



## Research Article

## ANTIVIRAL, ANTIPROTOZOAL, ANTIMALARIAL AND INSECTICIDAL ACTIVITIES OF *OCIMUM GRATISSIMUM L.*

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### ABSTRACT

*Ocimum gratissimum* has been used in traditional medicine for curing various ailments in tropical countries. The plant with unique bioactive compounds possesses a significant medicinal value. Among numerous curative properties the plant shows strong inhibition against human viruses and parasitic protozoans. The presence of eugenol makes it a safe mosquito repellent and anti-malarial agent. Moreover, the plant causes mortality of agricultural insects in both store seed and field crops. The researches have also proved the inhibitory action of plants against various plants and human nematodes harboring alimentary canal. Thus, there is a need to explore the potentiality of this plant to discover biological drug formulation with relevant action against plant and animal microbes, malaria parasites and nematodes. This systematic review presents the antiviral, antiprotozoal, antimalarial, insecticides and nematocidal activities of the plant for developing a standard therapeutic system.

**Keywords:** *Ocimum gratissimum*, Antiviral, Antiprotozoal, Antimalarial, Insecticidal, Nematocidal

### INTRODUCTION

The use of traditional medicine in health care and disease control is in practice in developing countries and is expanding rapidly in various other countries.<sup>[1]</sup> These countries are using medicinal plants for the treatment of multiple diseases, including cancer, infectious and parasitic diseases. Medicinal plants, mainly contain bioactive compounds in the form of secondary metabolite<sup>[2]</sup> which performs an active role to protect plants against microorganism infections and predator insects and can help in developing new biological products to combat plant and animal diseases.<sup>[3,4]</sup> The use of plant extracts as antiviral<sup>[5-7]</sup> antiprotozoal<sup>[8-11]</sup> and anthelmintic agents, have also been justified through numerous studies.

Today, most of the existing anti malarial drugs are taken from plant products.<sup>[2]</sup> These are readily obtainable, cheaper, biodegradable, non-toxic, and possess a broad-spectrum target-specific action against vector mosquitoes.<sup>[12]</sup> In recent years, various researches have been conducted on the traditional medicine of plant origin to evaluate their therapeutic potentials.<sup>[13-15]</sup>

*Ocimum gratissimum* an important tropical plant has been used in traditional medicine since ancient times.<sup>[16,17]</sup> The plant, belonging to family Lamiaceae commonly known as lemon basil or clove basil, is an erect aromatic shrub, polymorphic, branched perennial 50-200cm long<sup>[18]</sup> with opposite, ovate-lanceolate leaves, having dotted glands on the lower surface. The inflorescence is sparingly branched with dense calyx on the outer region. The mouth being held in private by a lower lip with small concealed white corolla (Fig. 1). The plant is distributed in Tropical Africa; Eastern Asia – Indonesia,

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Malaysia, Vietnam, Cambodia, Laos, Thailand, India, Bangladesh, Sri Lanka and Nepal. The plant generally occurs wild along disturbed land and lake shores<sup>[19]</sup> and is also cultivated in various countries.<sup>[20]</sup> The plant

produces essential oil and used in culinary and medicinal purposes like curing fever, antiseptic, antispasmodic, insect repellent, killing internal parasites etc.<sup>[20-22]</sup> There are numerous studies that have been carried with



**Fig. 1: *Ocimum gratissimum***

*Ocimum gratissimum* proving its antimicrobial, insect repellent, antimalarial, insecticides and pesticide activities.<sup>[23-29]</sup> This comprehensive review aims to focus the main antiviral, antiprotozoal, antimalarial and nematicide properties of the plant for future studies as an alternative medicine.

#### **Antiviral activities**

The literature on antiviral activities of clove basil oil reveals potent activities against virus pathogens causing human diseases. There are various literatures that provides the potentiality of Eugenol, a principal constituent of clove basil plant, as an antiviral agent.<sup>[30-32]</sup> Some study suggests the antiviral activities of plant oil with commercial importance and traditional uses.<sup>[33]</sup> Nevertheless the further study emphasizes in carrying more researches regarding the use of clove basil oil in encountering the lethal and highly infectious human viruses.<sup>[34]</sup> There are various studies that justified the antiviral action of the plant on human lethal viruses such as cytomegalovirus (CMV), murine CMV (MCMV)<sup>[6]</sup>, hepatitis C virus<sup>[35]</sup> herpes simplex-1 (HSV-1) and herpes simplex -2 (HSV-2) viruses.<sup>[26,36]</sup> In vitro studies of the plant leaf extract exhibits potent inhibition of HIV-1 and HIV-2 replication with antiviral indices value 110. Further, it also expresses cytotoxicity inhibiting reverse transcriptase and proviral

DNA copying of HIV-1, justifying the antiviral potential of the plant.<sup>[6]</sup> Moreover the plant also possesses effective curative properties for treating measles in children resulted from Paramyxovirus<sup>[37]</sup>. Depending on research, there should be a special concern on investigations that encourage the use of herbal essential oil that is economical with less side effects showing significant inhibitory activity against human viruses.<sup>[7]</sup>

#### **Antiprotozoal activities**

There are numerous studies on the efficacy of clove basil oil as an antiprotozoal agent. The use of plant volatile oil in traditional medicine as a protozoan controlling agent is given in the literature<sup>[8]</sup>. In the last three-four decades plenty of work has been done to analyze the clove basil oil in controlling leishmaniasis, a human cutaneous disease caused by Sand fly and other harmful protozoan parasites. Some study reports that clove basil oil inhibits the growth and modifies ultrastructure of mitochondria of *Herpetomonas samuelpessoai*, a non pathogenic trypanosomatid protozoan.<sup>[38]</sup> The in vivo test of plant oil in concentration between 200-500mg/kg in mice, against the growth of *Plasmodium berghei* a parasitic protozoa causing malaria, exhibit a strong percentage of inhibition ranging from 55-77.8%<sup>[39]</sup> whereas some other observation reported moderate action

of plant leaf extract against *Plasmodium falciparum*.<sup>[40]</sup>

An investigation using transmission electron microscopy on peritoneal macrophages of mouse observed that plant oil with chief constituent as eugenol when treated with promastigotes and amastigotes of *Leishmania amazonensis*, alters cell division and modify mitochondrial structure, minimizes association of promastigotes and macrophages, and increases production of nitric oxide by infected macrophages without any cytotoxicity.<sup>[24]</sup> According to some study methanolic extracts of clove basil oil was found effective against promastigotes *Leishmania chagasi*<sup>[41,42]</sup> and *L. amazonensis*, proving antileishmanial activity for treating several infectious and inflammatory diseases.<sup>[41]</sup> The steam distilled concentration dependent assay of essential oil expressed an effective control of *L. chagasi* promastigotes with IC<sub>50</sub> values 75µg/ml, and modifies the whole viability, including cell structure, lipid accumulation in the cytoplasm and rise in acidocalcisome of the protozoan.<sup>[43]</sup> The minor compounds of the plant oil such as myrcene with IC<sub>50</sub> value 2.24 ± 0.27µg/mL, limonene having IC<sub>50</sub> value 4.24 ± 2.27µg/mL, citronellal with IC<sub>50</sub> value 2.76 ± 1.55µg/mL, and ethanol crude extracts of the plant collected in full blossoming stages with leaves having IC<sub>50</sub> value 1.66 ± 0.48 µg/ml and seeds selected with IC<sub>50</sub> value 1.29 ± 0.42 µg/ml shows effective inhibition of *Trypanosoma brucei*, which causes vector-borne disease in vertebrates, including humans. Further, the plant oil and non-volatile extracts also show inhibition of *Plasmodium falciparum* to some extent.<sup>[26]</sup>

There are various studies that record the credentials of the use of *Ocimum* oil with eugenol as a chief ingredient with antiprotozoal activities.<sup>[9,10,44]</sup> In past four-five years, various studies, including the one in which the plant oil tested against protozoans *Leishmania* sp. and *Trypanosoma cruzi* a causal organism of leishmaniasis and trypanosomiasis, shows significant inhibition against the protozoans.<sup>[45]</sup> An in vitro investigation regarding efficacy of clove basil oil along with five other essential oils obtained through hydro distillation at a concentration necessary to suppress 50% of amastigotes growth IC (50) and trypanomastigote

forms LC(50) of *T. cruzi*, causal organism of chagas disease, shows a significant inhibition of the parasite mainly due to the presence of chemical substances monoterpenes and sesquiterpenes in all the products.<sup>[46]</sup> A study in Cameroon contends that Nefang a polyherbal anti-malarial folklore medicine with *O. gratissimum* leaves as an important constituent, exhibit an excellent action against multi-drug resistant *Plasmodium falciparum* along with in vivo suppression of *P. berghei* and *P. chabaudi* malaria parasites.<sup>[47]</sup> Accordingly, when the evaluation of ten plant species against bloodstream strains of *Trypanosoma brucei rhodesiense* was carried out, it was noted that the ethyl acetate leaf extract of *O. gratissimum* expressed highest activity IC(50) value of 2.08 0.01 µg/ml and a high selective index of 29 against the protozoan.<sup>[48]</sup> Moreover, a recent study on use of lemon basil oil against *L. amazonensis* also provided evidence of the leishmanicidal activity of the plant.<sup>[11]</sup>

#### Antimalarial activities

Malaria brought about by the bites of mosquitoes is a life-threatening disease, accounting for 90% of global cases in the African region, 7% in South-East Asia and 2% in Eastern Mediterranean Region.<sup>[49]</sup> Although there is a rapid development in active ingredients or new insecticides, but the pesticide increasing resistance in parasites is a matter of grave concern. In these circumstances, the bio-products can ensure non-toxicity to meet the environmental safety standards and public health. There are numerous works on the use of lemon basil as mosquito repellent and insect mortality. The synergistic formulation of essential oil of the leaves and peels of *Ocimum gratissimum* with *Eucalyptus globulus*, *Cymbopogon citratus*, *Azadirachta indica*, *Citrus sinensis* and *Hyptis suaveolens* expresses strong repellent activities and can be used as an effective bioproduct.<sup>[50]</sup> Indigenous communities of Massissip region in Ngumba Cameroon apply smashed plant leaves on the skin as a mosquito-repellent.<sup>[51]</sup> The lotion prepared by volatile oil with olive oil base exhibited highest repellencies whereas palm kernel and olive oil bases shows mosquito bite protection and expressed fast knockdown and paralyzing effect on the few mosquitoes expressing mosquito-repellent and

mosquitocidal potentials on human.<sup>[52]</sup> *O. gratissimum* petroleum ether extracts at 50% concentration causes 100% mortality of the larvae of *Culex* mosquito species.<sup>[53]</sup> Some study observed methanol<sup>[54]</sup>, whereas other reported n-hexane extract of the plant with an effective larvicide activity against *Culex quinquefasciatus* credited to presence of caryophyllene oxide, along with ethyl acetate and methanol leaf extract being effective against *Culex gelidus* larval and adult mosquitoes.<sup>[55]</sup> According to a study plant powder, oil and wax candle formulation of *O. gratissimum* results in 100% mortality of adult *Anopheles gambiae* mosquito.<sup>[56]</sup> The leaf essential oil possesses larvicide activity against *Aedes albopictus* which is credited due to chemical compounds 3-allyl-6-methoxyphenol, 4-(5-ethenyl-1-azabicyclo (2, 2, 2) octan-2), 1-(2, 5-dimethoxyphenyl) -propanol and 1-(1-hydroxybutyl) -2, 5-dimethoxybenzene in the concentration of 19.30 %, 16.82 %, 12.23 %, and 5.53 % respectively.<sup>[57]</sup> The mixture of *Lantana camara* leaves and *O. gratissimum* with ethyl acetate followed by methanol mixture of these plants shows synergistic effect<sup>[58]</sup> and in another experiment the combination of hexane fraction of the two plants and mixture of ethyl acetate (*O. gratissimum*) and hexane (*L. camara*) yielded synergistic actions. However, the same author also reported that the combination of ethyl acetate and hexane (*O. gratissimum*) and ethyl acetate (*L. camara*) shows antagonistic effects against *Aedes aegypti* mosquito.<sup>[59]</sup> Various studies observed that crude essential oils<sup>[3,60]</sup> and essential oils with liquid paraffin base, exhibits larval mortality or even repellent action against *Aedes aegypti* mosquitoes.<sup>[61]</sup> Further, this concept got strengthened by a study that demonstrated that n-hexane leaf extract contains terpenes, saponins, tannins, steroids and phenol possessing high larvicide mortality against mosquito larvae.<sup>[62]</sup> Another study suggests that the chloroform extract also has pupicidal and adulticidal activity against *Aedes aegypti* mosquito, which is credited to the presence of phenolic compounds Hentriaconate, tetracosahexane, Hepta 2-1 trimethyl, Benzopyran, Hexamethyl and Dihydro tetramethyl trimethyl acetate.<sup>[63]</sup>

## Insecticidal activity

*O. gratissimum* exhibit an effective control of larval and adult insects as evidenced in various in-vitro and field experiments. The plant can be used as a good natural protection agent for food commodities and its oil followed by constituents eugenol and  $\beta$ -(Z) -ocimene shows fumigant toxicity and repellent action against agricultural insects, *Tribolium castaneum* (Herbst), *Sitophilus oryzae* (L.), *Oryzaephilus surinamensis* (L.), *Callosobruchus chinensis* (L.) and *Rhyzopertha dominica* (F.).<sup>[64]</sup> According to a report, 1% of essential oils diluted with acetone cause 98% mortality of *Sitophilus zeamais* Motsch insect, after 24 h of treatment.<sup>[23]</sup> The other studies also reported that time and duration of the application of plant oil play a decisive role in inhibiting the insects and causes mortality of adult weevils up to 74 % after 4 days<sup>[65]</sup> and further the mortality decreases of 50% after 8 days.<sup>[66]</sup> The formulations based on plant oil and modified clays can act as a good bio-insecticide which was experimentally proved in a study which reports the strong affinity of essential oil with modified Cameroonian montmorillonite clay that persists even up to 107 days effecting maize weevil *S. zeamais*.<sup>[67]</sup> There are investigation that suggests that 25  $\mu$ l/vial of *O. gratissimum* causes 80% mortality of adult cowpea seed beetle *Callosobruchus maculate* after 12 hours of treatment and when 1 g of plant powder is used, 50% lethal concentration was noted to be 116  $\mu$ l/g of the oil which compared more sensitive to the male than to female adults after 48 hours. Further the use of essential oil at the concentration of 30  $\mu$ l reduces egg hatching rate up to 15%. The plant oil at the rate of 400  $\mu$ l also provided complete protection from the beetle even for more than 3 months duration under storage.<sup>[68]</sup> A study observed that a mixture of *Vernonia amygdalina* and *Ocimum gratissimum* in the ratio of 50%: 50% applied at the rate of 5% per 30 g of cowpea seeds, causes 33-80% mortality of *C. maculatus*, reduces egg count and impose least the adult emergence of the beetle at 24, 48 and 72 hours after treatment.<sup>[69]</sup>

**Table 1: Antimicrobial activities of *O. gratissimum* against Virus, Protozoa, Malaria Vector, Insects and Nematodes**

Target organisms	References
<b>Virus</b>	
i. Cytomegalovirus (CMV)	[5]
ii. Murine cytomegalovirus (MCMV)	[5]
iii. HIV-1	[6, 32]
iv. HIV-2	[6]
v. Herpes simplex-1(HSV-1)	[7,25,37]
vi. Herpes simplex-2 (HSV-2)	[7,37]
vii. Hepatitis C virus	[36]
viii. Paramyxovirus	[38]
<b>Protozoa</b>	
i. <i>Leishmania amazonensis</i>	[10,24,42]
ii. <i>Trypanosoma brucei brucei</i>	[26]
iii. <i>Plasmodium falciparum</i>	[26,41,48]
iv. <i>Herpetomonas samuelpessoai</i>	[39]
v. <i>Plasmodium berghei</i>	[40,48]
vi. <i>Plasmodium chabaudi</i>	[48]
vii. <i>Leishmaniachagasi</i>	[42-44]
viii. <i>Trypanosoma cruzi</i>	[46,47]
ix. <i>Trypanosoma brucei rhodesiense</i>	[49]
<b>Malaria Vector</b>	
i. <i>Culex quinquefasciatus</i>	[54-56]
ii. <i>Culex gelidus</i>	[56]
iii. <i>Anopheles gambiae</i>	[57]
iv. <i>Aedes albopictus</i>	[58]
v. <i>Aedes aegypti</i>	[3,60-64]
<b>Insects</b>	
<b>Agriculture insect</b>	
i. <i>Tribolium castaneum</i> (Herbst)	[65]
ii. <i>Sitophilus oryzae</i> (L.)	[65]
iii. <i>Oryzaephilus surinamensis</i> (L.)	[65]
iv. <i>Callosobruchus chinensis</i> (L.)	[65]
v. <i>Rhyzopertha dominica</i> (F.)	[65]
vi. <i>Sitophilus zeamais</i>	[66-68]
vii. <i>Callosobruchus maculatus</i>	[69,70]
viii. <i>Acanthscelides obtectus</i>	[71]
ix. <i>Plutella xylostella</i>	[72]
x. <i>Brevicoryne brassicae</i>	[72]
xi. <i>Aphis craccivora</i>	[73,74]
<b>Flies</b>	
i. <i>Musca domestica</i>	[75]
ii. <i>Simulium damnosum</i>	[76,77]
<b>Nematodes</b>	
i. <i>Haemonchus contortus</i>	[78]
ii. <i>Meloidogyne incognita</i>	[27,80,82]

According to a report 4% solution of the ethanolic leaf extracts causes 28.80 % mortality of beans weevil *Acanthscelides obtectus* 1.50 hours after treatment and rate of toxicity increases with the concentration and time of the exposures.<sup>[70]</sup> The detergent and water extracts of plant shows significant inhibition of cabbage diamondback moth, *Plutella xylostella*, and *Brevicoryne brassicae* aphid and is comparable to synthetic insecticide.<sup>[71]</sup> Some study found that hot solvent extraction of aerial parts of the plant and root causes of 73.33 and 86.66% mortality of groundnut aphid *Aphis craccivora* Koch.<sup>[72]</sup> The acetone extract of the plant also shows an influential activity against cowpea aphid, *Aphis craccivora* causing mortality, life span reduction and decrease in reproduction.<sup>[73]</sup> Plant oil diluted in 2% acetone expresses 100% repellency against housefly *Musca domestica* L. and is comparable to pesticides dimethyl phthalate, N, N-diethyl-meta-toluamide, malathion, thymol, pyrethrum extract, piperonylbutoxide and mineral oil.<sup>[74]</sup> The plant oil with liquid paraffin base shows a strong repellent against *Simulium damnosum* blackflies.<sup>[75]</sup> An investigation reveals that plant oil at 20% concentration with liquid paraffin as the carrier applied on forearms and legs gave protection up to three hours against blackfly, *S. damnosum* s.l., the insect vector of human onchocerciasis.<sup>[76]</sup>

### Nematicidal activities

There are numerous studies on inhibitory properties of lemon basil extracts against helminths and their herbal use. A study reports that *O. gratissimum* oil 0.50% concentration shows ovicidal activity against *Haemonchus contortus* a parasite present in gastrointestinal of small ruminants<sup>[77]</sup> and the eugenol isolated from the plant also possess nematicidal properties.<sup>[77,78]</sup> The aqueous extract of the plant at the rate of 20,000 mg/kg increases the yield and reduces *Meloidogyne incognita* pathogenicity in cowpea by inhibiting egg hatching 40% - 63.7% and juvenile mortality ranging from 82% - 93.8% of the root-knot nematode<sup>[79]</sup>, whereas water extract at the concentration of 20 ml exhibited nematicidal activity against

*M. incognita* and found comparable with carbofuran by increasing fruit production, controlling root damage and the growth of okra.<sup>[27]</sup> The powdered extract of *Azadirachta indica* and *O. gratissimum* possess nematicidal properties and when used in combination shows synergistic action against root knot nematode of a pepper plant as compared to be used individually and the soil treated with these plants has potentiality to kill other plant parasitic nematodes.<sup>[80]</sup> In vitro study suggest that plant essential oil at zero percent concentrations affects the survival of the eggs of *M. incognita* root nematodes of tomato in 48 hours and at higher concentration inhibits hatching or causes mortality.<sup>[81]</sup>

A study reports that more than 1000 ppm concentrations of the plant cause 50% mortality of brine shrimp nauplii and were significantly comparable to piperazine drug. The plant extract also shows toxicity against parasitic *Haemonchus infective* L3 stage nematode. The in vitro study further reveals the inhibitory action of plant with IC50s of 8 and 10 micro/ml against recombinant *Onchocerca* and *Ascaris* GSTs (glutathione-S-transferases).<sup>[82]</sup> The methanolic leaf extract based on dose and duration shows haematinic and haemopoietic activities which were justified by an experiment inducing phenylhydrazine anaemia in Albino wistar rats. The result reveals a significant decrease in the mean value of the red blood cell (RBC), an increase in the RBC indices, a slight increase in the mean values of white blood cell (WBC) and increase in lymphocytes.<sup>[83]</sup>

### CONCLUSION

There are various scientific studies on the uses of lemon basil as antimicrobial, antimalarial, nematicidal and insecticides agent. As most of the mentioned researches have been conducted using a crude preparation, few of them use ethanol or acetone base, but the chemical studies are less mentioned, thus requiring a need of controlled clinical trial. The plant potential in inhibiting the HIV virus, mosquito repellent and larvicidal activities makes it a novel bio-

product. The use of the plant and their essential oils in the form of insecticides is safer than synthetic pesticides. Most importantly, it is economical and easily available in tropical countries this special focus should be placed on the discovery of new drug formulations using these natural products to combat the microorganisms and insects.

## REFERENCES

1. WHO traditional medicine strategy: 2014-2023: ISBN 978 92 4 150609 0;2013.
2. Kaur R, Kaur S. Evaluation of in vitro and in vivo antileishmanial potential of bergenin rich *Bergenia ligulata* (Wall.) Engl. root extract against visceral leishmaniasis in inbred BALB/c mice through immunomodulation. *J Trad Compl Med* 2017; <https://doi.org/10.1016/j.jtcm.2017.06.006> (In Press, Corrected Proof)
3. Cavalcanti ESB, Morais SM, Lima MAA, Santana EWP. Larvicidal Activity of essential oils from Brazilian plants against *Aedes aegypti* L. *Mem. Inst. Oswaldo Cruz* [online] 2004;99:541-44.
4. Pandey S, Shukla A, Pandey S, Pandey A. Morphology, chemical composition and therapeutic potential of *Somlata* (*Sarcostemma acidum* Wight. & Arn.). *Pharma Sci Moni* 2017; 8:54-60.
5. Yukawa et al. Prophylactic treatment of cytomegalovirus infection with traditional herbs. *Antiviral Res* 1996;32:63-70.
6. Ayisi NK, Nyadedzor C. Comparative in vitro effects of AZT and extracts of *Ocimum gratissimum*, *Ficus polita*, *Clausena anisata*, *Alchornea cordifolia*, and *Elaeophorbiadrupifera* against HIV-1 and HIV-2 infections. *Antiviral Res* 2003;58:25-33.
7. Gavanji S, Sayedipour SS, Larki B, Bakhtari A. Antiviral activity of some plant oils against herpes simplex virus type 1 in Vero cell culture. *J Acute Med* 2015;5:62-68.
8. Holetz et al. Effect of plant extracts used in folk medicine on cell growth and differentiation of *Herpetomonas samuelpessoai* (Kinetoplastida, Trypanosomatidae) cultivated in defined medium. *Acta Scientiarum* 2002;24(3):657-62.
9. Monzote L, Alarcon O, Setzer WN. Antiprotozoal activity of essential oils. *Agriculturae Conspectus Scientifi cus* 2012;77:167-75.
10. Perez GS, Ramos-López MA, Sanchez-Miranda E, Fresán-Orozco MC, Perez-Ramos J. Antiprotozoa activity of some essential oils. *J Med Pl Res* 2012;6:2901-08.
11. Silva et al. Leishmanicidal Activity and structure-activity relationships of essential oil constituents. *Molecules* 2017;22:815.
12. Ghosh A, Chowdhury N, Chandra G. Plant extracts as potential mosquito larvicides. *Ind J Med Res* 2012;135:581-98.
13. Kumar S, Pandey S. An ethnobotanical study of local plants and their medicinal importance in Tons river area, Dehradun, Uttarakhand. *Indian J Trop Biodiv* 2015; 23: 227-31.
14. Pandey S, Shukla A, Pandey S, Pandey A. An overview of resurrecting herb 'Sanjeevani' (*Selaginella bryopteris*) and its pharmacological and ethnomedicinal uses. *The Pharm Innov* 2017; 6:11-14.
15. Grover JK, Yadav SP. Pharmacological actions and potential uses of *Momordica charantia*: a review. *J Ethnopharma* 2004; 93:123-32.
16. Prabhu KS, Lobo R, Shirwaikar AA, Shirwaikar A. *Ocimum gratissimum*: A review of its chemical, pharmacological and ethnomedicinal properties. *Open ComplemMed J* 2009;1:1-15.
17. Omosun G, Okoro IA, Ekundayo E, Ojmelukwe PC, Ibe O. Ethnobotanical study of medicinal plants useful for malaria therapy in eight local government areas of Abia State, Southeast Nigeria. *Adv Med Plant Res* 2013;1:39.
18. Savithramma N, Yugandhar P, Babu RH, Prasad KS. Validation of indigenous knowledge of Yanadi tribe and local villagers of Veyilingalakona- A sacred grove of Andhra Pradesh. *Ind J Pharm Sci Res* 2014;6:382.
19. <http://www.worldagroforestry.org>.
20. Achigan-Dako et al. Traditional Vegetables in Benin. Darwin Initiative; International Foundation for Science 2009.
21. Bown D: *Encyclopaedia of Herbs and their Uses*. ISBN 0-7513-020-31: Dorling Kindersley, London; 1995.
22. DeFilipps RA, Maina SL, Crepin J. *Medicinal Plants of the Guianas*: Smithsonian museum; 2004.
23. Kouninki et al. Potential use of essential oils from Cameroon applied as fumigant or contact insecticides against *Sitophilus zeamais* Motsch. (Coleoptera: Curculionidae). *Commun Agric Appl Biol Sci* 2005;70:787-92.
24. Ueda-Nakamura et al. Antileishmanial activity of eugenol-rich essential oil from *Ocimum gratissimum*. *Parasitol Int* 2006;55:99-105.
25. Astani A, Reichling J, Schnitzler P. Screening for antiviral activities of isolated compounds from essential oils. *Evid Based Complement Alternat Med* 2011; 253643
26. Kpadonou Kpoviessi et al. In vitro antitrypanosomal and antiplasmodial activities of crude extracts and essential oils of *Ocimum gratissimum* Linn from Benin and influence of vegetative stage. *J. Ethnopharmacol* 2014;155:1417-23.
27. Asimiea AO, Tanimola AA, Osaronwi MO. Nematicidal effect of aqueous extract of *Ocimum gratissimum* (L.) leaves on *Meloidogyne incognita* on Okra. *J Appl Sci Agric* 2015;10:22-28.
28. Pandey S. Antibacterial and antifungal activities of *Ocimum gratissimum* L. *Int J Pharm Pharma Sci* 2017;9(In press).
29. Iqbal J, Mishra RP. In vitro activity of medicinal plants against some bacterial and fungal isolates. *Asian J Pharm Clin Res* 2015;8:225-30.
30. Garg A, Singh S. Eugenol: A potential phytochemical with multifaceted therapeutic activities. *Pharmacology* 2010;2:108-20.
31. Behbahani M, Mohabatkar H, Soltani M. Anti-HIV-1 activities of aerial parts of *Ocimum basilicum* and its Parasite *Cuscuta campestris*. *J Antivir Antiretrovir* 2013;5:57-61.
32. Raja MRC, Srinivasan V, Selvaraj S, Mahapatra SK. Versatile and synergistic potential of eugenol: A Review. *Pharm Anal Acta* 2015;6:367.
33. Kalita J, Khan ML. Commercial potentialities of essential oil of *Ocimum* members growing in North East India. *Int J Pharm Life Sci* 2013;4:2559-67.
34. Pavithra B. Eugenol-A Review. *J PharmSci Res* 2014;6:153-54.

35. Hussein et al. Inhibitory effects of sudanese medicinal plant extracts on hepatitis C virus (HCV) protease. *Phytother Res* 2000;14:510-16.
36. Benencia F, Courreges MC. In vitro and in vivo activity of eugenol on human herpesvirus. *Phytother Res* 2000;14:495-500.
37. Sonibare MA, Moody JO, Adesanya EO. Use of medicinal plants for the treatment of measles in Nigeria. *J Ethnopharmacol* 2009;122:268-72.
38. Holetz et al. Effect of Essential Oil of *Ocimum gratissimum* on the Trypanosomatid *Herpetomonas samuelpessoai*. *Acta Protozool* 2003;42:269-76.
39. Tchoumboungang F, Zollo PH, Dagne E, Mekonnen Y. In vivo antimalarial activity of essential oils from *Cymbopogon citratus* and *Ocimum gratissimum* on mice infected with *Plasmodium berghei*. *Planta Med* 2005;71:20-3.
40. Kamaraj et al. Antiplasmodial potential of medicinal plant extracts from Malaiyur and Javadhu hills of South India. *Parasitol Res* 2012;111:703-15.
41. Obot MJ, Aluyi HAS. Treatment of superficial mycoses with *Ocimum gratissimum*. *Int J Infect Dis* 2002;6:151.
42. Braga et al. Antileishmanial and antifungal activity of plants used in traditional medicine in Brazil. *J Ethnopharma* 2007;111:396-402.
43. Oliveira et al. Effects of essential oils from *Cymbopogon citratus* (DC) Stapf., *Lippia sidoides* Cham., and *Ocimum gratissimum* L. on growth and ultrastructure of *Leishmania chagasi* promastigotes. *Parasitol Res* 2009;104:1053-59.
44. Musuyu et al. In vitro antiprotozoal and cytotoxic activity of 33 ethnopharmacologically selected medicinal plants from Democratic Republic of Congo. *J Ethnopharma* 2012;141:301-08.
45. Alviano et al. Conventional therapy and promising plant-derived compounds against trypanosomatid parasites. *Front Microbiol* 2012;https://doi.org/10.3389/fmicb.2012.00283.
46. Borges et al. Trypanocidal and cytotoxic activities of essential oils from medicinal plants of Northeast of Brazil. *Exp Parasitol* 2012;132:123-28.
47. Arrey Tarkang P, Okalebo FA, Ayong LS, Agbor GA, Guantai AN. Anti-malarial activity of a polyherbal product (Nefang) during early and established *Plasmodium* infection in rodent models. *Malaria J* 2014;13:456.
48. Abiodun OO, Gbotosho GO, Ajaiyeoba EO, Brun R, Oduola AM. Antitrypanosomal activity of some medicinal plants from Nigerian ethnomedicine. *Parasitol Res* 2012;110:521-26.
49. World malaria report 2016, WHO.
50. Lawal HO, Adewuyi GO, Fawehinmi AB, Etatuvi SO. Chemical evaluation of mosquito repellent formulation prepared from the essential oil of plants. *J Nat Pro* 2013;6:33-37.
51. Youmsi et al. Ethnobotanical survey of medicinal plants used as insects repellents in six malaria endemic localities of Cameroon. *J Ethnobiol Ethnomed* 2017;13:33.
52. Oparaocha ET, Iwub I, Ahanakuc JE. Preliminary study on mosquito repellent and mosquitocidal activities of *Ocimum gratissimum* (L.) grown in eastern Nigeria. *J Vector Borne Dis* 2010;47:45-50.
53. Okigbo RN, Okeke JJ, Madu NC. Larvicidal effects of *Azadirachta indica*, *Ocimum gratissimum* and *Hyptis suaveolens* against mosquito larvae. *J AgriTech* 2010;6:703-19.
54. Kamaraj C, Rahuman AA. Larvicidal and adulticidal potential of medicinal plant extracts from south India against vectors. *Asian Pacific J Trop Med* 2010;3:948-53.
55. Nzalibe HC, Chintem DGW. Larvicidal potential of leaf extracts and purified fraction *Ocimum Gratissimum* against *Culex Quinquifasciatus* mosquito larva. *Int J Sci Res* 2015;4:2254-58.
56. Afolabi O. J. Efficacy of *Ocimum gratissimum* on adult *Anopheles gambiae*. *J Mosquito Res* 2016;6:1-6.
57. Sumitha KV, Thoppil JE. Larvicidal efficacy and chemical constituents of *O. gratissimum* L. (Lamiaceae) essential oil against *Aedes albopictus* Skuse (Diptera: Culicidae). *Parasitol Res* 2016;115:673-80.
58. Keziah EA, Nukene EN, Danga SPY, Younoussa L, Esimone CO. Creams formulated with *Ocimum gratissimum* L. and *Lantana camara* L. crude extracts and fractions as mosquito repellents against *Aedes aegypti* L. (Diptera: Culicidae). *J Ins Sci* 2015;15:45.
59. Keziah EA, Nukene EN, Danga SPY, Esimone CO. Larvicidal effect of *Lantana camara* and *Ocimum gratissimum* leaves extracts and their isolates against *Aedes aegypti* Larvae (Diptera: Culicidae). *J Mosquito Res* 2016;6:1-10.
60. Sosan MB, Adewoyin FB, Adewunmi CO. Larvicidal properties of three indigenous plant oils on the mosquito *Aedes aegypti*. *Nig J Nat Prod Med* 2001;5: 30-33.
61. Gbolade AA, Oyedele AO, Sosan MB, Adewoyin FB, Soyelu OL. Mosquito repellent activities of essential oils from two Nigerian *Ocimum* species. *J Trop Med Pl* 2000;1:146-48.
62. Adefolalu FS, Ogbadoyi EO, Ndams IS, Mann A. Larvicidal activities of N-Hexane fraction of *Ocimum gratissimum* leaf against mosquito larvae and its GC-MS analysis of phytoconstituents. *J Appl Life Sci Int* 2015;2:175-88.
63. Pratheeba T, Prabhavathi O, Yuvarajan R, Murugan N, Natarajan D. Identification of mosquitocidal compounds from the leaf extracts of *Ocimum gratissimum* (Lamiaceae) against dengue and chikungunya vector *Aedes Aegypti* (L.) *Int J Entomol Res* 2015;03: 67-79.
64. Ogendo et al. Bioactivity of *Ocimum gratissimum* L. oil and two of its constituents against five insect pests attacking stored food products. *J Stored Prod Res* 2008;44:328-34.
65. Ngamo et al. Protection of stored maize against *Sitophilus zeamais* (Motsch.) by use of essential oils of spices from Cameroon. *Meded Rijksuniv Gent Fak Landbouwk Toegep Biol Wet* 2001;66:473-8.
66. Ngamo Tinkeu et al. Persistence of the insecticidal activity of five essential oils on the maize weevil *Sitophilus zeamais* (Motsch.) (Coleoptera: Curculionidae). *Commun Agric Appl Biol Sci* 2004;69:145-7.
67. Goletti NMM, Benoit NM, Richard K, Marc C, Pascale C. Insecticidal formulation based on *Ocimum gratissimum* essential oil and montmorillonite clays for maize protection. *International Symposia on Entomology September 04, 2013, Orlando, FL, USA.*
68. Moussa KS, Vincent C, Schmit Jean-Pierre, Arnason JT, Bélanger A. Efficacy of essential oil of *Ocimum basilicum* L. and *O. gratissimum* L. applied as an insecticidal fumigant and powder to control *Callosobruchus maculatus* (Fab.) [Coleoptera: Bruchidae]. *J Stored Prod Res* 2001;37:339-49.
69. Musa AK, Oyerinde AA, Owolabi FO. Evaluation of the efficacy of mixed leaf powders of *Vernonia amygdalina* L. and *Ocimum gratissimum* Del.



- against *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae). *Acad J Entom* 2009;2:85-87.
70. Adeniyi SA, Orjiekwe CL, Ehiagbonare JE, Arimah BD. Preliminary phytochemical analysis and insecticidal activity of ethanolic extracts of four tropical plants (*Vernonia amygdalina*, *Sida acuta*, *Ocimum gratissimum* and *Telfaria occidentalis*) against beans weevil (*Acanthscelides obtectus*). *Int J Phy Sci* 2010;5:753-62.
71. Amoabeng et al. Tri-Trophic insecticidal effects of African plants against cabbage pests. *PLoS ONE* 2013;8:e78651.
72. Hewage CM, Bandara KANP, Karunaratne V, Bandara BMR, Wijesundara DSA. Insecticidal activity of some medicinal plants of Sri Lanka. *J Natn Sci Foundation Sri Lanka* 1997;25:141-50.
73. Ofuya TI, Okuku IE. Insecticidal effect of some plant extracts on the cowpea aphid *Aphis craccivora* Koch (Homoptera: Aphididae). *Anzeiger für Schädlingkunde Pflanzenschutz Umweltschutz* 1994;67:127-29.
74. Singh D, Singh A. Repellent and insecticidal properties of essential oils against housefly, *Musca domestica* L. *Insect Sci Appl* 1991;12:487-91.
75. Usip LP, Opara KN, Ibanga ES, Atting IA. Longitudinal evaluation of repellent activity of *Ocimum gratissimum* (Labiatae) volatile oil against *Simulium damnosum*. *Mem Inst Oswaldo Cruz* 2006;101:201-5.
76. Aisien M, Imasuen A, Wagbatsoma V, Ayinde B. Preliminary evaluation of the repellent activity of some plant essential oils against *Simulium damnosum* s.l., the vector of human onchocerciasis. *Int J Trop Insect Sci* 2004;24:196-99.
77. Pessoa LM, Morais SM, Bevilaqua CML, Luciano JHS. Antihelmintic activity of essential oil of *Ocimum gratissimum* Linn and eugenol against *Haemonchus contortus*. *Vet Parasitol* 2002;109:59-63.
78. Chatterje A, Sukul NC, Laskel S, Ghoshmajumadar S. Nematicides principal from two species of Lamiaceae. *J. Nematol* 1982;14:118-20.
79. Claudius-Cole AO, Aminu AE, Fawole B. Evaluation of plant extracts in the management of rootknot nematode *Meloidogyne incognita* on cowpea [*Vigna unguiculata* (L) Walp] *Mycopath* 2010;8:53-60.
80. Elele K, Gboeloh LB. Efficacy of neem extract (*Azadirachta indica*) and scent leaf extract (*Ocimum gratissimum*) on root knot nematodes of pepper plant (*Capsicum annum*). *J Sci Tech Res* 2012;5:56-64.
81. Moreira FJC, Santos CDG, e Renato I. Hatching and mortality of second-stage juveniles of *Meloidogyne incognita* race 2 in essential plant oils. *Rev Cienc Agron Fortaleza* 2009;40:441-48.
82. Fakae et al. Inhibition of glutathione S-transferases (GSTs) from parasitic nematodes by extracts from traditional Nigerian medicinal plants. *Phytother Res* 2000;14:630-4.
83. Thomas et al. Methanolic crude leaf extract of *Ocimum gratissimum* reverses phenylhydrazine-induced anemia in albino wistar rats. *Niger J Exp Clin Biosci* 2013;1:23-27.