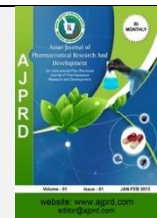


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Review Article

Ocimum Sanctum: Phytochemistry, Therapeutic Uses Pharmacological Activities and Its Anticancer Activities

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ABSTRACT

The practice of Ayurveda, which is based on the principles of living a healthy lifestyle and eating adaptogenic herbs on a regular basis, has the potential to alleviate a significant number of the chronic diseases that are the main cause of mortality across the globe. Tulsi, also known as *Ocimum sanctum* Linn, is considered to be the most significant herb in Ayurvedic medicine. Recent scientific research has demonstrated that it imparts a number of positive health effects. An increasing body of research indicates that the unique combination of pharmacological properties that tulsi possesses has the potential to alleviate stress on various levels, including the physical, physiological, metabolic, and psychological levels. Tulsi has been found to protect internal organs and tissues from the damaging effects of heavy metals and industrial pollutants, as well as against the impacts of physical stresses such as cold, excessive loudness, ischemia, and prolonged durations of physical exercise. This has been demonstrated through research. Plants that have medicinal properties are utilized on a daily basis by traditional healers in order to treat a wide variety of ailments. *Ocimum sanctum* Linn, a small herb that is widely distributed across India and is referred to as Tulsi in Hindi, has a long history of application in traditional medicine for the treatment of a wide range of ailments. These ailments include bronchitis, bronchial asthma, malaria, diarrhea, dysentery, skin diseases, arthritis, painful eye diseases, chronic fever, insect bites, and many others. There is some evidence to suggest that *O. sanctum* L. may possess qualities that include antibacterial, anticancer, antidiabetic, neuroprotective, and hepatoprotective. It has been demonstrated that *O. sanctum* L. possesses therapeutic value, as demonstrated by the pharmacological research that was incorporated into this study.

Keywords: Phytochemistry, pharmacological activity, anticancer, therapeutic uses

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INTRODUCTION

Since the beginning of Ayurvedic medicine, practitioners have relied on the numerous medical properties of *Ocimum sanctum* L., which is also known

as Tulsi or *Ocimum tenuiflorum*. This practice dates back thousands of years[1]. Tulsi, also known as the "Incomparable one" of India and the "queen of herbs," is considered to be among the most revered and cherished of the numerous medicinal and health-promoting herbs that are

native to the Eastern region. As a result of its lengthy history of application in Eastern holistic health systems like as Ayurveda and Unani, the sacred basil that is commonly referred to as Tulsi has received international recognition for the spiritual and religious importance it possesses[2]. In the volume of Ayurvedic literature known as the Charaka Samhita, the character Charaka makes a reference to it. Adaptability to stress is improved by tulsi, which is also considered an adaptogen due to the fact that it improves the balance of a number of different biological systems. According to Ayurveda, it is considered a "elixir of life" because of its powerful aroma and astringent flavor, both of which contribute to an increased longevity[3]. Tulsi extracts are used in the production of Ayurvedic medications for a wide range of conditions, including but not limited to malaria, inflammation, heart disease, headaches, stomach difficulties, and many sorts of poisoning and common colds[4]. Traditional preparations of *O. sanctum* L. include herbal tea, dried powder, and fresh leaf. However, there are also a range of other preparations available. The use of a combination of dried Tulsi leaves and grains has been used for a very long time in order to keep insects at bay[5].

The Charaka Samhita, which is a text from Indian traditional medicine, discusses the religious significance of holy basil (*Ocimum sanctum* L) as well as its numerous medical applications[6]. These applications include the plant's capacity to reduce blood sugar levels, alleviate anxiety, lower blood pressure, reduce inflammation, and eliminate microorganisms. It is also known as an adaptogen since it assists the body in adjusting to stressful situations, which is why it is called that. When it comes to traditional medicine

(ayurvedic and unani), medicines made from holy basil leaves have been shown to be effective in treating headaches, common colds, and inflammation[7]. There are numerous chemicals that may be discovered in the leaves of *Ocimum*, including β -caryophyllene, eugenol derivatives, vanillin, rosmarinic acid, ursolic acid, gallic acid, and vanillic acid. However, it is still difficult to extract and identify active flavonoids from these leaves, and there is no obvious association between the antioxidant effects of these substances and the flavonoids themselves. According to Iloki et al., flavones do not dissolve as well in water as they do in alcohols. However, flavones do dissolve better in water[8]. According to the findings of the study, the polarity of the solvent has the greatest impact on the extraction of phytochemicals and the antioxidant capabilities that plants possess. When attempting to determine which fraction includes the highest concentration of polyphenols, flavonoids, and active antioxidant components, it may be beneficial to employ successive fractionation with solvents that have variable degrees of polarity. On the other hand, there is no evidence that the leaves of this herb were extracted in a different manner. Therefore, the purpose of this work was to determine the most effective method for extracting flavonoids and polyphenols from the leaves of *O. sanctum* by means of a series of fractionations using a variety of solvents, and to investigate any potential relationships that may exist between the components and the antioxidant activity of the preparations that come about as a result of this process. The purpose of this study was to establish which fractions of flavonoids and their derivatives were the most effective in detecting the substances the researchers were looking for.



Figure 1: *Ocimum sanctum* whole plant

Phytochemistry

The complex phytochemical composition of the revered medicinal herb *Ocimum sanctum*, which is commonly referred to as Tulsi or Holy Basil, is responsible for the wide variety of positive properties that it possesses[9]. There are numerous essential oils that are found in the plant, including

as eugenol, methyl eugenol, β -caryophyllene, linalool, germacrene D, and camphor. The use of these oils has the potential to eliminate pests, lessen inflammation, alleviate pain, and prevent infections. The volatile compounds contained in the plant are responsible for both the plant's distinctive aroma and its wide range of medicinal applications[10]. There are a number of flavonoids and

phenolic acids that can be found in abundance in *Ocimum sanctum*, which also contains essential oils. Some of these flavonoids and acids include rosmarinic acid, apigenin, luteolin, quercetin, and cirsimaritin. Because of their powerful antioxidant characteristics, these polyphenolic compounds have anti-aging, neuroprotective, and hepatoprotective effects[11]. Through their ability to scavenge free radicals and reduce oxidative stress, these molecules have these effects. Furthermore, the plant holds a significant quantity of sterols and triterpenoids, such as β -sitosterol, ursolic acid, and oleanolic acid, which possess anti-inflammatory, cardioprotective, and anticancer properties[12]. The anti-inflammatory, tumor-inhibiting, and wound-healing capabilities of ursolic and oleanolic acids have been the primary focus of research in recent years. Additionally, the presence of alkaloids and ocimene derivatives, which further boost the plant's pharmacological potential, contributes to the enhancement of the adaptogenic and stress-relieving properties of *Ocimum sanctum*[13]. It has been demonstrated that the glycosides that are present in

Ocimum sanctum, such as vicenin and orientin, possess potent neuroprotective and radioprotective qualities. This further solidifies the herb's use in both traditional and alternative medicine[14].

The intricate phytochemical makeup of *Ocimum sanctum* is the reason for its widespread application in Ayurveda and other traditional medical systems for the treatment of a wide variety of ailments[15]. In addition to respiratory illnesses, diabetes, cardiovascular diseases, and immunological dysfunctions, these conditions also include immunological dysfunctions. In addition to the immunomodulatory benefits, the anti-diabetic action helps to regulate blood glucose levels, and the immunomodulatory activity strengthens the body's defense mechanisms[16]. *Ocimum sanctum* is utilized in the pharmaceutical industry, the nutraceutical industry, and the herbal medicine industry due to the way that its bioactive compounds interact with one another. This is a comprehensive treatment that has a great deal of potential in the medical field[17].

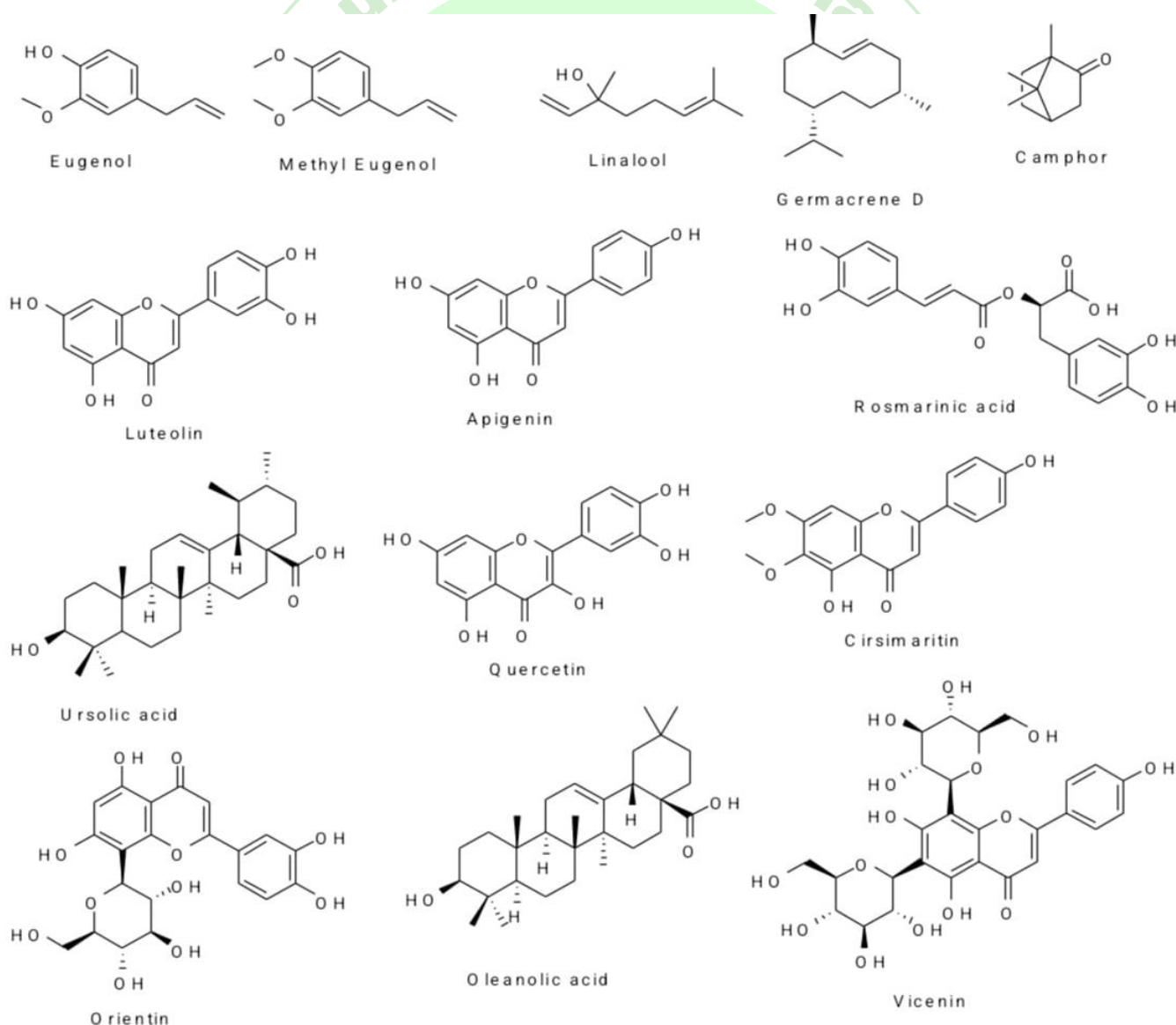


Figure 2: Some phytoconstituents structures of *Ocimum sanctum*

Therapeutic uses

Plants that have medicinal properties are utilized on a daily basis by traditional healers in order to treat a wide variety of ailments. *Ocimum sanctum* Linn, a small herb that is widely distributed across India and is referred to as Tulsi in Hindi, has a long history of application in traditional medicine for the treatment of a wide range of ailments[18]. These ailments include bronchitis, bronchial asthma, malaria, diarrhea, dysentery, skin diseases, arthritis, painful eye diseases, chronic fever, insect bites, and many others. It is possible that *Ocimum sanctum* L. possesses other therapeutic qualities, including those of an adaptogen, diaphoretic, hepatoprotective, cardioprotective, antiemetic, antispasmodic, analgesic, anticancer, antifungal, and antidiabetic. The active component that may be discovered in *Ocimum sanctum* L. is eugenol (1-hydroxy-2-methoxy-4-allylbenzene), which is primarily responsible for the therapeutic properties of Tulsi[19]. *Ocimum sanctum* L. has been used for a long time by traditional Indian healers to treat a wide range of maladies due to the plant's widespread occurrence and powerful therapeutic characteristics. However, there is a lack of evidence to justify the incorporation of this practice into contemporary medical thinking[20]. For the past few decades, Indian scientists and researchers have been investigating the pharmacological effects of steam-distilled, petroleum-ether, and benzene extracts of various parts of the Tulsi plant and eugenol on the immune system, reproductive system, central nervous system, cardiovascular system, gastric system, urinary system, and blood biochemistry[21]. These extracts have been used to study the effects of the Tulsi plant on these systems. The purpose of their work is to determine the therapeutic applications of *Ocimum sanctum* L. in contemporary medical practice. It has been suggested that tulsi possesses therapeutic relevance in the treatment of a variety of illnesses[22].

Pharmacological activities

• Antioxidant activity

Chaudhary *et al* The leaves of the medicinal plant *Ocimum sanctum* were first extracted in methanol (OsM), and then they were fractionated in a sequential manner with n-hexane (OsH), ethylacetate (OsE), and butanol (OsB)[23]. This was done in order to identify which solvent was the most effective for obtaining antioxidants from the leaves[24]. Both the total flavonoid contents (TFC) of OsB (54.51 ± 3.5 mg QE/g extract) and the total polyphenolic content (TPC) of OsB (212.26 ± 6.3 mg GAE/g extract) are found to be quite high[25]. Similarly, OsE displayed a significantly higher total phenolic content (TPC), at 202.71 ± 5.5 mg GAE/g extract. The EC₅₀ values for OsB were determined to be 3.91 ± 0.3 µg/ml for DPPH, 1.6 ± 0.1 µg/ml for ABTS, and 2.31 ± 0.1 µg/ml for phosphomolybdate[26]. For OsM, the hydroxyl radicals were found to be 5.3 ± 0.4 µg/ml, while for OsE, the phosphomolybdate radicals were found to be 2.43 ± 0.1 µg/ml, and ABTS (5.3 ± 0.4 µg/ml) for phosphomolybdate. These results indicate that OsE may possess potential antioxidant properties[27].

• Anti inflammatory activity

Anant *et al* Carrageenan-induced paw edema was utilized to evaluate the anti-inflammatory properties of OS extracts in hexane (STH), chloroform (STC), ethyl acetate (STE),

butanol (STB), and water (STW)[28]. The results of this study were given. The most active extract, STE, was subjected to additional testing in a dose-dependent manner. This was done in addition to verifying its oral toxicological profile and demonstrating its anti-inflammatory, analgesic, and antipyretic activities in small laboratory animals[29]. Establishing a chemical fingerprint through the utilization of high-performance liquid chromatography allowed for the determination of the solid-state terahertz (STE) fraction that was the most active[30]. The ethyl acetate fraction (STE) exhibits the highest level of anti-inflammatory activity, followed by the following fractions: STB, STW, STC, and STH[31]. According to the findings of a dose response study, STE displayed dose-dependent anti-inflammatory, analgesic, and antipyretic potential at a dosage of 2000 mg/kg. Furthermore, the study determined that STE did not cause any side effects[32]. The use of chemical fingerprinting allowed for the identification of flavanoids as being present.

Anti microbial activity

Sajjanshetty *et al* to assess controlled laboratory environment is used to evaluate the effectiveness of *Ocimum sanctum*, which is an extract derived from Tulsi leaves, as an antibacterial agent[33]. The preparation of the ethanolic extract of Tulsi was accomplished by the use of the cold extraction method. There were five different concentrations that could be obtained by diluting the extract with dimethyl formamide, which is an inert solvent. These concentrations were 0.5, 1, 2, 5, and 10% respective[34]. A positive control was carried out with doxycycline, whereas a negative control was carried out with dimethyl formamide. In order to determine the microbiological characteristics of the extract and the controls, *Aggregatibacter actinomycetemcomitans*, *Prevotella intermedia*, and *Porphyromonas gingivalis* examined the samples[35]. The agar well diffusion method was utilized in order to locate the concentration of Tulsi that resulted in the formation of an inhibitory zone that was comparable to that of doxycycline[36]. The Tukey post hoc test was applied for the purpose of comparing and contrasting the groups, and the one-way analysis of variance was utilized for the objective of analyzing the data. At dosages of 5% and 10%, the antibacterial activity of Tulsi extracts against *A. actinomycetemcomitans* was equivalent to that of doxycycline, with comparable inhibition zones ($P > 0.05$). This was the case when comparing the percentages of inhibition[37]. The presence of resistance to Tulsi extract was observed in *P. gingivalis* and *P. intermedia*, which exhibited a significant decrease in inhibitory zones ($P < 0.05$). In light of the fact that it possesses antibacterial properties against *A. actinomycetemcomitans*, tulsi has demonstrated that it has the potential to serve as an affordable and efficient "adjunct" to traditional periodontal therapy[38].

Antidiabetic activity

Leila *et al* A methanol extract of *Ocimum sanctum* L. leaves, which is a fraction that reduces blood sugar, was administered to diabetic rats in order to establish better control over hyperglycemia[39]. This was done in order to achieve the desired results. Additionally, assessments were conducted on the levels of α -amylase, α -glucosidase inhibitor, and insulin in the animals. In streptozotocin-induced rats, the researchers discovered that the highest dose of methanol extract considerably reduced blood glucose

levels in comparison to another dose[40]. This demonstrates that the extract has antihyperglycemic properties when it is taken orally. According to the findings of repeated administration of methanol fractions, the ethyl acetate-butanol fraction demonstrated a more effective antihyperglycemic impact than the chloroform and ethanol-water fractions[41]. In addition, the *in vitro* investigation indicated that the ethyl acetate-butanol fraction had the ability to regulate insulin levels as well as the activities of α -glucosidase and α -amylase enzymes, even when the concentration was low[42]. This was in contrast to the other fractions and acarbose, which was used as the positive control. Neither the methanol extract nor the fraction exhibited any change in insulin levels that could be considered statistically significant[43]. It was demonstrated by these findings that the presence of polyphenolics active components made it possible for the active crude extract (methanol) and its active fractions (ethyl acetate/butanol) to dramatically lower glucose levels[44].

Hepatoprotective activity

Lahon *et al* In the field of modern medicine, there is a deficiency in the availability of reliable hepatoprotective pharmaceuticals for the purpose of preventing and treating liver damage caused by drugs[45]. In traditional medicine, the hepatoprotective qualities of the leaves of sacred basil, also known as green tulsi (*Ocimum sanctum*), a plant belonging to the Lamiaceae family, have been recognized for a long time. It was the goal of this research to establish whether or not *Ocimum sanctum* possesses hepatoprotective properties on its own or whether or not it possesses synergistic effects when combined with silymarin[46]. The weight of the albino rats in each of the five groups ranged from 150 to 200 grams for each group. Group B had the experimental controls, while Group A contained the normal controls[47]. Normal controls were in Group A. Different quantities of an alcoholic extract of *Ocimum Sanctum* leaves (OSE) were administered to groups C, D, and E over the course of a period of ten days. The doses applied to groups C, D, and E were as follows: 200 mg/kg BW/day, 100 mg/kg BW/day of silymarin, and 100 mg/kg BW/day of OSE mixed with 50 mg/kg BW/day of silymarin, respectively. For the purpose of inducing hepatotoxicity, Groups B, C, D, and E were given paracetamol at a dosage of 2 grams per kilogram of body weight per day on the seventh day. In order to evaluate the hepatoprotective effect, serum protein tests, albumin globulin ratios, alkaline phosphatases, transaminases, and liver histology were utilized[48]. In order to present the results of the test for each group, the mean and the standard error of the mean (SEM) were utilized. An analysis of variance (ANOVA) using one-way and Bonferoni's test were utilized to compare the research group to the control group. In order for the P-value to be considered significant, it had to be less than 0.01. Furthermore, when compared to groups C, D, and E, which were much closer to normal ($P < 0.01$), group B had significantly higher levels of liver enzymes and an albumin globulin ratio. Upon histological examination, groups C, D, and E exhibited reduced sinusoidal congestion, hazy edema, fatty changes, and regenerating parts of the liver. Group B, on the other hand, solely had hepatic necrosis.

Fatemah *et al* *Ocimum sanctum* L. plant, often known as tulsi and belonging to the libiaceae family, has a long history of usage in traditional medicine as a treatment for a wide variety of medical conditions. These conditions include cancer, diabetes, heart disease, spasms, excessive sweating, and adaptogenic effects[49]. This experiment is being carried out with the purpose of determining the serum metabolites, determining the hepatoprotective efficacy of *O. sanctum* L. against galactosamine-induced toxicity, and analyzing the bioactive components that are present in each extract. High-performance thin-layer chromatography (HPTLC) was utilized in order to standardize and analyze the Ayurvedic method of extracting *O. sanctum* L. for the presence of biochemical markers. Rutin, ellagic acid, kaempferol, caffeic acid, quercetin, and epicatechin are some of the indications that are included in this category. In order to induce hepatotoxicity in adult albino rats, galactosamine was given intraperitoneally at a dose of 400 mg/kg. The quantitative hydroalcoholic and alcoholic extracts of *O. sanctum* L (100 and 200 mg/kg body weight/day) were compared in order to evaluate the hepatoprotective potential of the extracts. The evaluation included the reduction of histological damage, changes in serum enzymes (AST, ALT, and ALP), and an increase in TBARS. Twenty chemical components of *O. sanctum* serum metabolites were discovered and described through the process of comparing the mass spectra that were acquired by GC-MS with those that were obtained from the library-Wiley/NIST. We conducted *in vitro* investigations into the activity at each phase (hexane, chloroform, and ethyl acetate) to evaluate the hepatoprotective efficiency of the various hydroalcoholic extract fractions towards Chang liver cells. Our goal was to determine how the toxicity of CCL4 (40 mM) was affected by the different fractions of the hydroalcoholic extract. Because of its greater hepatoprotective efficiency in comparison to the other fractions, the ethyl acetate fraction of the selected plants was utilized for the purpose of vacuum liquid chromatography (VLC). The ethyl acetate fraction contains high amounts of a number of compounds that are responsible for hepatoprotection. These compounds include ruthin (0.34% w/w), ellagic acid (2.32% w/w), kaempferol (0.017 w/w), caffeic acid (0.005% w/w), quercetin (0.038 w/w), and epicatechin (0.057% w/w). Isolated bioactive compounds had a hepatoprotective activity that was significantly more extensive than that of standard silymarin in Chang liver cells that were subjected to CCl₄ toxicity. Purified ellagic acid exhibited a range of 70% at 100 μ g/ml to 81.33% at 200 μ g/ml, while purified rutin exhibited a range of 63.4% at 100 μ g/ml to 76.34% at 200 μ g/ml. This indicates a significant increase in hepatoprotection when compared to normal silymarin, which exhibited a range of 77.6% at 100 μ g/ml to 83.95% at 200 μ g/ml. One can observe that the concentration ranges of pure quercetin, epicatechin, and kaempferol are as follows: 54.33% at 100 μ g/ml, 60.64% at 200 μ g/ml, 53.22% at 100 μ g/ml, and 65.6% at 200 μ g/ml, respectively. On the basis of these findings, it would appear that the bioactive compounds found in *O. sanctum* L. greatly reduce the amount of liver damage that galactosamine is capable of causing[50].

Neuroprotective activity

Venuprasad *et al* Oxidative stress is a factor that contributes to the damage that cells sustain in a variety of diseases, including neurodegenerative disorders. The *ocimum sanctum*

plant is utilized extensively in the treatment of a wide variety of medical ailments in the Indian medical practice known as Ayurveda[51]. The purpose of this study was to investigate whether or not the hydroalcoholic extract of *O. sanctum* (OSE) might shield SH-SY5Y human brain cells from the oxidative stress that is brought on by hydrogen peroxide (H₂O₂). It was observed that the extract exhibited significant antioxidant activity against DPPH, 2,2'-azinobis (3-ethylbenzothiazoline-6-sulfonic acid) radical, and hydroxyl radicals, as indicated by the IC₅₀ values of 395 ± 16.2, 241 ± 11.5, and 188.6 ± 12.2 µg/ml, respectively[52]. One possible explanation for this is that the extract contains a significant amount of flavonoids and polyphenols. According to the data, the survival rate of cells decreased to 41.5% after being subjected to a challenge of 100 µM H₂O₂ over a period of 24 hours[53]. On the other hand, the survival rate of cells climbed to 73% after being pre-treated with OSE for a period of two hours. Furthermore, it prevented the cells from losing their form and decreased the amount of lactate dehydrogenase that was released. Lipide peroxidation, DNA damage, the formation of reactive oxygen species (ROS), and the depolarization of the mitochondrial membrane were all avoided by OSE[54]. In addition to restoring levels of catalase and superoxide dismutase enzymes and proteins, the extract resulted in a reduction in the overexpression of HSP-70 activity[55-57].

Anticancer activity

Mohammad *et al* leaf extract from *O. sanctum* was tested on the leukemic cell line, and the results were validated using the K562 cell line. In addition to this, we demonstrated that the cytotoxic effect was dose-dependent and that the activity was equivalent across all of the groups that were examined[58]. For as long as anyone can remember, medicinal herbs have been a source of information and inspiration for those working in the pharmaceutical industry[59]. In this study, the specific anti-cancer characteristics of the herb on leukemic cell lines are also explored that are not found in other studies[60]. Although the particular action mechanism was not studied in this study, the anticancer effects of the herb's components have been related to a number of different routes. For example, the herb has been shown to inhibit the growth of cancer cells[61].

Manaharan *et al* Among the several pharmacological properties that were demonstrated by the essential oil of *Ocimum sanctum* Linn, antifungal and antibacterial activity were among the most interesting[62]. The purpose of this study was to investigate the anticancer and apoptotic processes that are associated with *Ocimum sanctum* essential oil (OSEO). To extract OSEO from the leaves, hydrodistillation was utilized as the extraction method[63]. In order to evaluate the proliferation of cells, different dosages of OSEO were used. To investigate the process of apoptosis, tests were conducted using human breast cancer cells that were stained with propidium iodide (PI) and Hoechst. Based on the findings of our research, it was shown that OSEO had a significant impact on the proliferation of MCF-7 cells, with the dosage having a significant impact (IC₅₀ = 170 µg/ml)[64]. The fact that the OSEO caused an increase in the amount of apoptotic nucleic acids that were stained with PI in MCF-7 cells is additional evidence that the OSEO induced cell death. According to the analysis of flow cytometry, it

was shown that the apoptotic cell population increased by 16-84% dosage dependently when the cells were treated with OSEO (50-500 µg/ml) in comparison to the control group under study. Increasing the ratio of Bax to Bcl-2 and up-regulating the apoptotic genes p53 and Bid are both possible outcomes of OSEO[65].

CONCLUSION

Due to the numerous therapeutic advantages that it offers, *Ocimum sanctum* has been an indispensable component of traditional medicine for a considerable amount of time. It has been known for a long time for its antibacterial, antiviral, antifungal, antipyretic, antidiuretic, antidiabetic, and antimalarial activities. This is due to the high concentration of metabolites that it contains, which includes flavonoids, phenolics, and terpenoids. Because of the fact that these bioactive compounds have the potential to operate as antioxidants, anticancer agents, antibacterial agents, and antidiabetic agents, it is of the utmost importance to do additional research on them. Over the past several years, there has been a rise in the number of people who are interested in investing in the traditional health-promoting applications of tulsi. Based on the fact that the herb has been used historically in its natural, unprocessed state, it is likely that the nutritional and therapeutic benefits of the herb are the result of the interaction of a large number of phytochemicals that are active. Therefore, the advantages of Tulsi cannot be replicated in their entirety by using the extracts or components that make up the herbal plant. Because of the increasingly diverse range of chronic degenerative diseases that are caused by environmental factors, lifestyle choices, and individual stress, the traditionally fragmented approach of modern allopathic medicine has proven to be insurmountable. This is despite the numerous remarkable achievements that western medical science has accomplished. Traditional herbal therapies and holistic health practices are beginning to play an essential complementary role in the prevention and treatment of the condition known as passive sickness, which is prevalent in today's society. The World Health Organization has recognized the necessity of widening western medical viewpoints and has campaigned for the integration of traditional health and folk medicine systems with modern medical remedies. This is done with the goal of better addressing health problems on a worldwide scale. The body of research that suggests that Tulsi may have both preventative and therapeutic effects against stress-related degenerative diseases that are common in industrialized nations is expanding.

REFERENCES

1. Siva M, Shanmugam KR, Shanmugam B, Venkata SG, Ravi S, Sathyavelu RK, Mallikarjuna K. *Ocimum sanctum*: a review on the pharmacological properties. *Int. J. Basic Clin. Pharmacol.* 2016 May;5(3):558-65.
2. Pandey G, Madhuri S. Pharmacological activities of *Ocimum sanctum* (tulsi): a review. *Int J Pharm Sci Rev Res.* 2010 Nov;5(1):61-6.
3. Singh D, Chaudhuri PK. A review on phytochemical and pharmacological properties of Holy basil (*Ocimum sanctum* L.). *Industrial Crops and Products.* 2018 Aug 1;118:367-82.
4. Mahajan N, Rawal S, Verma M, Poddar M, Alok S. A phytopharmacological overview on *Ocimum* species with special emphasis on *Ocimum sanctum*. *Biomedicine & Preventive Nutrition.* 2013 Apr 1;3(2):185-92.

5. Rahman S, Islam R, Kamruzzaman M, Alam K, Jamal AH. *Ocimum sanctum* L.: A review of phytochemical and pharmacological profile. *American journal of drug discovery and Development*. 2011;1:1-5.
6. Pattanayak P, Behera P, Das D, Panda SK. *Ocimum sanctum* Linn. A reservoir plant for therapeutic applications: An overview. *Pharmacognosy reviews*. 2010 Jan;4(7):95.
7. Kumar R, Saha P, Lokare P, Datta K, Selvakumar P, Chourasia A. A systemic review of *Ocimum sanctum* (Tulsi): Morphological characteristics, phytoconstituents and therapeutic applications. *International Journal for Research in Applied Sciences and Biotechnology*. 2022 Apr 7;9(2):221-6.
8. Prakash PA, Gupta N. Therapeutic uses of *Ocimum sanctum* Linn (Tulsi) with a note on eugenol and its pharmacological actions: a short review. *Indian journal of physiology and pharmacology*. 2005 Apr 1;49(2):125.
9. Singh D, Chaudhuri PK. A review on phytochemical and pharmacological properties of Holy basil (*Ocimum sanctum* L.). *Industrial Crops and Products*. 2018 Aug 1;118:367-82.
10. Rahman S, Islam R, Kamruzzaman M, Alam K, Jamal AH. *Ocimum sanctum* L.: A review of phytochemical and pharmacological profile. *American journal of drug discovery and Development*. 2011;1:1-5.
11. Baseer M, Jain K. Review of Botany, Phytochemistry, Pharmacology, Contemporary applications and Toxicology of *Ocimum sanctum*. *International Journal of Pharmacy & Life Sciences*. 2016 Feb 1;7(2).
12. Mandal AK, Poudel M, Neupane NP, Verma A. Phytochemistry, pharmacology, and applications of *Ocimum sanctum* (Tulsi). In *Edible Plants in Health and Diseases: Volume II: Phytochemical and Pharmacological Properties* 2022 Mar 15 (pp. 135-174). Singapore: Springer Singapore.
13. Pradhan D, Biswasroy P, Haldar J, Cheruvanachari P, Dubey D, Rai VK, Kar B, Kar DM, Rath G, Ghosh G. A comprehensive review on phytochemistry, molecular pharmacology, clinical and translational outfit of *Ocimum sanctum* L. *South African Journal of Botany*. 2022 Nov 1;150:342-60.
14. Zahran EM, Abdelmohsen UR, Khalil HE, Desoukey SY, Fouad MA, Kamel MS. Diversity, phytochemical and medicinal potential of the genus *Ocimum* L.(Lamiaceae). *Phytochemistry Reviews*. 2020 Aug;19:907-53.
15. Chaudhary A, Sharma S, Mittal A, Gupta S, Dua A. Phytochemical and antioxidant profiling of *Ocimum sanctum*. *Journal of Food Science and technology*. 2020 Oct;57(10):3852-63.
16. Hanumanthaiha P, Panari H, Chebte A, Haile A, Belachew G. Tulsi (*Ocimum sanctum*)—a myriad medicinal plant, secrets behind the innumerable benefits. *Arabian Journal of Medicinal and Aromatic Plants*. 2020 Apr 14;6(1):105-27.
17. Enevide C. *Ocimum* Species: ethnomedicinal uses, phytochemistry and pharmacological importance. *International Journal of Current Research in Physiology and Pharmacology*. 2021 May 24;1-2.
18. Prakash PA, Gupta N. Therapeutic uses of *Ocimum sanctum* Linn (Tulsi) with a note on eugenol and its pharmacological actions: a short review. *Indian journal of physiology and pharmacology*. 2005 Apr 1;49(2):125.
19. Pattanayak P, Behera P, Das D, Panda SK. *Ocimum sanctum* Linn. A reservoir plant for therapeutic applications: An overview. *Pharmacognosy reviews*. 2010 Jan;4(7):95.
20. Singh N, Verma P, Pandey BR, Bhalla M. Therapeutic potential of *Ocimum sanctum* in prevention and treatment of cancer and exposure to radiation: An overview. *International Journal of pharmaceutical sciences and drug research*. 2012 Jul 4;4(2):97-104.
21. Singh V, Amdekar S, Verma O. *Ocimum sanctum* (tulsi): Bio-pharmacological activities. *Webmed Central Pharmacol*. 2010 Oct 22;1(10):1-7.
22. Siva M, Shanmugam KR, Shanmugam B, Venkata SG, Ravi S, Sathyavelu RK, Mallikarjuna K. *Ocimum sanctum*: a review on the pharmacological properties. *Int. J. Basic Clin. Pharmacol*. 2016 May;5(3):558-65.
23. Chaudhary A, Sharma S, Mittal A, Gupta S, Dua A. Phytochemical and antioxidant profiling of *Ocimum sanctum*. *Journal of Food Science and technology*. 2020 Oct;57(10):3852-63.
24. Rithichai P, Jirakiattikul Y, Nambuddee R, Itharat A. Effect of Salicylic Acid Foliar Application on Bioactive Compounds and Antioxidant Activity in Holy Basil (*Ocimum sanctum* L.). *International Journal of Agronomy*. 2024;2024(1):8159886.
25. Ashokkumar M, Palanisamy K, Ganesh Kumar A, Muthusamy C, Senthil Kumar KJ. Green synthesis of silver and copper nanoparticles and their composites using *Ocimum sanctum* leaf extract displayed enhanced antibacterial, antioxidant and anticancer potentials. *Artificial Cells, Nanomedicine, and Biotechnology*. 2024 Dec 31;52(1):438-48.
26. Pathak I, Niraula M, Sharma KR, Thapa P, Oli HB, Kalauni SK. Green Synthesis of Silver Nanoparticles from *Ocimum sanctum* Linn. and Study of Their Antioxidant Activity. *Amrit Research Journal*. 2024 Dec 31;5(1):82-90.
27. Zahid H, Saeed F, Lateef M. Anti-urease and antioxidant activity on selected medicinal plants: *Nigella sativa*, *Ricinus communis*, *Ocimum sanctum* and *Curcuma longa*. *Hamdard Medicus*. 2024;67(1).
28. Kumar A, Agarwal K, Maurya AK, Shanker K, Bushra U, Tandon S, Bawankule DU. Pharmacological and phytochemical evaluation of *Ocimum sanctum* root extracts for its anti-inflammatory, analgesic and antipyretic activities. *Pharmacognosy magazine*. 2015 May;11(Suppl 1):S217.
29. Varghese RM, Kumar A, Shanmugam R. Comparative anti-inflammatory activity of silver and zinc oxide nanoparticles synthesized using *Ocimum tenuiflorum* and *Ocimum gratissimum* herbal formulations. *Cureus*. 2024 Jan;16(1).
30. Bhattarai K, Bhattarai R, Pandey RD, Paudel B, Bhattarai HD. A Comprehensive Review of the Phytochemical Constituents and Bioactivities of *Ocimum tenuiflorum*. *The Scientific World Journal*. 2024;2024(1):8895039.
31. Dhanapal P, Shanmugam R, MD PM. *Ocimum gratissimum* mediated synthesis of AgNPs—An in vitro analysis of anti-inflammatory and antimicrobial effects. *Med J Malaysia*. 2025 Jan 1;80:1.
32. Sharma AD, Kaur I, Angish S, Thakur A, Sania S, Singh A. Comparative phytochemistry, antioxidant, antidiabetic, and anti-inflammatory activities of traditionally used *Ocimum basilicum* L. *Ocimum gratissimum* L., and *Ocimum tenuiflorum* L. *BioTechnologia*. 2022;103(2):131.
33. Mallikarjun S, Rao A, Rajesh G, Shenoy R, Pai M. Antimicrobial efficacy of Tulsi leaf (*Ocimum sanctum*) extract on periodontal pathogens: An in vitro study. *Journal of Indian Society of Periodontology*. 2016 Mar 1;20(2):145-50.
34. Al Ahdal K, Maawadh AM, Al Deeb L, Alshamrani AS, Almohareb T, Alrahlah A. Effect of malachite green, *ocimum sanctum*, and Er, Cr: YSGG laser on antimicrobial activity against *S. mutans* and CAD disinfection bonded to resin restoration. *Photodiagnosis and Photodynamic Therapy*. 2023 Jun 1;42:103571.
35. Panigrahi S, Rath CC. In vitro characterization of antimicrobial activity of an endophytic bacterium *Enterobacter cloacae* (MG001451) isolated from *Ocimum sanctum*. *South African Journal of Botany*. 2021 Dec 1;143:90-6.
36. Rai A, Singh A, Sharma A. Phytochemical analysis & antimicrobial activity of the leaves of *Ocimum sanctum*. *Journal of Pharmacognosy and Phytochemistry*. 2022;11(3):174-6.
37. Nasim I, Jabin Z, Kumar SR, Vishnupriya V. Green synthesis of calcium hydroxide-coated silver nanoparticles using *Andrographis paniculata* and *Ocimum sanctum* Linn. leaf extracts: An antimicrobial and cytotoxic activity. *Journal of Conservative Dentistry and Endodontics*. 2022 Jul 1;25(4):369-74.
38. Chaiyana W, Punyoyai C, Sriyab S, Prommaban A, Sirilun S, Maitip J, Chantawannakul P, Neimkhum W, Anuchapreeda S. Anti-Inflammatory and Antimicrobial Activities of Fermented *Ocimum sanctum* Linn. Extracts against Skin and Scalp Microorganisms. *Chemistry & Biodiversity*. 2022 Feb;19(2):e202100799.
39. Mousavi L, Salleh RM, Murugaiyah V. Antidiabetic and In Vitro Enzyme Inhibition Studies of Methanol Extract of *Ocimum tenuiflorum*

- Linn Leaves and Its Fractions. Tropical life sciences research. 2020 Apr;31(1):141.
40. Kesevan RK, Norakma MN, Zaibunnisa AH, Astuti PD. Total phenolic content and anti-diabetic activity of dried and fresh leaves of *Ocimum sanctum* (Linn). Food Research. 2023;7(4):139-45.
41. Vijay A, Nivethitha L, Mooventhan A. Anti-diabetic effect and its mechanism of action of *Ocimum* species (basil leaves) in type 2 diabetes mellitus: A narrative review. Advances in Integrative Medicine. 2024 Aug 14.
42. Arya R, Faruquee HM, Shakya H, Rahman SA, Begum MM, Biswas SK, Apu MA, Islam MA, Sheikh MM, Kim JJ. Harnessing the Antibacterial, Anti-Diabetic and Anti-Carcinogenic Properties of *Ocimum sanctum* Linn (Tulsi). Plants. 2024 Dec 16;13(24):3516.
43. Islam MT. Anti-diabetic potential of *Ocimum sanctum* Linn. Int J Adv Chem Res. 2022;6:38-41.
44. Mishra S, Sundaram S, Srivastava S, Dhar R. Phytosynthesis of Silver Nanoparticles Using *Ocimum sanctum* Leaf Extract and Studies on Its Antidiabetic, Antioxidant, and Antibacterial Properties. ACS Applied Bio Materials. 2023 Sep 26;6(10):4127-37.
45. Lahon K, Das S. Hepatoprotective activity of *Ocimum sanctum* alcoholic leaf extract against paracetamol-induced liver damage in Albino rats. Pharmacognosy research. 2011 Jan;3(1):13.
46. Kamel FO, Karim S, Bafail DA, Aldawsari HM, Kotta S, Ilyas UK. Hepatoprotective effects of bioactive compounds from traditional herb Tulsi (*Ocimum sanctum* Linn) against galactosamine-induced hepatotoxicity in rats. Frontiers in pharmacology. 2023 Oct 4;14:1213052.
47. Chatterjee A, Sarkar B. Polyphenols and terpenoids derived from *Ocimum* species as prospective hepatoprotective drug leads: a comprehensive mechanistic review. Phytochemistry Reviews. 2024 Jul 12:1-43.
48. Teofilović B, Tomas A, Martić N, Stilinović N, Popović M, Čapo I, Grujić N, Ilinčić B, Rašković A. Antioxidant and hepatoprotective potential of sweet basil (*Ocimum basilicum* L.) extract in acetaminophen-induced hepatotoxicity in rats. Journal of Functional Foods. 2021 Dec 1;87:104783.
49. Madhavalatha J, Kothamasu S, Kattapogu NK. Phytochemical profile, anti-oxidant and hepatoprotective activity *Ocimum tenuiflorum* leaves against carbon tetrachloride-induced hepatotoxicity in Mice. International Journal of Current Innovations in Advanced Research. 2024 Apr 1:15-35.
50. Almatroodi SA, Alsahli MA, Almatroudi A, Rahmani AH. *Ocimum sanctum*: role in diseases management through modulating various biological activity. Pharmacognosy Journal. 2020;12(5).
51. Venuprasad MP, Hemanth Kumar K, Khanum F. Neuroprotective effects of hydroalcoholic extract of *Ocimum sanctum* against H₂O₂ induced neuronal cell damage in SH-SY5Y cells via its antioxidative defence mechanism. Neurochemical research. 2013 Oct;38:2190-200.
52. Mataram MB, Hening P, Harjanti FN, Karnati S, Wasityastuti W, Nugrahaningsih DA, Kusindarta DL, Wihadmadyatami H. The neuroprotective effect of ethanolic extract *Ocimum sanctum* Linn. in the regulation of neuronal density in hippocampus areas as a central autobiography memory on the rat model of Alzheimer's disease. Journal of chemical neuroanatomy. 2021 Jan 1;111:101885.
53. Nandini HS, Krishna KL, Apattira C. Combination of *Ocimum sanctum* extract and Levetiracetam ameliorates cognitive dysfunction and hippocampal architecture in rat model of Alzheimer's disease. Journal of Chemical Neuroanatomy. 2022 Mar 1;120:102069.
54. Sarangi SC, Pattnaik SS, Katyal J, Kaleekal T, Dinda AK. An interaction study of *Ocimum sanctum* L. and levetiracetam in pentylenetetrazole kindling model of epilepsy. Journal of ethnopharmacology. 2020 Mar 1;249:112389.
55. Rodrigues V, Rao MS, Rao GS, KG MR. Neuroprotective potential of *ocimum sanctum* (Linn) leaf extract in preventing and attenuating stress induced substantia nigral neuronal damage in rats. Journal of Ayurveda and Integrative Medicine. 2022 Oct 1;13(4):100651.
56. Pattnaik SS, Sarangi SC, Sharma D, Sinha S, Nag TC, Nambirajan A, Tripathi M. *Ocimum sanctum* extract preserves neuronal echotexture and controls seizure in lithium-pilocarpine induced status epilepticus rats. Asian Pacific Journal of Tropical Biomedicine. 2023 Nov 1;13(11):479-87.
57. Seyed MA, Ayesha S, Azmi N, Al-Rabae FM, Al-Alawy AI, Al-Zahrani OR, Hawsawi Y. The neuroprotective attribution of *Ocimum basilicum*: A review on the prevention and management of neurodegenerative disorders. Future Journal of Pharmaceutical Sciences. 2021 Dec;7:1-4.
58. Harsha M, Kumar KM, Kagathur S, Amberkar VS. Effect of *Ocimum sanctum* extract on leukemic cell lines: A preliminary: in-vitro: study. Journal of Oral and Maxillofacial Pathology. 2020 Jan 1;24(1):93-8.
59. Hasan MR, Alotaibi BS, Althafar ZM, Mujamammi AH, Jameela J. An update on the therapeutic anticancer potential of *Ocimum sanctum* L.: "Elixir of life". Molecules. 2023 Jan 25;28(3):1193.
60. Perna S, Alawadhi H, Riva A, Allegrini P, Petrangolini G, Gasparri C, Alalwan TA, Rondanelli M. In vitro and in vivo anticancer activity of basil (*Ocimum* spp.): Current insights and future prospects. Cancers. 2022 May 11;14(10):2375.
61. Luke AM, Patnaik R, Kuriadom ST, Jaber M, Mathew S. An in vitro study of *Ocimum sanctum* as a chemotherapeutic agent on oral cancer cell-line. Saudi Journal of Biological Sciences. 2021 Jan 1;28(1):887-90.
62. Ahmad N, Ansari MA, Al-Mahmeed A, Joji RM, Saeed NK, Shahid M. Biogenic silver nanomaterials synthesized from *Ocimum sanctum* leaf extract exhibiting robust antimicrobial and anticancer activities: Exploring the therapeutic potential. Heliyon. 2024 Aug 15;10(15).
63. Boonyanugomol W, Rukseree K, Prapatpong P, Reamtong O, Baik SC, Jung M, Shin MK, Kang HL, Lee WK. An in vitro anti-cancer activity of *Ocimum tenuiflorum* essential oil by inducing apoptosis in human gastric cancer cell line. Medicina. 2021 Jul 30;57(8):784.
64. Khatoun S, Kalam N, Balasubramaniam VR, Shaikh MF, Ansari MT. Chemotherapeutic role of polyphenols present in *Ocimum sanctum*. Anti-Cancer Agents in Medicinal Chemistry-Anti-Cancer Agents). 2022 Dec 1;22(20):3325-42.
65. Manaharan T, Thirugnanasampandan R, Jayakumar R, Kanthimathi MS, Ramya G, Ramnath MG. Purified essential oil from *Ocimum sanctum* Linn. triggers the apoptotic mechanism in human breast cancer cells. Pharmacognosy magazine. 2016 May;12(Suppl 3):S327.