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### **Review Article**

# NIPAH VIRUS – THREAD TO PREGNANCY

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#### **ABSTRACT:-**

Nipah virus is a type of zoonotic virus. Which cause serious respiratory nervous problems along with serious fever it get easily transmitted to one infected person to another person. Nipah virus is a great thread to mankind. Nipah virus belongs to the family of Paramyxoviridae. Nipah virus positive person is called as NiV positive pigs various birds are also affected by the virus badly. So the study and proper knowledge about Nipah virus should be their.

Key words:- Zoonotic, Virus, Nipah, Inflammation, Health, Pteropusrufus, Transmission, Paramyxoviridae.

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## **INTRODUCTION**

ipah virus is generally known as Zoonotic virus. belongs to the family the family It paramyxoviridae and it is also a member of genus Henipa. Due to its highly morbificity it is classified as (BSL-4) agent and relative new findings. The Centres for Disease Control and Prevention and the National Institute of Allergy and Infectious Diseases have classified Nipah virus as a Category C main concern microorganisms. The name Nipah virus derives from Sungai Nipah (Nipah River Village), where the first separate were acquire<sup>1</sup>. The natural reservoir of the virus has been Materialise from the Bats belonging to genus Pteropus. NiV happens naturally as viruses of fruit bats generally known as 'flying foxes', Nipah virus swept through countless piggeries in Malaysia and killed 1100 people during the period from 1998 through 1999<sup>2</sup> .NiV has consisting of an outer layer of filamentous nucleocapsids, it consists of a single-stranded Ribose nucleic acid which is approx 18.2 kb. The major genome is encodes for six major structures of proteins: nucleocapsid (N), phosphor protein (P), matrix protein (M), fusion protein (F), glycoprotein (G), and large protein or RNA polymerase (L)<sup>3.The</sup> natural basin for NiV is pteropid fruit bats<sup>4</sup>, and instantly bat to human transmission can take place, frequently as a result of expenditure of date tribute sap which contaminated by saliva or urine from infected bats . NiVin Malaysia has been isolated from the spinal fluid and brain of victims<sup>5</sup> and also has been isolated from ecological samples of partially eaten fruits and urine of bats<sup>6</sup>. Madagascar

(Pteropusrufus, Eidolon dupreanum)<sup>7</sup> and Ghana (Eidolon *helvum*)<sup>8</sup> indicating awide geographic distribution of the viruses in which antibodies to Henipa virus is also present. In Cambodia, Thailand or Africa no infection of humans or other species has been observed. In Malaysia and Singapore NiV was recognized as the etiological mediator liable of an outbreak, in pigs and humans. Transmission may be occur by consumption of contaminated or infected food by secretion of bats or contact with spoiled pigs and other way can be human-tohuman mode of transmission . Since 1998 in Bangladesh and India there have been various cases of infections, the lethality rate sometimes excess of 70% in hundreds of cases<sup>9</sup>.Nipah included in first list i.e, the priority list of most emerging diseases of WHO that could be cause a global pandemic, along with various other disesse<sup>10</sup>. In Malaysia, when first case was reported of Nepah virus it was observed that the transmission was due to an amplifying host in which transmission occurred due to close contact with infected domesticated swine<sup>11</sup>. About 1.1 million pigs eventually culling Between September 1998 and April 1999 in Malaysia the peninsular major outbreaks of disease in pigs and humans resulted the death of 105 humans<sup>12</sup>. The diseases in pigs and human infectious and signalised by critical fever with respiratory attachment and sometimes nervous signs in all age classes. Sows and boars occasionally died per critically<sup>13</sup>. The main clinical syndrome in humans was inflammation in brain rather than respiratory with unsympathetic symptoms including fever, headache, myalgia,

drowsiness, and disorientation sometimes proceeding to coma within 48h<sup>14</sup>.The predominance of human occurrences history of direct contact with live pigs. Mostly were the pig farmers. Between 2001 and 2007, It was estimated that Nipah virus 50% of cases of humanto-human transmission in Bangladesh. NiV can be detected by testing sample of human urine, saliva, nasal and oropharyngeal secretions and epidemiological data suggest that direct contact with these secretions of Nipah virus infection. Three potential modes of human-to-human transmission of Nipah virus could be transmission via fomites, direct get in touch with or aerosol<sup>15</sup>.

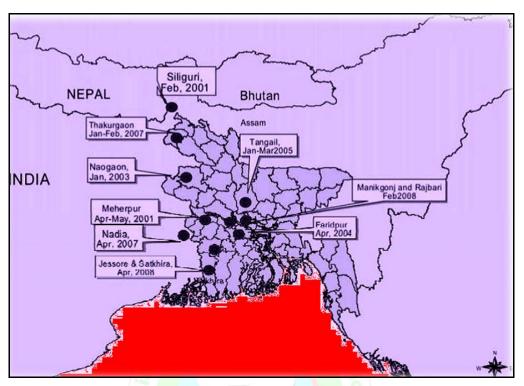


Figure. 1: Structure of Nipah Virus Infection (NiV)

#### WAY OF TRANSMISSION:

NiV disease is an emerging contagious disease spread by secretions of infected bats. It can spread to humans through close contact with infected humans or through eating infected fruit or came into direct contact with infected animals<sup>16</sup>. In Malaysia and Singapore the Nepah virus is spread by direct contact with sick pigs or their infected tissues. This transmission is occurred by respiratory globule, In keeping contact with throat or nasal secretions from the pigs, or keeping contact with the tissue of a sick animal. While in Bangladesh and India, infection is spread by using of fruits or fruit products (such as- date palm or its juice) which is infected with urine or saliva from contagious fruit bats was the most likely source of disease causes organisms<sup>17</sup>. It is reported that Nipah virus spread directly from human-to-human through close contact with people's secretions and excretions during the later outbreaks in Bangladesh and India. The possible perfunctory transmission by rhythmic use of same needles or equipment without further sterilization after each use for health involvement and simulated insemination and sharing of boar semen within a farm were also concerned. The possible role of communication by infected dogs and cats found in the affected farm could not be excepted<sup>18</sup>. Transmission also occurs from direct contact to ruined bats. A common example is using up of raw date palm sap contaminated with transmissible bat excretion,

Serologically positive dogs, cats, bats, horses and goats were found in the infected areas. To date, serum samples from rats attentive in infected area shear all been negative. Testing will also be conducted on blood from cattles (such as goats, sheep, squirrels, wild boar, wild birds, poultry and ostrich)<sup>19</sup>. Experimentally contagious pigs have shown capability to expel NiV as early as 4 days post-infection from the oropharynx, and NiV can also be shack in nasal secretions<sup>20</sup>.Direct contact with infected pigs was recognized as the major mode of conduction in humans when it was first recognized in a large outburst in Malaysia in 1999.90% of the infected people in the 1998-1999 outbreaks were pig farmers or had get in touch with pigs<sup>21</sup>. During the Bangladesh outbreak the virus is recommended to have been transmitted either directly or indirectly from infected bats to humans. Strong proof symptomatic of human- tohuman transmission of NiV was found in Bangladesh in  $2004^{22}$ .

#### **DIFFERENT DIAGNOSIS:**

A variety of laboratory test exists to test for confirmation of NiV virus. It is significant to note that NiV disease has human health implications and all field investigations should take essential safety measures to prevent infection .Presence of any respiratory or neurological disorder in swine in an area known to have *Pteropid* bats, should consider Nipah as a exclude also among swine; deaths of suckling pigs and piglets; Nepah results in unexpected death in boars and sows or abortions and other reproductive dysfunction respiratory diseases with harsh, non-productive coughing and cases with encephalitis (inflammation in brain) manifestations of wobbly, muscular in organization and myoclonus primary to lateral recumbence.

#### Laboratory Diagnosis:

Procedures for the laboratory analysis of NiV contain serology, histopathology, and PCR and virus isolation. Serum Neutralization Test, ELISA, RT-PCR are used for laboratory verification. Most countries in the South-East Asia area do not have sufficient amenities for diagnosing the virus or on ways of overprotective it. Bangladesh, India and Thailand have developed laboratory capability for diagnostic and research purposes. Nipah virus is classified internationally as a bio security level-4 agent and also Bio security level- 2facilities are sufficient if the virus can be first inactivated during sampling collection<sup>23</sup>.

### Virus Isolation:

Virus isolation is an important most important diagnostic move towards for NiV infections. 
□ NiV cultivate well in Vero cells, and the range of specimens elastic isolates in either natural or experimental cases. Brain, lung, kidney and spleen should always be submitted. Tissues are handled under disinfected conditions for preparing of 10% suspensions in cell culture media and are clarified by centrifugation and the supernatant used for immunization of cell cultures. A CPE usually develops within 3 days, but two 5-day passages are suggested before judging the effort ineffective . originally after low diversity infection of cell mono layer, the CPE is manifested by the configuration of syncytia that may contain up to 20 or more nuclei. Consequently syncytia lift from the substrate, leaving interrupt holes in the cell monolayer. The syncytia formed by NiV in Vero cell mono layers are significantly larger than those shaped by HeV in the same time period .□ fascinatingly, the sharing of nuclei differs between NiV-induced syncytia and can be used to distinguish between the two viruses (see Hyatt et al. in this Current focus for more details).Identification methodologies for virus isolates comprise immune staining of fixed, infected cells, neutralization with specific antisera, PCR of culture supernatants, immune electron microscopy and electron microscopy. The later techniques are useful for initial description of the separate since HeV and NiV have separate especially structural features<sup>24</sup>

# **By Electron Microscopy:**

- NiV grows in cultivated cells to titres as high as 108 TCID50 or PFU/ml.
- Apparition of viruses in the intermediate of infected cells by negative contrast electron microscopy and recognition of virus-antibody connections by immune electron microscopy quickly provide important information on virus structure and antigenic reactivity, even during primary separation of the virus.

• Other ultra structural techniques such as grid cell culture, in which cells are grown, infected and visualized on electron microscope grid and recognition of replicating viruses and inclusion bodies in thin sections of fixed, fixed cell cultures and contaminated tissues complement the diagnostic attempt24

### Virus Effective in The Case of Pregnancy:

In utero mother-to- foetus transmission of viruses is of particular interest, because it contains elements of both transmission within an infected host and spread to other hosts. On the one hand, viral spread occurs within the body of the mother through permanent close contact between infected maternal tissue and susceptible foetal tissue. In utero mother –to-mother foetus transmission of viruses can happen within the host and also can be transmitted to other host.

There are two possible ways of transmission to foetus happen, if the pregnant mother is infected with Nipah virus.

- Mother to foetus in-utero transmission
- Transmission during delivery

Placental barrier is a barrier between mother and foetus which allow only highly lipid soluble drugs to pass through it. If there is any complication in between the placental barrier then there is a possibility of transmission to infected maternal tissue. The infection can be occurred due to two reason i.e, by mother to foetus or during delivery of child. In most of the cases, however, only observed pathological changes in the foetus are accepted as evidence for infection of the foetus with utilisation of other approach in utero transmission has been conducted for some viruses, which including cytomegalovirus, varicella-zoster virus, rubella virus, poliovirus, Japanese encephalitis virus, coxsackie virus, echovirus, measles virus, mumps virus, and hepatitis B, C, and E viruses<sup>25</sup>. It is always seen that the infection causes pathological changes in foetus with infection in foetus. In some of the cases the infection in the time of pregnancy leads to more severe condition- example; women infected with hepatitis E virus<sup>26</sup>. This mechanism of in utero transmission of Nipah virus remain same and is also occur by trans cytosisas has been shown for hepatitis B virus<sup>27</sup>. This transfer of virus in pregnant lady was first hypothesized after the discovery that the index case of the 1994<sup>28</sup>.Now, in this issue of the Journal<sup>29-37</sup> report the results of a detailed investigation of in utero NiV transfer& provide the 1<sup>st</sup> experimental evidence that NiV, like HeV, can be vertically transfer in cats. Cats are naturally infected & consistently exhibit characteristic disease pathology even at a low MOI<sup>38</sup>. Isolated sufficient range of infectious NiV from placental fluid (1\_105TCID50/ml) as well as from placental tissue. Evidence was also provide for higher levels of viral replication in several tissues of a pregnant adult cat and in foetal tissues, suggesting both vertical and horizontal transfer of this virus-a finding that has important implications for the epidemiology of NiV infection as well as for the testing of inhibitors & vaccines in this animal model and to understand disease mechanisms. The pathology of cats are common as several features seen in human. Interestingly, temperature increase (as a measure of the development of infection) was initially the same for the pregnant cat and the infected non pregnant control cat during the first 5days; this was followed by a rapid increase for the control cat (figure 1 in Mungall etal.), whereas the infected pregnant cat showed a slight drop in temperature followed by a largely constant period until day 12. Person can hypothesize that the pregnancy can be delayed by this virus affect and thus the disease in the "secondary" infection in the foetus, In which the infection continued unchecked by maternal defence (immune) systems. It can be represents a unique example of protection (although partial in this case) as a result of pregnancy. One can further speculate, as the Mungall et al. did, that such protection could be due to hormonal changes that occur during pregnancy. (The data from this study were based on only two cats , and required to confirm this observation). They also suggesta possible role for cats in HeV and NiVoutbreaks that has never been fully investigated, even though cats were observed at the sites of both HeV and NiV outbreaks<sup>39-45</sup>. Finally, the similarity in major pathological features between this cat modelof disease and human infection could help to develop novel treatments, for example, by identification of maternal pregnancy factors that could delay progression to disease.

#### **Prevention and Control:**

NiV is a viral infectious diseases cause organisms which can transmit through secretion, saliva and by eating a fruits which has been partially eaten by fruits. It also can transmit from human to human. Currently, neither vaccines nor medicines have been proven to be effective in treating NiV infection. However, health care providers may offer supportive therapy to manage symptoms. At this time duration of fever and severity of diseases can be reduce and also may alleviate the symptoms of nausea, vomiting, and convulsions by ribavarin . A recombinant **REFERENCES:** 

- 1. Wang L, Harcourt BH, Yu M, TaminA, Rota PA, Bellini WJ, et al. Molecular biology of Hendra and Nipah viruses. Microbes Infect 2001; 3: 279-287.
- Centers for disease control and prevention. Outbreak of Hendralike virus-Malaysia and Singapore, 1998–1999. MMWR Morb Mortal Wkly Rep 1999; 48: 265-269.
- Broder CC, Xu Kai, Nikolov DB, Zhu Z, Dimitrov DS, Middleton D, Pallister J, Geisbert TW, Bossart KN, Wang LF. A treatment for and vaccine against the deadly Hendra and Nipah viruses. Antiviral Research, 2013;100(1): 8-13.
- 4. Halpin K, Hyatt AD, Fogarty R, Middleton D, Bingham J, Epstein JH, et al. Pteropid bats Are confirmed as the reservoir hosts of henipaviruses: a comprehensive experimental study of virus transmission. Am J Trop Med Hyg 2011; 85: 946-951.
- Chua KB, Bellini WJ, Rota PA, Harcourt BH, et al. Nipah virus: a recently emergent Deadly paramyxovirus. Science 2000; 288:1432-1435.
- Chua KB, Koh CL, Hooi PS, *et al.* Isolation of Nipah virus from Malaysian island flying-foxe. *Microbes Infect*.2002;4(2):145–51.
- 7. Lehlé C, Razafitrimo G, Razainirina J, *et al.* "Henipavirus and Tioman virus antibodies inpteropodid bats, Madagascar". *Emerging Infect. Dis.* 2007;13(1):159–61.
- Hayman D, Suu-Ire R, Breed A, *et al* "Evidence of henipa virus infection in West African fruit bats". *PLoS ONE*. 2008;3(7):2739.
- Hsu VP, Hossain MJ, Parashar UD, Ali MM, Ksiazek TG, Kuzmin I, et al. Nipah virus Encephalitis re-emergence, Bangladesh. Emerg Infect Dis. 2004; 2082-2087.

sub-unit vaccine formulation which protects against lethal NiV challenge in cats. ALVAC Canary poxes vectored Nipah F and G vaccine appears to be a shows potential vaccine for swine and has potential as a vaccine for humans<sup>46-47</sup>. Diseases which may cause by NiV can be prevented by such manners-

- Avoiding direct contact with infected animals,
- Avoid partially eaten fruits and unpasteurised fruit juice,
- Boil freshly collected date palm juice before consuming ,
- Maintain self and child's hygiene,
- Cover household properly,
- Wear NH95- grade and higher masks,
- Regularly wash hand with soap,
- It will be necessary to establishing appropriate surveillance system so that NiV can be detected quickly and appropriate control measures initiated.

#### **CONCLUSION:**

Nipah virus widely affecting pregnant women it may cause serious mortality and morbidity in pregnant women. This virus can be detected by blood sample, serum, urine, tonsil swabs etc. Pregnant women should take protection measures against this virus by protecting her from infected animals (like bat, monkey, cow, pig etc.) Maintain hygiene, avoid eating bitten or fallen fruits or wash fruits thoroughly in salt water. Avoid raw meat of pigs, to travel use N95 mask, avoid drinks made near palm tree etc. for the future outlook antibodies to nipah virus have been found in the fruit bats in India, Indonesia. So the vaccine will be made in future for (NiPV) Nipah virus which will shows potential protections in infected humans.

- Hughes JM, Wilson ME, Luby SP, Gurley ES, Hossain MJ. Transmission of human infectionwith Nipah virus. Clin Infect Dis. 2009;49(11):1743-1748.
- 11. Chua KB, Bellini WJ, Rota PA, Harcourt BH, Tamin A, Lam SK, et al. Nipah virus: a recently emergent deadly paramyxovirus. Science 2000; 288: 1432-1435.
- 12. Nor M.M., Gan C., Ong B., Nipah virus infection of pigs in Peninsular Malaysia, Ministry of Agriculture, Malaysia 1999.
- Nordin B., Ong B., Nipah virus infection in animals and control measures implemented in peninsular Malaysia, Comprehensive Reports on Technical Items presented to the International Committee or Regional Commissions, OIE, Paris, 1999.
- Chua K.B., Goh K.J., Wong K.T., Fatal encephalitis due to Nipah virus among pig-Farmers in Malaysia, lancet. 1999; 354, 1256–1259.
- 15. Devaux P, Hodge G, McChesney MB, Cattaneo R. Attenuation of V- or C defective measles viruses: infection control by the inflammatory and interferon responses of rhesus monkeys. J Virol. 2008;82(11):5359–5367.
- 16. Singh G, Raksha, Urhekar D, Nipah Virus: A killer virus. International journal of advanced microbiology and health research. 2018: 2(2): 40-55.
- Halder A, Chakravarty A. Nipah virus encephalitis: Cause for concern for Indian neurologist. Ann indianAcad Neurol. 2006; 9: 137-144.
- Gupta M, Lo. Mk, Spiropouloy CF. Activation and cell death in human dendritic cells infected with Nipah virus. Virology. 2013; 441(1): 49-56.
- 19. Middleton DJ, Westbury HA, Morrissy CJ, van der Heide BM, Russell GM, Braun MA, Hyatt AD. Experimental Nipah virus infection in pigs and cats. *J Comp Pathol*.2002;126(2):124-136.

- 20. Goh KJ, Tan CT, Chew NK, Tan PSK, Kamarulzaman A, Sarji SA, Wong KT, Abdullah BJJ, Chua KB, Lam SK. Clinical features of Nipah virus encephalitis among pig farmers inMalaysia. N. Engl. J. Med. 2000; 342: 1229– 1235.
- 21. Gurley E, Montgomery JM, Hossain MJ, Bell M, Azad AK, Islam MR, et al. Person- to person transmission of Nipah virus in a Bangladeshi community. Emerg Infect Dis. 2007;13:1031-7.
- Daniels, P., Ksiazek, TG, Eaton, BT. Laboratory diagnosis of Nipah virus and Hendra infections. Microbes Infect. 2001; 3: 289-295.
- Danielsa P, Ksiazekb T, Eatona BT. Laboratory diagnosis of Nipah and Hendrvirus infections. Microbes and Infection, 2001;3:289–295.
- 24. Avgil M, Ornoy A. Herpes simplex virus and Epstein-Barr virus infections in pregnancy: consequences of neonatal or intrauterine infection.ReprodToxicol**2006**; 21:436–45.
- 25. Sookoian S. Liver disease during pregnancy: acute viral hepatitis. Ann Hepatol**2006**; 5: 231–6.
- 26. Bhat P, Anderson DA. Hepatitis B virus translocates across a trophoblastic barrier. J Virol **2007**; 81: 7200–7.
- Halpin K, Young PL, Field HE, Mackenzie JS. Isolation of Hendra virus from pteropid bats: a natural reservoir of Hendra virus. J Gen Virol2000; 81:1927–32.
- Diseases of Swine. 10th ed. Ames, IA: Wiley-Blackwell;
   2012. Wild TF. Henipaviruses: A new Family of emerging Paramyxoviruses. PatholBiol (Paris). 2009;57(2):188-196.
- 29. Eshaghi M, Tan WS, Chin WK, Yusoff K. Purification of the extra-cellular domain of Nipah virus glycoprotein produced in Escherichia coli and possible application in diagnosis. J Biotechnol. 2005; 116(3):221–226.
- 30. Goh KJ, Tan CT, Chew NK, Tan PSK, Kamarulzaman A, Sarji SA, Wong KT, Abdullah BJ, Chua KB, Lam SK. Clinical features of Nipah virus encephalitis among pig farmers in Malaysia. New Engl J Med. 2000; 342:1229– 1235.
- 31. Goldsmith CS, Whistler T, Rollin PE, Chua KB, Bellini W, Rota P, Wong KT, Daszak P, Ksiazek TG, Zaki SR. Ultrastructural studies of Nipah virus, a newly emergent paramyxovirus, using thin section, negative stain, immunogold, and in situ hybridization electron microscopy. MicroscMicroanal. 2000; 6:644–645.
- 32. Harcourt BH, Tamin A, Ksiazek TG, Rollin PE, Anderson LJ, Bellini WJ, Rota PA. Molecular characterization of Nipah virus, a newly emergent paramyxovirus. Virology. 2000; 271:334–349.
- 33. Harcourt BH, Tamin A, Halpin K, Ksiazek TG, Rollin PE, Bellini WJ, Rota PA. Molecular characterization of the polymerase gene and genomic termini of Nipahvirus. Virology. 2001;287:192–201
- 34. Harcourt BH, Lowe L, Tamin A, Liu X, Bankamp B, Bowden N, Rollin PE, Comer JA, Ksiazek TG, Hossain MJ, Gurley ES, Breiman RS, Bellini WJ, Rota PA. Genetic characterization of Nipah viruses isolated during two

outbreaks in Bangladesh in Emerg Infect Dis. 2005; 11:1594–1597.

- 35. Marsh GA, de Jong C, Barr JA, , Tachedjian M, Smith C, Middleton D, Yu M, Todd S, Foord AJ, Haring V, Payne J, Robinson R, Broz I, Crameri G, Field HE, Wang LF. Cedar virus: A novelHenipavirus isolated from Australian bats. PlosPathog. 2012;8: 8.
- 36. Kumar D, Sharma A, Garg R, Formulation and Evaluation of Sublingual Tablet of Losartan Potassium. Asian Journal of Pharmaceutical Research and Development. 2018; 6(4): 54-66.
- 37. Mungall BA, Middleton D, Crameri G, et al. Vertical transmission and fetal replication of Nipah virus in an experimentally infected cat. J Infect Dis 2007; 196:812–6
- 38. Mackenzie JS, Field HE. Emerging encephalitogenic viruses: lyssaviruses and henipaviruses transmitted by frugivorous bats. Arch VirolSuppl2004; 18:97–111.
- 39. Chong HT, Kamarulzaman A, Tan CT, Goh KJ, Thayaparan T, Kunjapan SR, et al. treatment of acute Nipah encephalitis with ribavirin. Ann Neurol. 2001; 49(8):10-3.
- 40. McEachern JA, Bingham J, Crameri G, Green DJ, Hancock TJ, Middleton D, et al. A recombinant subunit vaccine formulation protects against lethal Nipah virus challenge in cats.Vaccine.2008;26(31): 3842-3852.
- 41. Weingartl et al. Recombinant Nipah virus vaccines protect pigs against challenge. Journal of Virology 2006; 80: 7929-38.
- 42. Harit AK, Ichhpujani RL, Gupta S, Gill KS, Lal S, Ganguly NK, Agarwal SP. Nipah/Hendra virus outbreak in Siliguri, West Bengal, India in 2001. Indian J Med Res. 2006; 123:553–560.
- 43. Rahman MA, Hossain MJ, Sultana S, et al. Date Palm Sap Linked to Nipah Virus Outbreak in Bangladesh. VectorBorne and Zoonotic Disease 2012;12(1):65-73.
- 44. Rollin PE, Rota P, Zaki S, Ksiazek TG. Hendra and Nipah viruses. in: Versalovic J, Carroll KC, Funke G, Jorgensen JH, Landry ML, Warnock DW, editors. Manual of Clinical Microbiology. Washington, DC: ASM Press; 2011; 10: 1479-87.
- 45. Wacharapluesadee S, Boongird K, Wanghongsa S, et al. A Longitudinal Study of the Prevalence of Nipah Virus in Pteropuslylei Bats in Thailand: Evidence for Seasonal Preference in Disease Transmission. Vector-Borne and Zoonotic Disease 2010;10(2): 183-90.

46. Breed AC, Meers J, Sendow I, Bossart KN, Barr JA, Smith

- I, Wacharapluesadee S, Wang L, Field HE. The distribution of henipaviruses in Southeast Asia and Australasia: is Wallace's line a barrier to Nipah virus? PLoS One. 2013; 8(4): 61316.
- 47. Chowdhury S, Khan SU, Crameri G, Epstein JH, Broder CC, Islam A, Peel AJ, Barr J, Daszak P, Wang LF, Luby SP. Serological evidence of henipavirus exposure in cattle, goats and pigs in Bangladesh. PLoSNegl Trop Dis. 2014; 8(11): 3302.