial effect for *Leishmania major*. In Iran, it is one of the most etiological agents of cutaneous leishmaniasis.

Linalool (11.22%) and 1, 8-cineol (21.98%) were the chief components of *Lavandula angustifolia* (lavender) essential oils. The Nanoemulsion of *Lavandula angustifolia* and *Rosmarinus officinalis* exhibited antileishmania effects on promastigote with $IC_{50} = 0.12 \mu$ L/mL & $IC_{50} = 0.07 \mu$ L/mL, correspondingly. The study was compared with similar non-emulsified essential oils of *Rosmarinus officinalis*, which showed $IC_{50} = 2.5 \mu$ L/mL respectively [77]. The study reported that a greater strength of the Nanoemulsion kills parasites. The effects of *Zataria multiflora* essential oil on *Echinococcus granulosus*, caused cystic echinococcosis, a zoonotic contamination with financial and human health importance globally [78]. The *in-vivo* results presented that the scolicidal effect of the Nanoemulsion (1 mg/mL concentration) were 90% and 100% after 12 to 22 min, correspondingly, although the Nanoemulsions (2 mg/mL concentration) exhibited 100% of scolicidal effect after 10 minute [79]. Thymol is the chief component of *Zataria multiflora* essential oil involved in the scolicidal activity [80-82]. It also exhibited negative effect on the germinal layer of hydatid cysts [83]. The following table shows some of the Nano emulsion essential oils with antiparasitic activity (Table 3).

Table: 3. Essential Oil Nanoemulsion for antiparasitic activity

S.No.	CommonName	Main Essential Oil Compound	Emulsificant	Parasite
1.	Avishan Shirazi	Thymol	Tween 80	Protoscoleces of the hydatid cysts
2.	Lavander and Rosemary	1,8-cineole and linalool	Tween 80	Leishmania major
3.	Lavender	1,8-cineole	Tween 80	<i>Trichomonas vaginalis</i>

The Nanoemulsions with essential oils have been used as active mediators in control of illnesses caused by several agents. The study reported the variety of targets and effectiveness of nanoemulsions when compared to additional agents already used, as well as their easiness of production [84, 85]. Technology of Nanoemulsion formulations provide various benefits with respect to the active principle and their stability, such as:

- Better protection of the active constituent against chemical or biological degradation

- Lower probability of creaming or sedimentation of droplets
- Greater contact surface of the target with the droplets that contains the active agent
- Possibility of dispersion of immiscible substances in a certain solvent, which in the case of Essential Oils is usually water, besides the simplicity of production,
- Low cost of reagents.
- Less residual damage to the environment when compared to synthetic products, widely used in modern times [85].

These facts could be verified through related studies made with nanoemulsified oils in which the essential oils alone were not able to balance the activity propagation of the nano-formulations. The variety of the components of the essential oils such as monoterpenes, sesquiterpenes and Phenylpropanoids, contributes to the diversity of mechanisms of action for controlling the vectors & etiological agents of infectious/parasitic diseases which increase the possible action of synergistic effect^[86–88].

5. CONCLUSIONS:

Nanomodification of essential oils, is hydro-immiscible in nature, expressively improves life value and efficiency as a pesticide and/or antiparasitic. Nanoformulations require a lesser amount of essential oils to develop/prepare nanoemulsions, and the use of water & nonionic surfactants also makes these Nano-formulations favorable & ecofriendly. In case of pesticide preparation, removal of the unpredictable chemical components increases their biological effects and makes “greener” method to the control of vectors of pathogenic illness. The properties of essential oil Nano-formulations such as a greater degree of delivery to the target of action, water dispersion, and stability of formulation, cost-effective and low toxicity are very effective and highly ecological.

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REFERENCE:

1. Guenther, E., 1972. The Essential Oils. Krieger Publishing Company, Florida, USA.
2. Hadis M., Lulu M., Mekonnen Y., Asfaw T. Field trials on the repellent activity of four plant products against mainly *Mansonia* population in Western Ethiopia. *Phytother. Res.* 2003; 17:202–205.
3. FAO, 1995. Flavours and Fragrances of Plant Origin, Rome.
4. Trongtokit Y., Rongsriyam Y., Komalamisra N., Apiwatnasorn C. Comparative repellency of 38 Essential Oils against mosquito bites. *Phytother. Res.* 2005; 19:303–309.
5. Zygadlo JA., Juliani HR. 2003. Recent progress in medicinal plants. In: Majundar DK., Govil JN., Singh VK., Shailaja MS., Gangal SV. (Eds.), *Phytochemistry and Pharmacology II, VIII*. Studium Press LLC, Texas, pp. 273–281.
6. Urzua A., Santander R., Echeverría J., Cabezas N., Palacios SM., Rossi Y. Insecticide properties of the Essential Oils from *Haplopappus foliosus* and *Bahia ambrosoides* against the house fly, *Musca domestica* L. *J. Chil. Chem. Soc.* 2010; 55:392–395.
7. Urzua A., Santander R., Echeverría J., Villalobos C., Palacios, S.M.; Rossi, Y. Insecticidal properties of *Peumus boldus* Mol. Essential Oil on the house fly, *Musca domestica* L. *Bol. Latinoam Caribe Plantas Med. Aromat.* 2010; 9:465–469.
8. Urzua, A., Di Cosmo D., Echeverría J., Santander R., Palacios SM., Rossi Y. Insecticidal effect of *Schinus latifolius* Essential Oil on the housefly, *Musca domestica* L. *Bol. Latinoam Caribe Plantas Med. Aromat.* 2011; 10:470–475.
9. Espinoza J., Urzua A., Bardehle L., Quiroz A., Echeverría J., Gonzalez-Teuber M. Antifeedant effects of Essential Oil, extracts, and isolated sesquiterpenes from *Pilgerodendron uviferum* (D. Don) Florin heartwood on red clover borer *Hylastinus obscurus* (Coleoptera: Curculionidae). *Molecules.* 2018; 23:1282.
10. Ferhat MA., Meklati BY., Chemat F. Comparison of different isolation methods of Essential Oil from Citrus fruits: Cold pressing, hydrodistillation and microwave dry distillation. *Flavour Fragr. J.* 2007; 22:494–504.
11. Kaufmann B., Christen P. Recent extraction techniques for natural products: Microwave-assisted extraction and pressurized solvent extraction. *Phytochemical. Anal.* 2002; 13:105–113.
12. Brooker M & Kleinig D. Field Guide to the Eucalyptus, Vol. 1. Revised edn. Melbourne and Sydney, South-Eastern Australia, Inkata Press (1990).
13. Batish DR., Singh HP., Kohli RK., & Kaur S. Eucalyptus Essential Oil as a natural pesticide. *Forest Ecology and Management.* 2008; 256:2166–2174.
14. Papachristos DP & Stamopoulos D.C. Fumigant toxicity of three Essential Oils on the eggs of *Acanthoscelides obtectus* (Say) (Coleoptera: Bruchidae). *Journal of Stored Products Research.* 2004; 40:517–525.
15. Trigg JK. Evaluation of eucalyptus-based repellent against *Anopheles* spp. in Tanzania. *Journal of the American Mosquito Control Association.* 1996; 12:243–246.
16. Kumar P., Mishra S., Malik A & Satya S. Compositional analysis and insecticidal activity of Eucalyptus globules (family: Myrtaceae) Essential Oil against housefly (*Musca domestica*). *Acta Tropica.* 2012; 122:212–218.
17. Figueiredo AC., Barroso JG., Pedro LG., Scheffer JJC. Factors affecting secondary metabolite production in plants: Volatile components and Essential Oils. *Flavour Fragr. J.* 2008; 23:213–226.
18. Mills C., Cleary BV., Walsh JJ., Gilmer JF. Inhibition of acetylcholinesterase by tea tree oil. *J. Pharm. Pharmacol.* 2004; 56:375–379.
19. Priestley CM., Williamson EM., Wafford KA., Sattelle DB. Thymol, a constituent of thyme Essential Oil, is a positive allosteric modulator of human GABA receptors and a homo-oligomeric GABA receptor from *Drosophila melanogaster*. *Br. J. Pharmacol.* 2003; 140:1363–1372.
20. Enan EE. Molecular response of *Drosophila melanogaster* tyramine receptor cascade to plant Essential Oils. *Insect Biochem. Mol. Biol.* 2005; 35:309–321.
21. Munro IC., Ford RA., Kennepohl E., Sprenger JG. Correlation of structural class with no-observed-effect levels: A proposal for establishing a threshold of concern. *Food Chemical Toxicology.* 1996; 34:829–867.
22. Smith RL., Cohen SM., Doull J., Feron VJ., Goodman JI., Mamett LJ., Portoghesi PS., Waddell WJ., Wagner BM., Hall RL. A procedure for the safety evaluation of natural flavor complexes used as ingredients in food: Essential Oils. *Food Chemical Toxicology.* 2005; 43:345–363.
23. Monzote L., Alarcón O., Setzer WN. Antiprotozoal activity of Essential Oils. *Agric. Conspec. Sci.* 2012; 77: 167–175.
24. Ramos-Lopez MA., Sanchez-Mir E., Fresan-Orozco MC., Perez-Ramos J. Antiprotozoal activity of some Essential Oils. *J. Med. Plants Res.* 2012; 6:2901–2908.
25. Salager JL., Anton RE., Anderez JM., Aubry JM. Formulation des micro-emulsions par la method HLD. In *Techniques de l'Ingenieur*, 1st ed. Editions T.I.: Paris, France, 2001; 1–20.
26. Solans C., Izquierdo P., Nolla J., Azemar N., Garcia-Celma MJ. Nano-emulsions. *Curr. Opin. Colloid Interface Sci.* 2005; 10:102–110.
27. Forgiarini A., Esquena J., Gonzalez C., Solans C. Studies of the relation between phase behavior and emulsification methods with Nanoemulsion formation. In *Trends in Colloid and Interface Science XIV*; Springer: Berlin/Heidelberg, Germany, 2000; pp. 36–39.
28. Chime SA., Kenchukwu FC., Attama AA. Nanoemulsions—advances in formulation, characterization and applications in drug delivery. In *Application of Nanotechnology in Drug Delivery*; In Tech Open: London, UK, 2014.
29. Lu WC, Huang DW, Wang CC, Yeh CH, Tsai JC, Huang YT, Li PH. Preparation, characterization, and antimicrobial activity of nanoemulsions incorporating citral essential oil. *Journal of food and drug analysis.* 2018 1; 26(1):82-9.
30. Galho AR, Cordeiro MF, Ribeiro SA, Marques MS, Antunes MF, Luz DC, Hädrich G, Muccillo-Baisch AL, Barros DM, Lima JV, Dora CL. Protective role of free and quercetin-loaded nanoemulsion against damage induced by intracerebral haemorrhage in rats. *Nanotechnology.* 2016 ;27(17):175101.
31. Chen H, Hu X, Chen E, Wu S, McClements DJ, Liu S, Li B, Li Y. Preparation, characterization, and properties of chitosan films with cinnamaldehyde nanoemulsions. *Food Hydrocolloids.* 2016; 61:662-71.
32. Singh KK, Vingkar SK. Formulation, antimalarial activity and biodistribution of oral lipid nanoemulsion of primaquine. *International Journal of Pharmaceutics.* 2008; 347(1-2):136-43.

33. Craparo EF., Bondi ML., Pitarresi G., Cavallaro G. Nanoparticulate systems for drug delivery and targeting to the central nervous system. *CNS Neurosci. Ther.* 2011; 17:670–677.
34. Sole I., Pey CM., Maestro A., Gonzalez C., Porras M., Solans C. *et al.* Nanoemulsions prepared by phase inversion composition method: preparation variables and scale up. *J Colloid Interface Sci.* 2010; 344:417-23.
35. Ravi TPU., Padma T. Nanoemulsions for drug delivery through different routes. *Res Biotechnology.* 2011; 2:1-13.
36. De Campos VEB., Ricci-Junior E., Mansur CRE. Nanoemulsions as delivery systems for lipophilic drugs. *J. Nanosci. Nanotechnol.* 2012; 12:2881–2890.
37. Gahruie HH, Ziaee E, Eskandari MH, Hosseini SM. Characterization of basil seed gum-based edible films incorporated with Zataria multiflora essential oil nanoemulsion. *Carbohydrate polymers.* 2017 Jun 15; 166:93-103.
38. Wang L., Li X., Zhang G., Dong J., Eastoe J. Oil-in-water nanoemulsions for pesticide formulations. *J. Colloid Interface Sci.* 2007; 314:230–235.
39. Hazra DK. Nano-formulations: High definition liquid engineering of pesticides for nano-formulations: High definition liquid engineering of pesticides for advanced crop protection in agriculture. *Adv. Plant. Agric. Res.* 2017; 6:1–2.
40. Mishra P., Balaji APB., Tyagi BK., Mukherjee A., Chandrasekaran N. Nanopesticides: A boon towards the control of dreadful vectors of lymphatic filariasis. In *Lymphatic Filariasis*; Springer: Singapore, 2018; 247–257.
41. Mishra P., Balaji APB., Mukherjee A., Chandrasekaran N. Bio-based nanoemulsions: An eco-safe approach towards the eco-toxicity problem. In *Handbook of Ecomaterials*; Springer: Singapore, 2018; 1–23.
42. Sugumar S., Clarke SK., Nirmala MJ., Tyagi BK., Mukherjee A., Chandrasekaran N. Nanoemulsion of Eucalyptus oil and its Larvicidal activity against *Culex quinquefasciatus*. *Bull. Entomol. Res.* 2014; 104:393–402.
43. Ghosh V., Sugumar S., Mukherjee A., Chandrasekaran N. Cinnamon oil Nanoemulsion formulation by ultrasonic emulsification: Investigation of its bactericidal activity. *J. Nanosci. Nanotechnol.* 2013; 13:114–122.
44. Chang CL., Kyu Cho I., Li QX. Insecticidal activity of basil oil, trans-anethole, estragole, and linalool to adult fruit flies of *Ceratitis capitata*, *Bactrocera dorsalis*, and *Bactrocera cucurbitae*. *J. Econ. Entomol.* 2009; 102: 203–209.
45. Duarte JL., Amado JRR., Oliveira AEMFM., Cruz RAS., Ferreira AM., Souto RNP., Falcão DQ., Carvalho JCT., Fernandes CP. Evaluation of Larvicidal activity of a Nanoemulsion of *Rosmarinus officinalis* Essential Oil. *Rev. Bras. Farmacogn.* 2015; 25:189–192.
46. Conti B., Canale A., Bertoli A., Gozzini F., Pistelli L. Essential Oil composition and Larvicidal activity of six Mediterranean aromatic plants against the mosquito *Aedes albopictus* (Diptera: Culicidae). *Parasitol. Res.* 2010, 107, 1455–1461.
47. Conti B., Canale A., Cioni PL., Flamini G. Repellence of Essential Oils from tropical and Mediterranean Lamiaceae against *Sitophilus zeamais*. *Bull. Insectol.* 2010; 63:197–202.
48. Volpato A., Baretta D., Zortéa T., Campigotto G., Galli GM., Glombowsky P., Santos RCV., Quatrin PM., Ourique AF., Baldissera MD. Larvicidal and insecticidal effect of *Cinnamomum zeylanicum* oil (pure and nanostructure) against mealworm (*Alphitobius diaperinus*) and its possible environmental effects. *J. Asia Pac. Entomol.* 2016; 19:1159–1165.
49. Botas GDS., Cruz RAS., De Almeida FB., Duarte JL., Araújo RS., Souto RNP., Ferreira R., Carvalho JCT., Santos MG., Rocha L et al. *Baccharis reticularia* DC. and limonene nanoemulsions: Promising Larvicidal agents for *Aedes aegypti* (Diptera: Culicidae) control. *Molecules* 2017; 22:1990.
50. Balasubramani S., Rajendhiran T., Moola A., Kumari B. Development of Nanoemulsion from *Vitex negundo* L. Essential Oil and their efficacy of antioxidant, antimicrobial and Larvicidal activities (*Aedes aegypti* L.). *Environ. Sci. Pollut. Res.* 2017; 24:15125–15133.
51. Osanloo M., Amani A., Sereshti H., Abai MR., Esmaeili F., Sedaghat MM. Preparation and optimization Nanoemulsion of tarragon (*Artemisia dracunculus*) Essential Oil as effective herbal Larvicidal against *Anopheles stephensi*. *Ind. Crops Prod.* 2017, 109, 214–219. (a)
52. Osanloo M., Sereshti H., Sedaghat MM., Amani A. Nanoemulsion of dill Essential Oil as a green and potent Larvicidal against *Anopheles stephensi*. *Environ. Sci. Pollut. Res.* 2018; 25:6466–6473.
53. Amer A., Mehlhorn H. Larvicidal effects of various Essential Oils against *Aedes*, *Anopheles*, and *Culex* larvae (Diptera, Culicidae). *Parasitol. Res.* 2006; 99:466–472.
54. Sundararajan B., Moola AK., Vivek K., Kumari BDR. Formulation of Nanoemulsion from leaves Essential Oil of *Ocimum basilicum* L. and its antibacterial, antioxidant and Larvicidal activities (*Culex quinquefasciatus*). *Microb. Pathog.* 2018; 125:475–485.
55. Ibrahim J., Zaki ZM., Development of environment-friendly insect repellents from the leaf oils of selected Malaysian plants, *ASEA. Rev. Biodiv. Environ. Conserv.* 1998, 6, 1-7.
56. Jaenson TG., Palsson K., Borg-Karlson AK. Evaluation of extracts and oils of mosquito (Diptera: Culicidae) repellent plants from Sweden and Guinea-Bissau. *J. Med. Entomol.* 2006; 43:113–119.
57. Park BS., Choi WS., Kim JH., Lee SE. Monoterpenes from thyme (*Thymus vulgaris*) as potential mosquito repellents. *J. Am. Mosq. Control Assoc.* 2005; 21:80–83.
58. Yang YC., Lee EH., Lee HS., Lee DK., Ahn YJ. Repellency of aromatic medicinal plant extracts and a steam distillate to *Aedes aegypti*. *J. Am. Mosq. Control Assoc.* 2004; 20:146–149.
59. Gillij YG., Gleiser RM., Zygodlo JA., Mosquito repellent activity of Essential Oils of aromatic plants growing in Argentina. *Bioresour. Technol.* 2008; 99:2507–2515.
60. Kiran S., Devi P. Evaluation of mosquitocidal activity of Essential Oil and sesquiterpenes from leaves of *Chloroxylon swietenia* DC. *Parasitol. Res.* 2007; 101:413–418.
61. Jaenson TG., Palsson K., Borg-Karlson AK. Evaluation of extracts and oils of mosquito (Diptera: Culicidae) repellent plants from Sweden and Guinea-Bissau. *J. Med. Entomol.* 2006; 43:113–119.
62. Sukumar K., Perich MJ., Boobar LR. Botanical derivatives in mosquito control: a review. *J. Am. Mosq. Control Assoc.* 1991; 7:210–237.
63. Odalo JO., Omolo MO., Malebo H., Angira J., Njeru PM., Ndiege IO., Hassanali A. Repellency of Essential Oils of some plants from the Kenyan coast against *Anopheles gambiae*. *Acta Trop.* 2005; 95:210–218.
64. Tunón H., Thorsell W., Mikiver A., Malander I., rthropod repellency, especially tick (*Ixodes ricinus*), exerted by extract from *Artemisia abrotanum* and Essential Oil from flowers of *Dianthus caryophyllum*. *Fitoterapia.* 2006; 77:257–261.
65. Toloza AC., Zygodlo J., Mougabure Cueto G., Biurrun F., Zerba E., Picollo MI. Fumigant and repellent properties of Essential Oils and component compounds against permethrin-resistant *Pediculus humanus capitis* (Anoplura: Pediculidae) from Argentina. *J. Med. Entomol.* 2006; 43:889–895.
66. Omolo MO., Okinyo D., Ndiege IO., Lwande W., Hassanali A. Repellency of Essential Oils of some Kenyan plants against *Anopheles gambiae*. *Phytochemistry.* 2004; 65:2797–2802.
67. Ramar M., Manonmani P., Arumugam P., Kannam SK., Erusan RR., Baskaran N., Murugan K. Nano-insecticidal formulations from Essential Oil (*Ocimum sanctum*) and fabricated in filter paper on adult of *Aedes aegypti* and *Culex quinquefasciatus*. *J. Entomol. Zool. Stud.* 2017; 5:1769–1774.
68. Sakulku U., Nuchuchua O., Uawongyart N., Puttipipatkachorn S., Sootitiantawat A., Ruktanonchai U. Characterization and mosquito repellent activity of citronella oil nanoemulsion. *Int. J. Pharm.* 2009; 372:105–111.
69. Sritabutra D., Soonwera M., Waltanachanobon S., Pongjai S. Evaluation of herbal Essential Oil as repellents against *Aedes aegypti* (L.) and *Anopheles dirus* Peyton & Harrion. *Asian Pac. J. Trop. Biomed.* 2011; 1:S124–S128.
70. Nuchuchua O., Sakulku U., Uawongyart N., Puttipipatkachorn S., Sootitiantawat A., Ruktanonchai U. In vitro characterization and mosquito (*Aedes aegypti*) repellent activity of essential-oils-loaded nanoemulsions. *Aaps Pharmscitech.* 2009; 10:1234.
71. Santos DS., Boito JP., Santos RCV., Quatrin PM., Ourique AF., dos Reis JH., Gebert RR., Glombowsky P., Klauk V., Boligon AA. Nanostructured cinnamon oil has the potential to control *Rhipicephalus microplus* ticks on cattle. *Exp. Appl. Acarol.* 2017; 73:129–138.
72. Monteiro IN., dos Santos Monteiro O., Costa-Junior LM., da Silva Lima A., de Aguiar Andrade EH., Maia JGS., Mouchrek Filho VE. Chemical composition and acaricide activity of an Essential Oil from a rare chemotype of *Cinnamomum verum* Presl on *Rhipicephalus microplus* (Acari: Ixodidae). *Vet. Parasitol.* 2017; 238:54–57.

73. Galli GM., Volpato V., Santos RCV., Gebert RR., Quatrin P., Ourique AF., Klein B., Wagner R., Tonin AA., Baldissera MD. Effects of Essential Oil of *Eucalyptus globulus* loaded in nanoemulsions and in nanocapsules on reproduction of cattle tick (*Rhipicephalus microplus*). Arch. Zootec. 2018; 67:494–498.
74. Baldissera MD., Da Silva AS., Oliveira CB., Zimmermann CEP., Vaucher RA., Santos RCV., Rech VC., Tonin AA., Giongo JL., Mattos CB. Trypanocidal activity of the Essential Oils in their conventional and nanoemulsion forms: In vitro tests. Exp. Parasitol. 2013; 134:356–361.
75. Ziaei Hezarjaribi H., Nadeali N., Saeedi M., Soosaraei M., Jorjani ON., Momeni Z., Fakhar M. The effect of lavender Essential Oil and nanoemulsion on *Trichomonas vaginalis* in vitro. Feyz J. Kashan Univ. Med. Sci. 2017;21:326–334.
76. Shokri A., Saeedi M., Fakhar M., Morteza-Semnani K., Keighobadi M., Teshnizi SH., Kelidari HR., Sadjadi S. Antileishmanial activity of *Lavandula angustifolia* and *Rosmarinus officinalis* Essential Oils and nano-emulsions on Leishmania major (MRHO/IR/75/ER). Iran. J. Parasitol. 2017; 12, 622–631.
77. Bouyahya A., Et-Touys A., Bakri Y., Talbaui A., Fellah H., Abrini J., Dakka N. Chemical composition of *Mentha pulegium* and *Rosmarinus officinalis* Essential Oils and their antileishmanial, antibacterial and antioxidant activities. Microb. Pathog. 2017; 111:41–49.
78. Moazeni M., Borji H., Darbandi MS., Saharkhiz MJ. In vitro and in vivo antihydatid activity of a nano emulsion of *Zataria multiflora* Essential Oil. Res. Vet. Sci. 2017; 114:308–312.
79. Mahmoudvand H., Mirbadie SR., Sadooghian S., Harandi MF., Jahanbakhsh S., Saedi Dezaki E. Chemical composition and scolicidal activity of *Zataria multiflora* Boiss Essential Oil. J. Essent. Oil Res. 2017; 29:42–47.
80. Moazeni M., Larki S., Saharkhiz MJ., Oryan A., Lari MA., Alavi AM. Efficacy of the aromatic water of *Zataria multiflora* on hydatid cysts: An In vivo study. Antimicrob. Agents Chemother. 2014; 58:6003–6008.
81. Elissondo MC., Albani CM., Gende L., Eguaras M., Denegri G. Efficacy of thymol against *Echinococcus granulosus protoscoleces*. Parasitol. Int. 2008; 57:185–190.
82. Yones DA., Taher GA., Ibraheim ZZ. In vitro effects of some herbs used in Egyptian traditional medicine on viability of protoscolices of hydatid cysts. Korean J. Parasitol. 2011; 49:255
83. Elissondo MC., Pensel PE., Denegri GM. Could thymol have effectiveness on scolices and germinal layer of hydatid cysts? Acta Trop. 2013; 125:251–257.
84. Schummer J. Multidisciplinarity, interdisciplinarity, and patterns of research collaboration in nanoscience and nanotechnology. Scientometrics. 2004; 59:425–465.
85. Choi H., Mody CCM. The long history of molecular electronics: Microelectronics origins of nanotechnology. Soc. Stud. Sci. 2009; 39:11–50.
86. Tripathi AK., Upadhyay S., Bhuiyan M., Bhattacharya PR. A review on prospects of Essential Oils as biopesticide in insect-pest management. J. Pharmacogn. Phyther. 2009; 1:52–63.
87. Meira CS., Menezes LRA., dos Santos TB., Macedo TS., Fontes JEN., Costa EV., Pinheiro MLB., da Silva TB., Teixeira Guimarães E., Soares MBP. Chemical composition and antiparasitic activity of Essential Oils from leaves of *Guatteria friesiana* and *Guatteria pogonopus* (Annonaceae). J. Essent. Oil Res. 2017; 29:156–162.
88. Dos Santos Sales V., Monteiro ÁB., de Araújo Delmondes G., do Nascimento EP. Antiparasitic activity and Essential Oil chemical analysis of the *Piper tuberculatum* Jacq fruit. Iran. J. Pharm. Res. IJPR 2018; 17:268–275.

