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Review Article

Pharmacognostic Characteristics, Chemistry, Biological Activity and Toxicity of *Lolium* Species

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ABSTRACT

Around seven species of the genous lolium poisonous grasses belonging to the family Poaceae are mutually grown in corps field over the world. In Iraq the prennel ryegrass is locally called "rewatta". The toxicity of these gasses are related to three chemically distinct alkaloids groups; the aminopyrrolizidine; lolines, indole-diterpenes (ergots, loliterms, and paxillines) as well as peramine alkaloids mostly concentrated in their seeds although indole-diterpene alkaloids loliterm B and paxilline biosynthesis requires endophytes symbiosis. The level of loline alkaloids enhances in both late summer-autumn of the year as well as in the infected dry plant materials up to 10 fold. However, paxilline and ergovaline are believed to be the precursor of the most toxic lolium species alkaloids, loliterm B, although, indole-diterpene alkaloids paxillines, loliterms and ergovaline are the actual indicators of Lolium species. In this review we summarize chemical characteristics, biological and toxicological influences as well as their interrelation of the plant of lolium genus. Central as well as peripheral biological/toxicological manifestations are summarized for both loline and indole-diterpene alkaloids. Finally, toxic influences of lolium alkaloids are function of their biological influences mostly exhibited via resembling molecular mechansims centrally as well as peripherallyare concluded.

Key words: Pharmacognostic, lolium, alkaloids, chemistry, biological, phytochemicals, toxicity.

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INTRODUCTION

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here are several species of the cool season grass that belongs to the genus Lolium which belongs to the family Poaceae or Gramineae subfamily Pooideae [1], including the perennial ryegrass *Lolium perenne* (meadow fescue) [2-5] or also called Lolium pratense (=F. pratensis, meadow fescue) [6-12], the tall fescue Loliumarundinaceum (Festuca arundinacea) [7, 13, 14], Lolium giganteum (=F. gigantea) [6], Lolium multiflorum, Lolium rigidum [15], Loliumcuneatum Nevski [16], and Lolium temulentum [15, 17]. In addition, there are hybrids of tall fescue and Lolium (tall fescue)or Lolium-Festuca [18, 19]. The ryegrasses and broad-leaf fescues of Lolium species is distributed Europe and the Mediterranean [1]. In the central parts of Asia the herb Lolium cuneatum Nevski grows as a poisonous weed within the fields wheat, barley, as well as flax [20] while Lolium temulentum L. is one of the predominant poisonous plants in Pakistan in the discrete Bannu, Khyber Pakhtunkhwa known as darnel or poisonryegrass besides, its global abundance in the cereal fields of the developing contries where it is considered as the worst weed [21-23]. Remarkably, in Egypt, Lolium temulentum (L. cuneatum) is reported to be one of five species belongs to the genus Lolium[24] contaminating the wheat crop where it is commonly known by the Arabic name "zawan", yet, dernel and invraie in other countries [25]. Nevertheless, thePerennial Ryegrass Lolium perenneis detected in Australia and North America, while, the Wimmera ryegrass Lolium rigidum Gaud. is located in Australia, South Africa, and, rarely, North America [26]. The common names of the Lolium species are listed in table (1) while their local names are listed in table (2) adapted from (Thomas, et al. 2011) [27]. It has been reported that the seed transmitted fungal clavicipitaceous endophytes including Neotyphodium or Epichloe" (Clavicipitaceae) are

associated with plants' production of nitrogen rich loline [1, 2, 6, 7, 9, 11, 15, 28-33], ergot lolitrems, peramine alkaloids [4, 34-38] and Festucine [14] that are chemoactive plant protective mechanism against vertebrate and invertebrate [2, 39]. Besides, various endophytes type fungus symbiosis including Acremonium coenophialum for is reported to be crucial for the biosynthesis of the pyrrolizidine as well as pyrrolopyrazine alkaloid in addition to their accumulation in the grasses [9, 35, 36] particularly peramine in *Lolium perenne* in symbiosis with *N. lolii*. [3]. however, Neotyphodium uncinatum is involved in the production of loline alkaloids in Lolium pretense [11, 12], Acremonium coenophialum Morgan-Jones and Gams are involved in the production of the considerable quantities of loline alkaloids in L. arundinacea Schreb. [40], yet, E. uncinata is involved in these alkaloids production in L. pratensis [41]. In this context, (Stegelmeier et al, 2013) have reported that such endophytes symbiosis with different Lolium species is associated with their neurotoxicity particularly that of Lolium perenne [26]. Remarkably, (Burhan, 1984) have reported the involvement of both endophytes symbiosis as well as plant age are related to the N-acetyl-loline (NAL) and N-formyl-Loline (NFL) production/concentration. In addition, the NAL and NFL levels are the highest after 9-11 weeks of seeding which declines after clipping (8≥weeks after seeding) [42, 43] which is greater in the Epichloe" Lolium-Festuca hybrids [44] as well as other pyrrolizidine alkaloids [18, 19], however, pyrrolizidine [18, 45] and ergopeptine [44] alkaloids is detected in non-infected(non-Epichloe")grasses. Moreover, (Liu, et al., 2001) have reported the direct involvement of L-homoserine in the loline alkaloids biosynthesis pathway in the Acremonium chrysogenum fungi [46]. Furthermore, (Craven et al., 2001) have reported that the type/amount of loline alkaloids phytosynthesis is associated with the genotype of fungus as well as its degree of symbiosis with the grass particularly Lolium pratense, reporting that the alkaloids level may mount 2% of the grass dry weight [8] which is in accordance to (Justus et al., 1997) who reported the highest level of loline alkaloids in the seeds infected with N. uncinatum [10].For example, theperamine mean level have been reported to be 26.21 \pm 2.97 μ g/g dry weight, of considerable toxic lolitrem B quantity, hence, exceeding 2 μ g/g of dry plant weight (the toxicity level) in E. festucae var. lolii infected L. perenne seeds from New Zealand in late summer, while, their level in fresh grass is 24-37% of the dry plant [2] on one hand.The loline and other related alkaloids producing lolium species-endophytes symbiosis is listed in table (3) adapted from (Schardl, 2007) and other reports [1, 5, 6, 8-10, 13, 15, 17, 30].

Table 1: common names of the Lolium species [27].

Species	Common name	
Lolium perenne	Perennial ryegrass	
Lolium multiform	Italian ryegrass	
Lolium rigidum	Annual ryegrass, wimmera ryegrass	
Lolium canariense	Canary island ryegrass	
Lolium loliaceum		
Lolium remotum	Hardy ryegrass	
Lolium temulentum	Darnel, poisonous ryegrass	
Lolium persicum	Persian darnel	

Table: 2 local names of the Lolium species ^[27].

language	Common names of Lolium species	
English	Bearded darnel, Poison darnel, Annual darnel, Red	
	darnel, Poison ryegrass, Darnel ryegrass, Ray-grass,	
	Tarse, Drake, Drawke, Drunk, Dragge, Study ryle,	
	Cheat, Wonwoer, Chess, Virginina oat, Cokil,	
	Cockle, Evir.	
Arabic	Zirwan, Samma, Aqoullab, Zawan, Zuwan	
	Shaylam, Suwal, Sikra, Danaqa	
Basque	Iraka	
Breesciano	Fraina, Lergheta, Loi	
Breton	Draog, Ivre, Pigal, Pilgere'h	
Calabrese	Giogghju	
Chinese	Du mai	
Colombia	Ballico	
Czech	Jelik	
Dutch	Dolik, HandsdarivK	
Estonian	Uimastav raihein	
French	Ivraie annuelle, Ivraie enivrante, Herb a couteau	
	Herb d'ivrogne, Zizanie	
German	Taumellolch, Taumel-Raygras	
Hungarian	Konloly	
India	Machni, Mochni, Mostaki	
Italian	Loglio del Veleno, Loglio ubriacaute, Zizzania	
Latin	Lolium temulentum	
Morocco	Zwan, Zuwan, Gesmata, I -medhum, Sirkran, Sikra	
	Saylam, Laichour	
Peru	Ballico, Cerisuelo, Sirisuela	
Polish	Kakol, Zycicac roczna	
portuguese	Joio	
Spanish	Borrachuela, Cizana commun, Cizana embriagente,	
8	Cominillo, Joyo, Trigollo, Mala hierba, Rabillo	
South Africa	Drabok raaigras, Dronkgras, Drabok	
Swedidh	Darrepe	
Romagnolo	Zizagna, Zizania	
Valencian	Brossa	
Zulu	Shesi	
Welsh	Efrau, Efryn, Yd meddw, Edrau coliog, Pabi'n	
	gwenith, Drewg, Pabi gwenith, Ller, Graban yn	
	hwylydd, Lleren	

 Table 3: Lolium species-endophytes reported symbiosis
 [1, 5, 6, 8-10, 13, 15, 17, 30, 31]

Specie	Endophyte	Country
Lolium arundinaceum	N. coenophialum	USA
L. arundinaceum	N. coenophialum	Morocco
L. giganteum (L.) S. J. Drabyshire	E. festuca	Europe
L. multiflormLam	N. occultans	South Africa
L. persicum Bioss et Hohen	N. occultans	Iran
L. pretense	N. uncinatum	Europe
L. pretense	N. siegelii craven et al.	Germany
Lolium sp.	Neotyphodium sp. FaTG- 3	Tunisia
L. rigidum Gaud.	N. occultans	Egypt
L. temulentum	N. occultans	Greece

However, (Blankenship, 2004) have also reported that the pyrrolizidine alkaloids such as loline class of these alkaloids in *Lolium pretense* is through ornithine-homospermidine pathway [4, 47] where spermidine [48].

LOLINE ALKALOIDS AND THEIR CHEMISTRY:

Globally, pyrrolizidine alkaloids are located in 3% of the flowering plants [49]. The level of these alkaloids varies with season, where in Germany grass lands it inclines to their optimum peak during summer to exceed the toxicity threshold in dry grasses (three folds of the fresh grasses), while, remains bellow this threshold in fresh ones particularly those of the toxic alkaloids peramine, lolitrem B, and ergovaline [2]. Similarly, in Kentucky, USA it has been reported that the levels of these alkaloids particularly the various loline alkaloids in tall fescue are within the range of low (200-300 μ g/g) during winter then inclines gradually during spring to approach their peak level during late summer [50] are reported in other studies [51-54]. In addition, (Bauer et al., 2018) have reported that pyrrolizidine/loline as well as indole-diterpene alkaloids such as paxilline and ergot levels are greater (up to ten folds) in the dry grasses as compared to an equivalent weight of the fresh grasses using immunochemical analysis [55].

(Hartmann, 1999) have classified these alkaloids into five classes these are class I and III which have α,β unsaturated necine core structure along with a macrocyclic, diester bridge between C9 and C7 including senecionine and monocrotaline. Second, Class II which have α,βunsaturated core structure along with open chaine diesters linked to C9 and C7 comprising triangularine type alkaloid. The third, is class IV and V with single ester side chain at C9 comprising pyrrolizidines [56] as shown in figure (2). Chemically, the core structure of this class pyrrolizidine alkaloids is composed of two fused saturated heterocyclic pentagonal rings with a nitrogen atom at one of the bridgehead with amine group substitution as what is identified in the loline alkaloids causing ring staggering [26]. In addition, these saturated amino-pyrrolizidine alkaloids isolated from Lolium species have exocylic oxygen bridge occurs between C2 and C7. These loline alkaloids isolated from endophytes infected Lolium perenne L., are reported to be in the greatest level in the seeds while the lowest in rachis, stem, leaf sheath, and leaf blade, nevertheless, the synthesis site is unknown, while, the plant age is also involved in these biologically significant pyrrolizidine alkaloids levels [9]. The seasonal effect on the level of loline alkaloids N-acetylloline (NAL) and Nformylloline (NFL) is also reported to be enhanced from April to be peaked during summer during July up to 1000 μ g/g [55, 57] as what is reported for samples from Alabama, USA. However, their level is not affected by the growth conditions as what is observed for perioline [56, 57]. Furthermore, plant clipping 6-7 weeks of seeding has been reported to bring about enhancement of NAL and NFL mean levels [42]. In addition, it is reported that water stress as well as temperature dramatically enhanced the level of NAL and NFL from 2236 to 11063 μ g/g within 12 weeks at 21/15 C⁰ particularly when tall fescue infected with endophytes which is also inclined as the nitrogen or phosphorus level inclined [45, 49, 58].

In general, loline chemically named hexahydro-N-methyl-2,4-methano-4H-furo[3,2-b]pyrrol-3-amine is a small size molecule a saturated 1-aminopyrrolizidines-type alkaloid originally isolated from *Loliumcuneatum* Nevski with a remarkable rigid simple structure of a characteristic polarity leading to an extraordinary physicochemical properties [4, 16, 30, 59, 60]. Structurally, their two fused saturated pentagonal rings sharing carbon and nitrogen atoms at their fusion ring with an oxygen ether bridge occur at the two carbons C2 and C7 (C2–O–C7 bridge) making this tricyclic ring system a very stained system [4, 30]. Interestingly, the endophyte infecting fungi are responsible for the oxygen ether bridge hence, completing the pyrrolizidine ring system [1]. Nevertheless, its exo-amino group (–NRR') occur at C1 group of different substitutions in various loline alkaloids plant secondary metabolite such as formyl, acetyl, and methyl groups [4] on its unusual tricyclic strained necine ring system (that is of –CH2OR group at C1 position) [33, 60-64]. There are other 1-aminopyrrolizidine alkaloids with α , β -unsaturation besides neither ether bridge at C2 and C7 positions nor amine functionalities at C1 position such as senecionine and retrorsine [65-67].

Loline alkaloids of exo-1- aminopyrrolizidine-2,7-ether nucleus structure are first isolated from tall fescue grasses (Lolium species) by (Petroski et al., 1989) who have synthesized the other loline alkaloids [40] then are detected in other plants although (Yunusov, Akramov, 1955) have reported the isolation of loline alkaloids from the seeds of Lolium cuneatum but with an incorrect elucidation of its chemical structure [16]. The first loline alkaloid temuline, later on called norloline, with no N-methyl substitution occur during isolation process as reported by (Dannhardt and Steindl, 1985) [17], is isolated from Lolium temulentum for the first time in 1892 by Hofmeister [17, 33, 68, 69] which was reported Longley later on to be the dominant alkaloids in this plant [17], while, (Katz, 1949) has reported no alkaloids existed in L.temulentum [70]. In fact, (Yunusov, Akramov, 1955) have reported is reported to be major alkaloid in L. temulentum [17]. However, its Nmethyl derivatives -NHCH₃ substituent are in *exo* position of the pyrrolizidine moiety as proposed by (Yates and Tookey, 1965) [14]. In addition, the term loline is proposed by Yunusov and Akramov who have isolated loline for the first time from rye grass, L. cuneatum Nevski at 1955 then its structure is conformed in 1965 and 1972 who conformed the existence of pyrrolizidine core of unique ether linkage at C2 and C7 [16, 69, 71-73]. Besides, the isolation and identification of other loline alkaloids from the same plant including norloline, N-acetylloline (or lolinine) [74], Nacetylnorloline [75] which is available along with other loline alkaloids in L arundinancea as a volatile alkaloids [31], N-methylloline, N-formylloline [76], Nformylnorloline [77] are lolines isolated from darnel and tall fescue [31, 78] in addition to N-acetylloline N-oxide [77] and a dimeric chlorine containing alkaloid lolidine is also isolated as [77, 79, 33] where one loline molecule is joined to a saturated pyrrolizidine that exhibit a chlorine at C7 and hydroxyl group at C2 instead of the ether bridge as reported by others [33, 77, 79, 80]. Moreover, (Yunusov and Akramov, 1960) have produced chlorinated and hydroxylated compounds without affecting the oxygen bridge of the pyrrolizidine which are then selectively removed to produce a mixture of N-methylpyrrolizidine resulting in endo-N-methyl-1-aminopyrrolizidine structure and free pyrrolizidine [81, 82] however, festucine without exo-position C1-N-methyl substitution have been isolated from Lolium arundinaceum (Schreb.) S.J. Darbyshire [14, 73]. In addition, the extraordinarily chlorinated alkaloid, lolidine is also isolated from the Lolium species [79]. Lolidine is structurally hetrogenous dimeric compound composed of loline and N-acetylnorloline fraction which is always isolated from their alkaloids extract ether fraction using chlorine containing solvents in all stages of isolation and purification. The chlorine atom in this halogenated in

the N-acetylnorloline part located at C6 carbon with contact oxygen bridge, yet, its opening in Chlorohydroxyloline and lolidinein [80] that occurs in the seeds of Lolium plants in the methanolic fraction [83]. In addition, norloline is obtained from the chlorine containing fraction of lolidine through alkalinization that cause oxygen ether bridge formation [83]. Moreover, (Dannhardt and Steindl, 1985) have reported the isolation of two major alkaloids loline as well as perioline from the carvopses and stem of the aerial parts of Lolium temulentum L while no detection of the loline demethylation alkaloid, norloline [17]. Nevertheless, the loline alkaloids isolated from the seeds of L. temulentum are loline chemical metabolic intermediate analogues as well as fungal infection final degradation products, norloline, loline, 6-methylloline and lolinine where the final one is supposed to occur due to norloline Nmethylation as well as acetylation [84-86].

It is necessary to note that N-formyl loline is a biosynthetic analogue of loline while, N-Senecioyl norloline and acylnorloline, are loline alkaloids metabolites isolated from hours urine which feed on tall fescue grass [1, 87] which are reported to exhibit potent DNA binding potential [65, 66] as well as hepatotoxicity [56]. The Lolium species germinated seeds have been reported to exhibit a high level of loline alkaloids. In this context, (Yu et al., 1955) have reported the isolation of total alkaloids of Lolium cuneatum Nevski using chloroform and have been found to constitute 0.23% of the dry plant weight and are composed of loline, nortoline, lolinine (N-acetylloline), N-methylloline, and Nacetylnorloline while the aqueous fraction of the extract contains N-formylloline as it is quaternary base alkaloid. Loline constitutes 45% of the chloroform extract while, lolinine constitute 41.7% [79]. Thus, N-formylloline, Nacetylloline, N-methylloline, norloline, N-acetylnofloline and N-formylnofloline are isolated from Lolium cuneaturn and Loliurn temulentum. Moreover, these alkaloids including the two enantiomers of N-formylloline and Nacetylloline are reported to be existed in the plant parts of endophyte-infected tall fescue Lolium pretense, yet, the highest concentration is detected in the seeds, while, 1000 μ g/ml concentration is reported in the fungal filtrate. Interestingly, (Blankenship, 2004) have reported that the biosynthetic final steps of loline alkaloids follows the following order: norloline \rightarrow loline \rightarrow methylloline \rightarrow Nformylloline while ring C1 amine methylation happens before ring cyclization [4]. Nevertherless, the pharmacologically interesting new properties of loline alkaloids [9] has lead to the successful synthesis of (\pm) loline by (Tufariello et al, 1986) [88] while, N-formylloline as well as N-acetylloline have been synthesized from loline using ethyl formate at room temperature and acetyl chloride in chloroform respectively [9], however, N-acetylloline structure have been elucidated by (Bates, Morehead, 1972) Moreover, both of paxilline and ergovaline are [72]. reported to be the precursors of the indole diterpene alkaloid loliterm B as end product obtained from the Neotyphodium lolii and Epichloë infected grasses including perennial ryegrass, Lolium perenne [3, 89-94]. In England immunoassay EIA has demonstrated that the seeds infected with endophyte perennial ryegrass contains (3000 µg/kg) paxilline [95] while, (3000-5200 µg/kg) for that from

England and France [96, 97] while, ergovaline level in perennial ryegrass seeds from France is $6200 \ \mu g/kg$ [97]. Others have been reported that ergovaline level in perennial ryegrass dry material from Franse is $2300 \ \mu g/kg$, yet, $4700 \ \mu g/kg$ from Czech [97, 98]. However, approximately close concentration of paxilline as well as lolitrem B have been reported for perennial ryegrass from New Zealand [99, 100] in addition to lolitrem B in Germany and other European countries [53, 97, 101-103].

Furthermore, (Vikuk, et al., 2020) have reported that the E. festucae var. lolii seeds of L. perenne from New Zealand contains peramine, lolitrem B and ergovaline in a season dependent concentrations that incline during summer while decline during winter. The concentration of peramine is found to be within the range above the toxic range 0.04 and 23.38 μ g/g dry weight (2.00 + 0.32 μ g/g, 6.57 + 1.09 μ g/g and 3.23 \pm 0.61 μ g/g are the mean levels in July, August and September respectively) which is above the toxic one and half of that reported in Germany while its concentration in fresh plant is 13-40% of the dry weight. While, the detected level of lolitrem B is within the range of 0.07 and 23.81 μ g/g which is above the toxic one particularly during summer. However, paxilline is not detected while ergovaline is very low 0.3–0.4 μ g/g (DW) which is bellow the toxicity threshold however, ergovalline level reaches its peak in July to be $1.33 \pm 0.30 \,\mu \text{g/g}$ dry weight while in the fresh plant is 20-40% of its concentration in dry material [2]. Moreover, (Bauer, et al., 2018) have reported that paxilline congers are the dominant lipophilic secondary loline alkaloids metabolites in the ethyl acetate extracts of the seeds and the fresh plant of perennial ryegrass, L. perenne L. as well as in the seeds of the Italian ryegrass L. multiflorum Fabio obtained from Germany which are 1'-O-acetylpaxilline and 13desoxypaxilline. Besides, the existence of paxilline-like indole diterpene and ergot alkaloids in the seeds as well as the fresh plant of perennial ryegrass, however, weak concentrations, 7.3 µg/kg, of ergot alkaloids is detected in the seeds of the Italian ryegrass while, no detection to paxilline alkaloid. Remarkably, immunoassay have indicated the availability of high concentration of paxillin alkaloid, (5400 µg/kg), and ergot alkaloids, (260 µg/kg), in the dry matter of perennial ryegrass. Nevertheless, in south Germany the level of paxilline is 110 µg/kg in fresh plant while 270 µg/kg in dry matter which mostly related paxilline-like analogues due to the cross-reactivity to paxilline since according to immunoassay paxilline mean level of 190 190 μ g/kg represent > 3% of the total paxilline alkaloids (6800 µg/kg) in perennial ryegrass. In addition, the mean level of ergot alkaloids in the seeds and dry matter of perennial ryegrass is reported to be 1600 and 180 µg/kg ergometrine equivalents, respectively, yet, ergovaline actual levels are of 24000-80000 µg/kg and 3000-9000 µg/kg in the seeds and plant dry mater that make it the major indolediterpene alkaloids. Thus, paxilline as well as its paxillinelike indole-diterpene analogues levels are good indicators for the plant toxicity [55]. The chemical structures of the summarized loline alkaloids is illustrated in figure (3).

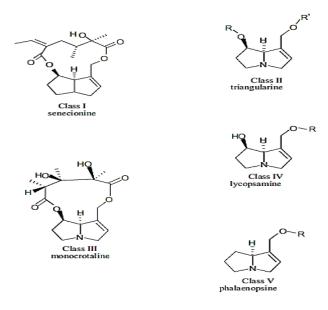


Figure 2: Classes of plants origin pyrrolizidine type alkaloids^[56].

Finally, some authors have reported that there is a relationship between the accumulations of peramine and ergovaline through out the year while no correlation between N-acetylloline and N-formylloine accumulation and the accumulation of peramine or ergovaline meaning that both N-acetylloline and N-formylloine are not synthesized by the infecting endophytes [4].

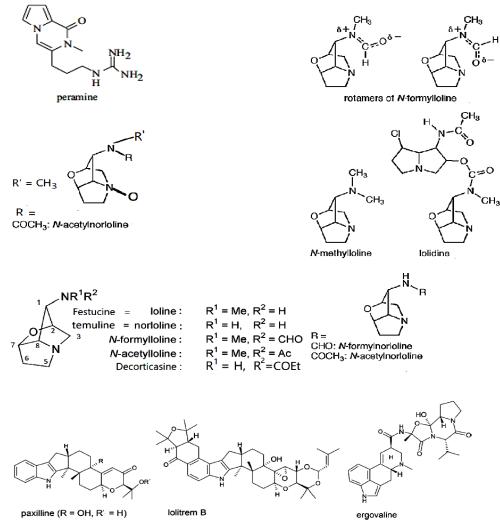


Figure 3: Chemical structure of the summerized lipophilic indole-diterpene and hydrophilic loline alkaloids.

Loline As Well As Paxilline/Paxilline-Like Diterpene Alkaloids Biological Influences and Their Toxicity:

Loline alkaloids particularly temuline from Lolium temulentum in darnel beer have been traditionally used as sedative for reliveing pain as well as sedative for nervous conditions [104]. Nevertheless, it is earlier reported that loline alkaloids as well as ergovaline interacts with the α_2 adrenergic, D₂ dopamine, or serotonergic receptors in the blood vasculatures leading to vasoconstriction besides, vasculatures thickening [105, 106], in addition to their immunosuppersive influence in murine model [107]. However, (Dannhardt, Steindl. 1985) have reported that the α 1-adrenergic, α 2-adrenergic, D₂ dopamine, cholinergic, serotonergic and benzodiazepine receptors bindings in rat and calf brains are not interfered by loline dihydrochloride as it has no affinity to these receptors [17, 108-112]. Whereas, other have suggested various central nervous system receptor antagonizing influences particularly the dopaminergic receptors [17, 113, 114]. Interestingly, unlike ergot alkaloids which are of no influence at concentration of 1.9 ppm, anorexic feed as well as prolactin depressing influences have been reported to a partially purified of tall fescus extract fraction of these pyrrolizidine alkaloids in rat model [115, 116]. In addition, (Strickland, et al. 1994) have reported a dose dependent prolactin suppression influence of loline at elevated concentration of 10⁻⁴ M via D₂ receptor agonistic effect in rat model [117]. Nevertheless, at similar level other loline derivatives such as N-acetylloline, Nformylloline and N-methylloline have exploited no prolactin inhibition activity in rats [118], particularly, Nacetylloline and N-formylloline that have no D₂ receptor or cAMP counteracting effect even at milimolar level [114]. Tall fescue extract has been reported to exhibit α_2 adrenergic receptor powerful agonistic influence leading to a potent contractility in the cattle's lateral saphenous veins [119] while, N-acetylloline and N-formylloline mixture at 3 x 10^{-5} M have been reported to cause partial inhibition of the norepinephrine mediated vasoconstriction [120]. However, N-acetylloline have been also reported to exploit briefly lower cow lateral saphenous veins and arteries vasoconstrictive effect in hourses [121, 122] indicating serotonergic and α_2 adrenergic receptor antagonistic influences [123]. Nevertheless, N-acetyl loline have been reported to elicit in vitro vascular smooth muscle cells at lower concentration while, at higher levels of 10⁻⁸ and 10⁻⁹ M have prohibited their growth, yet, at concentration range of 10⁻⁸-10⁻¹¹ M stimulate the growth of the dormant cells in bovine models [105]. Meanwhile, it is necessary to note that the in vitro tall fescue lolines vascular as well as endocrinal influences are exploited only at quietly high levels [117, 119, 120]. Moreover, it has been reported that these lolium pyrrolizidine alkaloids in excessive dose may inhibit thiaminase I enzyme responsible for thiamin catabolism through acting as enzyme substrate leading to thiamin deficiency in castles [124, 125].

Loline dichorohydrate as well as N-benzoyl iodomethylate derivatives have been found to decline coronary blood flow as well as blood pressure in mammalians such as cats and doges. In this context, loline alkaloids are reported to exhibit cardiac arrest subsequent to their negative inotropic effect at 0.1 and 1 mM concentration, however, cardiac arrest in froge model. Where as, at 10 mM level they have demonstrated instant cardiac arrest while diastole. In addition. intravenous administration of loline dihydrochloride at doses of 1-60 mg/Kg have been reported to cause dose-dependent reduction of blood pressure as well as respiratory deepening for 2-15 minutes [126]. Moreover, (Hammouda, et al., 1988) have reported that the loline alkaloids of the seeds ethanolic extract of Lolium temulentum exploit fast onset/short duration reversible cardiac arrest via depressing the predilective inotropic rather than the chorotropic function of the atrium in rabbit heart model along with very weak antagonistic effect against the tone as well as contractility of isolated rabbit aortic spiral strip vascular smooth muscle treated with epinephrine as compared to loline dihydrochloride which lacks such effect despites the later higher adrenaline tissue sensitizing influence. Furthermore, against the GIT, they have demonstrated very potent muscle tone depression on rabbit's jejunum smooth muscles model greater than their effect on their rhythmic contractility. However, on guinea pig ilium smooth muscles, the lower concentration of the loline alkaloids containing ethanolic extract have found to potentiate acetylcholine activity while, opposite activity is observed in the higher concentration along with uniform histamine effect antagonism at both concentrations explained by the total loline alkaloids synergistic influences. Meanwhile, loline dihydrochloride has exhibited no considerable effect on muscle tone on the same model while great enhancement of ilium contractility. However, both loline dihydrochloride as well as ethanolic extract total loline alkaloids have lacked any considerable influences against the tone of smooth muscle. Yet, the extract's total alkaloid have exploited resembling partial antagonistic action against Ach and histamine-induced contractions, but, loline dihydrochloride has elicited a selective histaminergic contraction antagonistic influence in ilium model. Moreover, the total alkaloids extract have elicited less Ach blood level influencing effect than loline since, the later exhibits more potent serum pseudocholine esterase inhibitory influence explaining its ability to provoke the transmission of partially irreversible neuromuscular blocking impulse in a perfused diaphragm rat's phrenic nerve model. On the CNS, the ethanolic extract total loline alkaloids have demonstrated CNS depression accompanied by hind limbs paralysis as well as ataxia hence, ultimately have brought about complete motor incapacitation, total skeletal muscle hypotonia along with postural reflexes/external stimuli response in mice model which have been found to be significantly greater than that to loline dihydrochloride. In addition, both ethanolic extract alkaloids as well as loline dihydrochloride have exploited dose-dependent time prolongation of barbiturate based hypnoptic activity [84]. Furthermore, (Putnam, et al, 1991) have reported that feeding of pregnant mares on endophytes infected Acremonium coenophialum tall fescue containing loline alkaloid derivatives 1610 μ g/g NAL plus NFL as well as 0.39 μ g/g of ergovaline plus ergovalinine has caused abortion or after lethality of birth fetus during pregnancy, after birth or during lactation in 90% of these fetuses. However, this toxic effect is not known whether due to a direct or indirect influences on mammalians [58].

Remarkably, N-acetylloline has been reported to exploit an in vitro mitogenic on the stationary smooth muscles of the blood vessels 10-1000 pM concentration while prohibit cell growth of others at 100-1000 pM concentration [105]. In addition, (Petroski et al., 1994) have been reported that loline alkaloids as well as their semisynthetic derivatives of 8-12 carbon acyl chain substituting C1 amino group have significant antineoplastic influences against solid tumor in brine-shrimp model of 0.274 μ g/ml mean ED₅₀ against human lung carcinoma A-549, breast carcinoma MCF-7, and colon adenocarcinoma HT-29, yet, through an unknown mode of action. However, N-Acyllolines of >12 carbon length have a non-significant with weak cyctotoxicities as what is observed for the parent congener loline, yet, those with 12-18 carbon atom length acyl substitutions have exploited significant cytotoxic influences although much weaker than that of the anticancer drug adriamycin. Interestingly, the most active N-acylloline congener is N-Oleoylloline which has exhibited some degree of selectivity against HT-29 human colon adenocarcinoma [127]. Whereas, in vivo studies have demonstrated that the metabolism of the naturally occurring pyrrolizidine alkaloids including loline alkaloids gives rise to non-cytotoxic metabolites that can't interact with the intracellular macromolecules [128].

Moreover, the Lolium species indole-diterpene neuro-active mycotoxin alkaloid, lolitrem B rich in seeds of endophyteinfected ryegrass seed have been reported to exhibit an antimammalian influences involving neurological symptoms of hyperexcitability, muscle tremors and ataxia. Yet, it develops clonic seizures then death in sever cases of toxicity in male mice model as it has been detected in liver, kidney, cerebral cortex and thalamus as they are presumed to be its primary site of influence, but not in the cerebellum as well as brain stem which also encountered in sheep and cattle [95, 129, 130]. It mode of action may involve alteration in the metabolic pathways of essential neurotransmitters like caticholeamines as well as amino acids like tyrosine as their profile have been found to be significantly altered with time. Thus, lolitrem B exhibits its toxic influence in the CNS via regions specific manner particularly in the cerebral cortex that involves the emotion, mental and cognition functions of the brain as it accumulate in this tissue within a short period of time through perturbation of neurotransmitters metabolism leading to tremors and behavioral alterations in both high and low doses in a dose dependent mode. Lolitrem B, can disturb the metabolism of branched amino acids leading to their accumulation in the brain, hence, it deregulates the sedative neuromodulatory/catecholamine pathways leading to enhance its tremorgenic and non-tremorgenic excitatory influence in the forebrain [130]. Centrally mediated, Loliterm B, prolonged/reversible tremorgenic influence have been also reported in sheep and murine models [90, 131-136] which may have a pharmacological, discovery as well as drug design importance [132]. The mode of action of loliterms is reported to undergo structure dependent large conductance calcium-activated potassium channels potent blockage [90, 137, 138, 139] that maintain its neurological, emotional, behavioral as well as motor activity disturbance 137] besides, explaining its potential [90, 133. pharmacological action [138]. Although others have reported that tremorgenicity is indirectly related to the

blockage of these channels besides, not seldom mechanism contributing to its symptoms of toxicity [138]. However, their metabolic alteration in vivo declines their activity due to the resulted structural alteration as it have observed in animal models [137-139]. However, stereochemistry have a role in the determining loliterm termjorgenic influence. For example, its natural stereoisomer lolitrem F. of A ring of alpha-phase is of no such influence [140] Remarkably, loliterm B binds to the charybdotoxin binding allosteric site located in the pore of the channel [141]. However, due to its lipophilicity, large molecular weight and non-volatility, lolitrem B tends to accumulate in the fatty tissues including the brain tissues [142-144] which is detected in fatty tissues of lactating and mature animals [143-147] as in case of sheep [142, 143]. Authors have speculated that loliterm as well as its biosynthetic intermediates including paxilline and terpendole C induce its termorgenic influence via close molecular mechanisms in mice model [92, 131]. In addition, a contractile tension inducing synergistic influence on the sheep distal colon smooth muscle longitudinal preparation have been reported for it combination with ergotamine leading to diarrhea [148], while its combination with ergovaline leads to decline bovine milk production [149-151]. In addition, in sheeps its GIT influence against duodenum via interfering the acetylcholine release [152]. It is reported that interpertoneal loliterm B tremorgenic influence lasts longer than other indole-diterpene alkaloids like aflatrem while much potent than paxilline [131, 145]. Remarkably, both lolitrem B as well as its intermediate metabolite, 31-epi-lolitrem B, significantly attenuate the production of IL-6 and TNF α cytokines production in murine macrophages, whereas, no cytotoxic influence have been observed against the viable cells even at 100-250 folds higher levels making them an excellent candidates for designing immune modulator drugs [153]. It is noteworthy to know that lolitrem is structurally related to the other lolium indole-diterpene alkaloid tremorgen, paxilline [154], however, loliterm B, maximum tremorgenic influence is exhibited at a dose of 8.0 mg/Kg of body weight [131]. While other Lolium metabolites such as lolilline and lolitriol have no tremorgenic influence [155, 156]. Nevertheless, despite that both of Lolitrem A, B and ergovaline are of the major mycotoxins in endophyteinfected perennial ryegrass and tall fescue respectively [147, 157], however, the level of lolitrem B is five to ten folds than ergovaline knowing that in lamb as well as lactating ews the required toxic threshold of ergovaline [140, 158-161] is much lower that of lolitrem B [147, 162, 163] of 1800-2000 μ g/kg in cattle and sheep [164].

Furthermore, un like loline alkaloids which has weaker influence as well as peramine which has no influence on mammalians than ergot alkaloids and other indole-diterpene alkaloids, ergot alkaloids of lolium specie including ergovaline exhibit their potent toxicities through prohibiting CNS neurotransmitters metabolism besides, endocrine function counteraction influence potentially mediated through dopaminergic pathway interaction mode of action indicated by the low blood melatonin and prolactin levels [39]. In this context, (Larson, et al. 1995) have reported that ergovaline exploits an elevated affinity to the dopaminergic receptors, in addition to, its intestinal vasoactive peptide as well as cAMP production stimulants [106]. In fact, ergovaline is the most toxic and abundant ergopeptide alkaloid in the infected tall fescue exhibiting similar neurological/motor symptoms of lolitrem B [165, 166]. However, in lactating ewes both ergovaline in particular and lolitrem B exhibit mild activity against certain drug-metabolizing enzymes [147, 162].

Moreover, the other Lolium species indole-diterpene alkaloid, paxilline of weak tremoergenic influence [29, 95] induce GIT smooth muscles various stimulation responses particularly in sheep duodenum [156] which is reported by (McLeay, et al. 1999) to be coincided with its skeletal muscles directed tremoring influence, hence, disturbing digestion as encountered with lolitrem B [132]. In murine model, its tremorgenic influence prolongs for many hours with LD₅₀ of 150 mg/kg body weight [167]. In fact, paxilline is a powerful smooth muscle high conductance calcium-activated BKca channel blocker [138] in very low Ka value range of (2-10) nM as encountered in bovine aortic smooth muscle channels [168, 169]. In addition, (Selala, et al. 1991), have also reported their contribution to smooth muscles contractions in guinea-pig ileum model via blocking these BKca channels along with enhancing acetylcholine release [170]. Interestingly, paxilline along with its novel congeners pyrapaxilline and 21isopentenylpaxilline, have been reported to prohibit nitrous oxide NO production in murine RAW264.7 cell line, nevertheless, they elecited their influences at 30 mg/ml and 10 mg/ml concentration respectively. The later compound greater activity is related to its additional dihydropyran ring [171]. Further more, paxilline has been also reported to attenuate the macrophages lipopolysaccharides-induced singnaling of the IkB-a/NF-Kb signaling pathway [172]. In addition, other remarkably unexpected antiviral against H1N1 influenza virus is reported for paxilline as well as other related congeners, 21-isopentenylpaxilline, paspaline, and dehydroxypaxilline [173].

In addition, paxilline at 0.1-10 μ M level has been reported to incline rodents urinary bladder, and duodenum spontaneous tension which can not be reversed by atropine, while, treachea spasm in guinea-pig in a dose dependent manner via blocking the high conductance Ca²⁺-activated potassium channel, despite, its inactivity against their isolated portal vein and aortic rings at 1-10 μ M concentration. Yet, authors have expected stimulation may happens at concentration higher than 10 μ M. However, at 10 μ M paxilline inclines the integrated myogenic of the bladder by (9.6 ± 2.8) folds of their basal level [138] via promoting acetylcholine release from nerve terminals [170] although it dose not include muscarinic receptors agonistic influence. In addition, at concentration of 10 μ M paxilline induces treacheal spasm to an extent around one quarter of the maximum influence of carbacol similar dose within 20 min. In deed, (DeFarias, et al. 1996) have spectulated that paxilline through blocking the BKca channel conductance prolongs the action potential hence, inclines the intracellular calcium inflex to the sarcolemma while the excitation-contraction coupling process as the molecular mechanism for its smooth muscle stimulation influence on the rodent bladder as well as that of GIT. Besides, they have concluded that paxilline synergistically potentiates charybdotoxin stimulatory influence on guinea-pig bladder

[174]. Earlier, (Knaus, et al. 1994) have reported that paxilline enhances the binding of charybdotoxin to the BKca channel through paxilline binding to an allosteric site that enhances the receptor, located in the channel pore, affinity to charybdotoxin, although paxilline by itself is a powerful channel blocker as it permeate through the affected cell plasma membrane, hence, exploiting full blocking influence [138]. Remarkably in sheeps both paxilline, and lolitrem B have been also reported to stimulate skeletal muscles while, both stimulate and inhibits duodenum smooth muscles although their stimulatory effect can be partially antagonized by atropine [175, 176]. Furthermore, since, paxilline's BKca channel blockage is calcium dependent thus, this blockage effect is declined with the incline of calcium ion level that enhances the channel conductance [138, 177, 178]. Paxilline have been reported to be detected in rats brain membrane besides, inhibiting the GABA-induced chlorine influx into microsacs through binding to GABAAreceptors as it can pass the blood brain barrier passing rapidly into the synapse by mean of their characteristic lipophilicty, hence, eliciting its central influence [179, 180]. Thus, (Gant, et al. 1987) have postulated that brain GABA receptors is its major site for electing its tremorgenic influence [179]. As compared to lolitrem B that elicit its maximum tremorgenic influence at 8 mg/kg dose [131], paxilline is considered as a weaker tremorgenic agent as it elicit an intermittent tremorgenic influence at 35 mg/kg while a sustained influence at 227 mg/kg intraperitoneal dose in murine model [181]. However, in sheep it exhibits extensive tremorgenic influence at 1.2 mg/kg body weight intravenous dose [182]. Through comparing both paxilline and lolitrem B pharmacological/toxic influence, it is obvious that a tiny structural difference modifies these mycotoxines binding properties to the calcium-activated BK channels as what is observed in wild and genetically modified mice models, yet, lolitrem B is still much more potent/longer acting blocker to these channels of motor functions than paxilline in vitro as well as in vivo [183-185]. Nevertheless, in contrast to lolitrem B, paxilline has more rapid onset/shorter duration of action [145]. In this context, brief tremorgenic influence at 4-80 mg/kg dose of complete inhibition BK/Maxi or hSlo Channel at 1 μ M concentration in mice, while, 70% channel blockage at 1.0 mg/kg dose and 10 nM concentration in sheep exploited as moderate to strong tremor 2 minutes after administration which disappear within an hour [132, 138, 140, 186]. Finally, it is necessary to not that *Lolium* species alkaloid biological influence is pH dependent as it has its impact on their chemical structure [187].

PHARMACOKINETICS OF LOLINES AND INDOLE-DITERPENS ALKALOIDS:

Loline alkaloids are speculated to be retained intact in the blood after ingestion, however, in lambs it is reported that little fraction of loline is absorbed via passive diffusion mechanism while the majority of loline quantity is absorbed by other mechanism due to its molecule hydrophilicity, small molecular weight along with neutral charge, making this compound easily cleared out of the GIT mucosa.Hence, it could be an excellent potential anthelmintic agents of local GIT action for pharmaceutical investigation [107, 188,

189], although it is reported to have good bioavailability in the blood/gastric mucosa of horses and sheep next to oral intake [188, 190] as it is reported to be readily absorbed beside, rapidly excreted renally in hours as well as bovine models [87, 191, 192]. Loline congeners including loline base, N-acetylloline and N-formylloline can cross passively across all of the GIT cross-section tissues of epithelium particularly ileum that exhibit the maximum 5% capacity. However, the greatest detected level in the blood was of loline base followed by N-formylloline followed by Nacetylloline, while, solely loline metabolites is detected in the liver and kidney tissues in lambs model as it suffers rapid metabolism. Yet, only small amount of Nformylloline are detected in these two organs and blood as compared to loline base metabolites availably in abundance [188] while some N-acetylloline and N-formylloline are located in hours blood [190]. Interestingly, (Seawright, et al. 1991) have reported that these pyrrolizidine alkaloids metabolites bind to the hemoglobin's globulin thiol groups of the hours's red blood cells [193].

In addition, these four loline alkaloids are renally excreted 2 hrs next to dosing in lamb model in addition to the metabolites of loline base as well as N-formylloline [194]. In this context, (Froehlich, 2020) have speculated that Nformylloline is the active form of loline metabolites while the simple loline base is un effective due to its rapid metabolism to an un effective metabolites while parasites counteracting N-formylloline metabolite is of good oral bioavailability while of poor urine excretion, thus remains in the blood for several hours [188, 194]. Some have supposed that loline alkaloids are absorbed, metabolized and excreted quickly, hence, exerts no symptoms of poisoning [194, 195]. Some of the loline alkaloids are metabolized in the intestinal mucosa in sheep, while, loline alkaloids are detected in urine in cow urine where over 50% of the absorbed loline is renally excreted followed by Nformylloline (bout 20%) followed by N-methylloline in sheep model meaning that their renal excretion is fast process occur within 15 minutes post dosing along with slow metabolism [194]. However, N-formylloline and Nacetylloline are excreted in hair as reported in hours model [196]. Nevertheless, (Ruan, et al.) have reported that the metabolites of these unsaturated pyrrolizidine alkaloids, particularly the platynecine type, are readily excreted without any binding to renal tissues protein adducts [128].

The Lolium species indole-diterpene, Loliterm B, is a lipophilic molecule insoluble in water, however, after oral administration, it has been reported to exhibit poor oral bioavailability due to poor GIT absorption [146, 197]. However, unlike, paxilline which is detected in murine brains at very low levels, high intravenous (75 μ g/kg BW) dose has exploited fast clearance from the systemic circulation despite the observed long term termorgenic influence in sheep model [145] as well as in lactating goat treated with (23 μ g/kg BW) dose [197]. It is hypothesized that loliterm B is trapped in certain body compartments then gradually released to the systemic circulation to find its way to the brain. This hypothesis is explained by the rapid blood clearance along with long term tremorgenic influence and encouraged by its detection in goat milk 32 hours and 75 hours post 23 μ g/kg BW IV dosing of 3% excretion rate and 100 μ g/kg BW oral dosing of slower excretion rate respectively [197, 145]. Remarkably, a resembling long term detection of loliterm B in bovine milk is reported by (Finch, et al. 2013) [146]. Finally, both paxilline and loliterm is metabolically oxidized hepatically into a detoxification more polar metabolites excreted billary [198].

TOXICOLOGY OF LOLIUM SPECIES AND THEIR ALKALOIDS

Human as well as animal toxicities happens in certain instances due to food, medicine and herbal products contaminated with plant toxins particularly pyrrolizidine alkaloids found in 3% of the flowering plants including grass that, regardless their long term consequences on health, have bring about fatalities in animals and human globally. Several poisoning cases have been encountered in case of using herbal preparations and teas of these alkaloids [26, 49]. Most of these alkaloids are hepatotoxic or even carcinogens; however, others are non-toxic or targets organs other than liver by their toxicity [26, 199]. Lolium species toxicity mostly known as ryegrass toxicity of often resembling causative toxin, clinical outcomes, case development as well as toxicological mechanism. In mammalians including humans and cattle the most common clinical manifestation include neurological toxicity expressed as tremor, diarrhea, loss of appetite, endocrinal outcomes expressed by reduction of milk production, and late manifestations including jaundice as well as photosensitivity. The most characteristic clinical manifestation, tremor happens via blocking the CNS inhibitory pathways through allosteric binding to GABA receptor chloride channel as well as chloride and calcium channel. However, despite no marked histological findings of their toxicities, yet, signs of adipose stores loss as well as emaciation are reported [26]. Several determinant factors related to the enzymatic as well as structural aspects lies behind these compounds' toxicity. The structural factors is related to their necine core basic or acidic structure and their substitutions nature/position mostly the ester one which are non-toxic unless they are bio-transformed into active metabolites such as pyrrolic esters [200, 201]. However, their bio-activation is hepatically performed, rather than detoxification, via consecutive series of oxidation-reduction as well as conjugation reactions [201-203] although chemically different metabolites are obtained from the metabolism of different classes of these pyrrolizidines alkaloids. The detoxification of the two classes retronecine and otenecine which are with single unsaturation at C1, C2 positions of the necine basic ring leads to their oxidative activation via cytochrome P450 hepatic enzymes into extensively reactive unstable pyrrolic esters (dehydropyrrolizidine) hepatotoxins that interact with thiol group of essential intracellular biomolecules such as enzymes or other proteins forming toxic pyrrole-protein adducts that targets the liver, lung besides other organs even acting as carcinogens [9, 128, 204]. However, these pyrrole active metabolites if interact with DNA may lead to genotoxicity bringing about carcinogencity [128]. In addition, if these reactive metabolites can be endogenously detoxified via binding to endogenous glutathione forming glutathione-pyrrole inactive water soluble adduct that us easily excreted [200, 201, 204, 128] on one hand. On the other hand, platynecines class of the pyrrolizidines alkaloids, which are of saturated necine bases nucleus are not hepatotoxic compounds as the previously mentioned classes [201, 202] although they pass a resembling oxidative hepatic metabolic fate via cytochrome P450 enzymes, yet, the resulted pyrrolic esters formed (dehydropyrrolizidine) is stable, un reactive, water soluble carboxylic acid that can not undergo conjugation reaction with thiol groups due to the absence of the necine base unsaturation, thus, needs no glutathione for their excretion [200]. The proposed metabolic pathway of *Lolium* species pyrrolizidine alkaloids is illustrated in figure (4) adapted from (Ruan et al. 2014) report [200]. However, (Stegelmeier, et al. 2013) consider pyrrolizidine alkaloids are generally non-toxic while, some of them are of toxicities other than hepatic one. In fact, (Stegelmeier, et al. 2013) have reported that although N-oxidation hepatic detoxification reaction of pyrrolizidine alkaloids along with increasing its solubility, these N-oxide metabolites are readily reduced in the GIT to re-establish their toxicity [26].

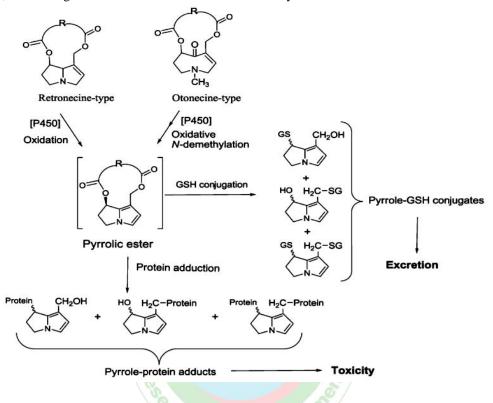


Figure (4): Metabolism of pyrrolizidine alkaloids proposed by (Ruan et al. 2014)^[200].

In general loline alkaloids are with no necine ring α_{β} unsaturation, with C2 and C7 oxygen bridge or without C1 or C7 ester substitution thus they lack the common pyrrolizidine alkaloids cytotoxicity [9, 65, 205], nevertheless, unlike oral administration, IV administration of loline alkaloids do exert toxic influences on mammalians [205]. While, other authors considers loline alkaloids have no anti-mammalian influence exhibited by other pyrrolizidine alkaloids particularly those related to genotoxicities [65-67] although liver cytochrome P450 oxidative enzymes have the ability of production of different genetic materials cross linking active pyrrolizidine alkaloids metabolites [66]. Generally, dimethylated or 1amino functionality aceylated loline alkaloids are not/ nonconclusively mammalian poisoning substances [1, 48]. Remarkably, in mice models loline alkaloids have been reported to exploit immunosuppressive as well as anorexic influence in rodents model [107, 205]. In addition, (Wang, et al. 2019) have reported that pyrrolyzidine alkaloids is also detected in honey [206], while, other reported their detection in bovine milk contributing to toxic influence against lactating infants although in both cases their concentration bellow the toxic thresholds of adults indicating low toxicity to humans consuming animal meat,

fats and products as reported in Germany [142, 146, 207]. However, number of reports have been issued regarding the incidence of several human poisoning cases due to ingestion of wheat products contaminated with the L. temultentum seeds [208, 209]. In this context, pure loline dihydrochloride have exhibited no toxicity up to 200 mg/kg body weight dose administered intraperitoneally, lethal at 400 mg/kg IV dose while nontoxic at oral 100mg/kg dose in murine model [14, 17, 108]. Besides, it has no interaction α_1 -adrenergic with receptor [108]. similarly with serotonergic [109]. receptors cholinergic nicotinic/muscarinic receptors [110, 111] or benzodiazepine receptors in calf brain [112], thus, L. temulentum or L. arundinaceum, is likely no associated with loline alkaloid as earlier believed [1]. Probably, the poisonous influence of L. temulentumseeds is exhibited by a mixture of its loline alkaloids, loline, lolinine, lolinidine, temuline and temulentine [16, 209] although (Bush, et al. 1983) have reported that loline analogues, N-formylloline, Nacetylloline, N-methylloline, norloline, N-acetylnofloline and N-formylnofloline are not hepatotoxines, although they may promote the biological membranes penetration as well as toxicity of other Lolium species toxins like ergot alkaloids, hence, exploiting indirect toxicity in large excess

concentration [9]. Nevertheless, other authors have reported that lolines including N-acetyl loline, N-acetyl norloline, Nmethyl loline, and loline base are generally not toxic to human beings or other mammalians as compared to other infected lolium alkaloids like ergot alkaloids [1, 9, 192, 194], despite, some have believed that N-formylloline and N-acetylloline are involved in equine fescue oedema [190, 196]. In general, lolines obtained from leaves, stems and head stems of L. temulentum are less toxic than nicotine in animal model [210]. However, in certain reports toxic/ lethal doses of loline alkaloids are specified. For example, the lethal dose of festucine (loline base) is 400 mg/Kg when administered IV, while, it is safe up to 1000 mg/kg oral dose [14] as what is encountered in murine models [17, 211, 212]. In addition, daily oral administration of Nformylloline, N-acetyl loline, N-acetyl norloline, N-methyl loline, and loline base mixture in a dose of 415 mg/kg has exhibited no obvious pathological, histological, hematological influence besides, no apparent influences on heart rate, blood pressure or motor coordination in murine model, although anorexia influence as well as cessation of weight gain have been reported particularly for Nformylloline [205, 212]. Nevertheless, (Jackson, et al. 1996) have correlated between N-formylloline and growthstimulating factor for its anorexic influence although they have reported that it has no influence on testes, hypothalamus and corpus striatum mass or on, prolactin and alkaline phosphatase levels [205]. Furthermore, as a dominant component (45.46%) of the in the total alkaloids (2.7%) seeds alcoholic extract of Lolium temulentum, loline has been reported to exhibit acute toxicity in rodents (mice and rats) model post oral as well as intraperitoneal dosing manifested as CNS depression that is deteriorated to coma then death due to respiratory failure. Yet, lethality of the total alkaloids toxicosis is greater in mice model by 1.58 and 1.35 for the two routs respectively while, lethality after oral intraperitoneal rout is 30 folds greater than oral routs mostly due to the total alkaloids neuroleptic influence, rather than loline alone, that starts to affect animals behavior, without interfering the learning capability in a dose dependent manner. Remarkably, at doses of 280 and 440 mg/kg doses loline lacks fatal acute toxicities in mice and rat models respectively [40]. It is reported that like classical ergotism, N-Acetylloline in fescue exploits its pituitary gland directed prolactin release inhibition, reproductive abnormality issues, hyperthermia and dry gangrene of extremities due to vasoconstriction inducing influence toxicity characteristics in animal models [32, 117]. Moreover, in Pakistan, Lolium temulentum L seeds consuming toxicity are rarely lethal to humans, nevertheless, the toxicity characteristics of diarrhea, gastroenteritis, vomiting, ataxia, nausea. giddiness, apathy and mydriasis are reported to be attributed to Cynoide as well as loline alkaloids like temuline and loliine [21]. Interestingly, toxicity case of endophyte-infected tall fescue, containing 1610 pg/g (N-formylloline and Nacetylloline) combination in addition to 0.39 μ g/g of ergovaline plus ergovalinine combination, ingestion by pregnant mares have caused teratogencity so that only 3 of 11 fetus have been delivered alive while solely one of them passed the natal stage to lactate despite the two classes of alkaloids are bellow the toxic concentration [58]. However, other loline metabolites like loliline and Lolitriol are also

exhibit nontremorgenic toxicity at doses of 8 mg/kg and 20 mg/kg respectively in murine model [155] on one hand. On the other hand, lolitriol have been reported exhibit its influence via targeting BK/Maxi or hSlo ChannelIC₅₀ = 196nM as compared to IC₅₀ of 43 nM of loliterm B [139, 140]. However, the indole-diterpene alkaloid of Lolium species, paxilline has its tremorgenic toxicity on vertebrate that need further investigations for its neurological effect on K⁺ channels [90, 213], as it is metabolically converted in vivo into the other indole-diterpene alkaloid, loliterm B that mediate most of the neural transmission disturbance fescue poisoning symptoms attributed to the alkaloid-promoted thiamine deficiency in animal model [9]. In this context, (Miles, et al. 1992) have reported that Lolitriol plus β -Paxitrol (16 mg/kg and 100 mg/kg) combination have exhibited lethal toxicosis at doses of 200 μ L dosage post initial lethargy period [140]. As compared to loliterm B, paxilline is a weaker termorgenic toxine [129]. In fact, loliterm B, is considered the most toxic indole-diterpene alkaloid of Lolium species, particularly in perennial ryegrass, that primarily contribute to their motor/neurological (tremorgenic) toxicities [129, 157] as well as in any other plant seeds containing loliterm B [157, 214], via binding to BK channels, however, the duration/severity of its toxicity beside its excitatory influences are location dependent. In addition, these influences/toxicity are also dose, lipophilic character and metabolic fate dependent [215]. Moreover, (Craig, et al. 2014) have reported that loliterm B exhibits its toxic effect at threshold level > 1.8 μ g/g dry weight of lolium plants in cattle [216], while, sever/prolonged tremor toxicity is exhibited by loliterm B, at dose range of 0.5-8.0 mg/kg due to inhibition of BK/Maxi or hSloChannel with IC₅₀ of 4 nM at 70 µg/kg dose [131, 135, 185]. Furthermore, both of alkaloids including ergovaline ergot of antivertibrate/invertebrate toxicity and loliterms of solely antivertibrate toxicity are the active poisonous alkaloids responsible for endophytes infected Lolium species, in which they are detected, toxicities including *L.perenne* [5, 13] to which most of toxicity cases in mammalians are reported [100, 217-219]. In this context, both of Lolitrem B and ergovaline are responsible these grasses toxicity when available in concentrations of 1.8 μ g/g and 0.3 μ g/g dry weight of the plants [89, 91, 220]. Yet, trace amount of ergot alkaloids including Ergovaline and related ergopeptines in tall fescue is also associated with its toxicity, characterized by reduce weight, hypethermia, blood flow restriction, reduced reproduction/milk production [1, 150]. In addition, like loline alkaloids, peramine is of weak toxicological characteristics against mammalians [215, 221]. In this context, peramine hydrochloride, at oral dose of 1000 mg/kg body weight exhibit highest toxicity level in murine model manifested by sluggish motor activity along with acute liver damage as revealed by autopsy, while, not influencing food intake, behavior, growth rate at dose of 50 μ g/g [221]. In murine model, infected ryegrass seeds consuming CNS toxicity manifestations involves hyperexcitability as well as nervousness [222]

Regarding the Lolium plants toxicities like perennial ryegrass (*Lolium perenne*) motor/neurological toxicity,

characterized by tetanic muscle spasms that leads to severe incoordination as well as hypersensitivity to external stimuli, is to be a reversible case in animals in Australia and North America [26, 223], however, no human toxicities are reported. While, toxicity of annual ryegrass (Lolium rigidum Gaud.) can be lethal manifested as neurological disorders as commonly reported in in Australia, South Africa, but, rarely reported in North America [26]. In Germany, symptoms of stiffness and movement disorders are reported in horses ingested perennial ryegrass due to ergot alkaloid [224]. L. temulentum seeds of the loline alkaloid temuline, toxicity is manifested as CNS as well as GIT symptoms [68, 225]. Other fescue poisoning manifestations involve interference with energy metabolic processes due to thiamine deficiency [226], interference with brain/hypothalamic functions including, gammaaminobutyrate (GABA), glutamine serotonergic and melatonin pathways [227]. However, (Watt and Breyer-Brandwijk, et al. 1962) have reported that lolium plants human intoxication symptoms are mutually similar to alcoholic sedation characterized with headache, dizziness, vertigo, mental confusion, difficulty in speech, inability to walk, vomiting, hypothermia and generalized shivering although a decoction of these plants is traditionally used in Moroccans folokloric medicine for haemorrhage and urine incontinence. In addition, other traditional medicine use of the powdered plant seeds for suppress the psychological and vasomotor disturbances associated with menopause when taken orally besides, being used topically for various skin disorders [209].

CONCLUSIONS:

The genus Lolium belonging to the family Poaceae or Gramineae involves around seven species, of poisonous grass plants grown globally particularly in Asia in corps especially wheat fields, however, in Iraq are called "rewatta". Their toxicity is mostly related to their characteristics alkaloids, the pyrrolizidine; lolines, indolediterpenes (ergots, loliterms, and paxillines) as well as peramine alkaloids mostly concentrated in their seeds, for which endophytes symbiosis/infection are involved in their synthesis particularly Acremonium, Neotyphodium or Epichloe species although these plants are capable of producing of amino-pyrrolizidine alkaloids like lolines individually without the need for fungal infection. These alkaloids are described to be contributing to both neuroand non neuro-toxicities. The levels of these alkaloids ranging from 0.2-1 mg/g inclines to their optimum levels in the areal parts as well as seeds during later summer and autumn reaching up to 10 folds in dry plant and constitute 45% of Lolium cuneatum Nevski of the total alkaloids (0.23%) of the chloroform extract. Chemically, the core nucleus of their pyrrolizidine alkaloids is necine composed of two fused saturated heterocyclic pentagonal rings with a nitrogen atom at one of the bridgehead with C1 amine group substitution which is characteristic to their loline alkaloids, including loline base, loline, Norloline, Nacetylloline, N-formylloine, N-acetylnorloline and Nformylnorloline. besides, a third exocyclic ring structure due to exocylic oxygen bridge occurs between C2 and C7. The difference between loline and norloline alkaloids is the existence of N-methyl group substitution at C1 amino

group while the acetyl, methyl as well as formyl derivatives of loline are results of C1 amino group acylation or alkylation. Nevertheless, other dimeric loline alkaloids like lolidine in addition to other tricyclic alkaloid perloline have been isolated from Lolium temulentum L. both of the indole-diterpene alkaloids paxilline and ergovaline are reported to be the precursor of the most toxic lolium species alkaloid loliterms including loliterm B of levels ranging 3-6 mg/g dry plant weight, which are biosynthesized with aid of endophytes symbiosis especially in the perennial ryegrass. In general, lolium species loline alkaloids are considered as relatively polar molecules as compared to the other lipophilic indole-diterpene alkaloids paxillines, loliterms and ergovaline which are the actual indicators of these plants toxicity. In some mammalians loline alkaloids are of poor oral bioavailability due to limited passive absorption although it is spectualted to remain unchanged within circulation, local systemic thus, intra-lumen pharmacological influences of these alkaloids are expected while in horses for example loline base, N-acetylloline and N-formylloline have exhibited good oral bioavailability potentially due to absorption mechanisms other than passive one particularly along with rapid renal excretion as in bovines. Only loline metabolites are detected in mammalian liver and kidney tissues as they suffer extensive/rapid metabolism, hence, loline base level > Nacetylloline > N-formylloline in their blood blinding to hemoglobin SH groups. For such kinetics the solely systemically active form of loline alkaloids is Nformylloline due to poor renal excretion, while, loline base is inactive due to fast first pass metabolism as well as rapid renal excretion since bovines for example excrete 50% of the absorbed loline renally. However, the indole-diterpene alkaloids have poor GIT absorption due to extensive lipophilicity as well as poor brain tissues accumulation in murine model due to rapid clearance from the systemic circulation through entrapment in body fat depots that may contributes to their prolonged influence. In general, this type of lolium alkaloids are detoxified through metabolic N-oxidation to a more polar metabolites excreted through the biliary rout. Regardless some reported traditional uses from Africa, the loline alkaloids of these plants have been reported to exploit diverse neuronal/motor as well as nonneurological influences. The neurological influences primarily demonstrated as a depressive activity, exhibited via affecting the central nervous system through interacting the α 1-adrenergic, α 2-adrenergic, D_2 dopamine, cholinergic, serotonergic and benzodiazepine receptors in the hypothalamic and cerebral cortex regions leading to tremorgenic as well as anorexic influences. Nevertheless, along with these neurological effects they affect the pituitary function leading to decline prolactin production. In the brain loline alkaloids particularly targets the dopaminergic receptrs. However, peripherally, they have influenced the α_2 -adrenergic, D₂ dopamine, or serotonergic receptors in the blood vasculatures smooth muscles leading to vasoconstriction and blood vessels thickening. While, in rabbits, cats and doges loline causes hypotension that could be accompanied by cardiac arrest at diastole due to its negative inotropic effect along with declining coronary blood flow. Moreover, loline alkaloids exhibits a remarkable immunosuppressive influence in murine model.

N-acetylloline, N-formylloline and N-methylloline have no central pituitary influences as they don't target brain's dopaminergic receptors. While, in excessive dose these loline alkaloids affects the energy production via affecting thiamine metabolism leading to thiamine deficiency. GI influences are also reported for lolines including increasing intestinal smooth muscles tone and contractility leading to diarrhea via promoting acetylcholine release and along with blocking histaminergic receptors. Moreover, both loline alkaloids as well as ergot alkaloids causes miscarriage as well as teratogenic influences in horses that causes 90% mortalities in fetuses. Although, semisynthetic derivatives of 8-12 carbon acyl chain substituting C1 amino group have significant antineoplastic influences against solid tumor in brine-shrimp model against human lung carcinoma A-549, breast carcinoma MCF-7, and colon adenocarcinoma HT-29. Similar CNS directed tremorgenic as well as GIT directed stimulatory influences have been reported to the indoel-diterpene alkaloids paxilline and loliterm B that could fatal for loliterm overdoses exhibited through high conductance calcium-activated BKca channel blockade in addition to similar endocrinal as well as peripheral influences. In the context of lolium alkaloids toxicity, these with C1 and C2 necine nucleus unsaturation are with hepatotoxicity, genotoxic as well as carcinogenic as they are activated metabolically through oxidation into a very reactive N-oxide metabolites, while, those with no unsaturation are not. However, most of lolines are of saturated necine nucleus thus they share with other indolediterpene alkaloids particularly loliterm B and paxilline toxicity, diarrhea, endocrinal as well CNS as photosensitivity through allosteric binding to GABA receptor chloride channel as well as chloride and calcium channel determined by structural aspects particularly the their necine core basic or acidic structure and their substitutions nature/position for loline alkaloids. Finally, toxic influences of lolium alkaloids are function of their biological influences mostly exhibited via resembling molecular mechansims centrally as well as peripherally. Unfortunately, an extensively little is reported regarding their pharmacological, toxicological as well as kinetics in humans despite many spectulations for pharmacological benifits, some traditional uses as well as some biological activities of their acyl derivitives are reported which requires future investigations.

REFERENCE:

- Schardl CL, Grossman RB, Nagabhyru P, Faulkner JR, Mallik UP. Loline alkaloids: currencies of mutualism. Phytochemistry. 2007; 68(7): 980-996.
- Vikuk V, Fuchs B, Krischke M, Mueller M J, Rueb S, Krauss J. Alkaloid concentrations of *Lolium perenne* infected with *Epichloë festucae* var. lolii with different detection methods—A re-evaluation of intoxication risk in Germany?. Journal of Fungi. 2020; 6(3): 177.
- Reddy P, Deseo MA, Ezernieks V, Guthridge K, Spangenberg G, Rochfort S. Toxic indole diterpenes from endophyte-infected perennial ryegrass *Lolium perenne* L.: Isolation and stability. Toxins. 2019; 11:16.
- Blankenship JD. (2004). Loline alkaloid biosynthesis in *Neotyphodium uncinatum*, a fungal endophyte of *Lolium pratense*. Doctor of Philosophy dissertation, College of Agriculture at the University of Kentucky.
- Arechavaleta M, Bacon CW, Hoveland CS, Radcliffe DE. Effect of the tall fescue endophyte on plant response to environmental stress. Agronomy Journal. 1989; 81: 83-90.

- Leuchtmann A, Schmidt D, Bush L P. Different levels of protective alkaloids in grasses with stroma forming and seed-transmitted *Epichloe/Neotyphodium* endophytes. Journal of Chemical Ecology. 2000; 26:1025-1036.
- Siegel MR, Latch GCM, Bush LP, Fannin FF, Rowan DD, Tapper B A, Bacon C W, Johnson M C. Fungal endophyte-infected grasses: alkaloid accumulation and aphid response. Journal of Chemical Ecology. 1990; 16:3301-15.
- Craven KD, Blankenship JD, Leuchtmann A, Hignight K, Schardl CL. Hybrid fungal endophytes symbiotic with the grass *Lolium pratense*. Sydowia. 2001; 53:44-73.
- Bush LP, Fannin FF, Siegel M R, Dahlman DL, Burton HR. Chemistry, occurrence and biological effects of saturated pyrrolizidine alkaloids associated with endophyte-grass interactions. Agriculture, Ecosystems & Environment. 1993; 44(4-1): 81-102.
- Justus M, Witte L, Hartmann T. Levels and tissue distribution of loline alkaloids in endophyte-infected *Festuca pratensis*. Phytochemistry. 1997; 44:51-57.
- Blakemore P R, Kim S-K, Schulze V K, White J D, Yokochi A F T. Asymmetric synthesis of (+)-loline, a pyrrolizidine alkaloid from rye grass and tall fescue. J. Chem. Soc., Perkin Trans. 2001; 1 :1831– 1847.
- Blankenship JD, Spiering MJ, Wilkinson HH, Fannin FF, Bush LP, Schardl CL. Production of loline alkaloids by the grass endophyte, *Neotyphodium uncinatum*, in defined media. Phytochemistry. 2001; 58 :395-401.
- Bacon CW. Abiotic stress tolerances, moisture, nutrients, and photosynthesis in endophyte-infected tall fescue. Agriculture Ecosystems & Environment. 1993; 44:123-141.
- 14. Yates SG, Tookey HL, Festucine, an alkaloid from tall fescue (*Festuca arundinacea* Schreb.): chemistry of the functional groups. Aust. J. Chem. 1965; 18(1): 53–60.
- 15. TePaske MR, Powell RG, Clement SL. Analyses of selected endophyte-infected grasses for the presence of loline-type and ergottype alkaloids. Journal of Agricultural and Food Chemistry. 1993; 41 :2299-2303.
- Yunusov SY, Akramov ST. Alkaloids of seeds of *Lolium cuneatum*. Zhurnal Obshchei Khimii. 1955; 25: 1765-71.
- Dannhardt G, Steindl L. Alkaloids of *Lolium temulentum*: isolation, identification and pharmacological activity. Planta Medica. 1985; 51(3):212-214.
- Bush LP, Cornelius PL, Buckner RC, Varney SR, Chapman RA, et al. Association of N-acetyl loline and N-formylloline with *Epichloe typhina* in tall fescue. Crop Sci. 1982; 22:941-943.
- Jones TA, Buckner RC, Burrus II, PB, Bush LP. Accumulation of pyrrolizidine alkaloids in benomyl-treated tall fescue parents and their untreated progenies. Crop Sci. 1983; 23: 1135-1140.
- Yu SY, Akramov ST. Alkaloids of seeds of Lolium cuneatum. Zh. Obshch. Khim.1955; 25 :1813-1820.
- Khan RU. Mehmood S. Khan S U. Toxic effect of common poisonous plants of district Bannu, Khyber Pakhtunkhwa, Pakistan Pakistan Journal of Pharmaceutical Sciences. 2018; 31(1):57-67.
- 22. Holm LG, Plucknett DL, Pancho JV, Herberger JP. (1977) The World's worst weeds. University Press of Hawaii, Honolulu
- Senda T, Tominaga T. Genetic diversity of darnel (Lolium temulentum L.) in Malo, Ethiopia depends on traditional farming systems. Econ Bot. 2004; 58:568–577.
- 24. Tackholm V. Students Flora of Egypt. Anglo-Egyptian Bookshop Cairo. 1965.
- Tackholm V, Drar M, Täckholm G. *Flora of Egypt* (Vol. 1, pp. 113-114). Giza, Egypt: Fouad I University. 1941.
- Stegelmeier BL, Field R, Panter KE, Hall JO, Welch KD, Pfister JA. (2013). Selected poisonous plants affecting animal and human health. In *Haschek and Rousseaux's Handbook of Toxicologic Pathology* (pp. 1259-1314). Academic Press.
- 27. Thomas H, Archer E., Turely R. M., (2011), Evolution, Physiology and Phytochemistry of the Psychotoxic Arable Mimic Weed Darnel (Lolium temulentum L.). In Progress in Botany, genetics physiology, systematic ecology. Volume 72. Editors: Lu^{*}ttge U, Beyschlag W, Bu^{*}del B, Francis D. (pp. 76, 80) Springer-Verlag Berlin Heidelberg.
- Yates SG, Fenster JC, Bartelt RJ. Assay of tall fescue seed extracts, fractions, and alkaloids using the large milkweed bug. J. Agric. Food Chem. 1989; 37:354–357.
- Miles CO, Di Menna ME, Jacobs SWL, Garthwaite I, Lane GA, Prestidge RA, Marshall SL, et al. Endophytic fungi in indigenous Australasian grasses associated with toxicity to livestock. Appl. Environ. Microbiol. 1998; 64 :601–606.

- Siegel, MR, Latch, GCM., Bush, LP, Fannin, FF, et al. Fungal endophyte infected grasses: alkaloid accumulation and aphid response. J. Chem. Ecol. 1990; 16:3301–3315.
- Robbins JD, Sweeny JG, Wilkinson SR, Burdick D. Volatile alkaloids of Kentucky 31 tall fescue seed (Festuca arundinancea). Journal of Agricultural and Food Chemistry. 1972; 20(5):1040-1043.
- 32. Hemken RW. Bush LP. Toxicants of Plant Origin, Vol. I, Cheeke PR, Ed., CRC Press, Boca Raton, Florida, 1989, pp. 281-289.
- 33. Powell RG, Petroski RJ. The loline group of pyrrolizidine alkaloids. Alkaloids: Chemical and Biological Perspectives. 1992; 8 :320-38. Powell RG, Petroski RJ. (1992). The loline group of pyrrolizidine alkaloids. In *Alkaloids: chemical and biological perspectives* (pp. 320-338). Springer, New York, NY.
- Porter JK. Analysis of endophyte toxins Fescue and other grasses toxic to livestock. Journal of Animal Science. 1995; 73:871-880.
- Bacon CW. Procedure for isolating the endophyte of tall fescue and screening isolates for ergot alkaloids. Applied and Environmental Microbiology. 1988; 54:2615-2618.
- Rowan DD. Lolitrems, perimine, and paxilline--mycotoxins of the ryegrass-endophyte interaction. Agriculture Ecosystems & Environment. 1993; 44:103-122.
- Gurney KA, Mantle PG, Penn J, Garthwaite I, Towers NR. Loss of toxic metabolites from *Acremonium lolii*, the endophyte of ryegrass, following mutagenesis. Naturwissenschaften. 1994; 81:362-365.
- Porter JK. (ed.) 1994. Chemical constituents of grass endophytes, pp. 103-123. CRC, Boca Raton, FL.
- Bush LP, Wilkinson HH, Schardl CL. Bioprotective alkaloids of grass-fungal endophyte symbioses. Plant Physiology. 1997; 114:1-7.
- Petroski RJ, Yates SG, Weisleder D, Powell RG. Isolation, semisynthesis, and NMR spectral studies of loline alkaloids. Journal of Natural Products. 1989; 52(4):810-817.
- Schardl CL, Young CA, Faulkner JR, Florea S, Pan J. Chemotypic diversity of Epichloë, fungal symbionts of grasses. Fungal Ecol. 2012; 5:331–344.
- 42. Burhan W. 1984. Development of Acremonium coenophialum and accumulation of N-acetyl and N-formyl loline in tall fescue (*Festuca arundinacea* Schreb.). Master's Thesis, University of Kentucky, Lexington, KY, p. 64.
- Belesky DP, Robbins JD, Stuedemann JA, Wilkinson SR, Devine OJ. Fungal endophyte infection-loline derivative alkaloid concentration of grazed tall fescue. Agron. J. 1987; 79:217-220.
- 44. Buckner RC, Bush LP, Burrus II, PB. 1981. Improvement of forage quality of tall rescue through *Lolium-Festuca* hybridization. In: Smith JA, Hays VW (Editors), Proc. 14th Int. Grassland Congress 15-24 June, 1981, Lexington, KY, Westview Press, Boulder, CO, pp.157-159.
- 45. Belesky DP, Stringer WC, Plattner RD. Influence of endophyte and water regime upon tall fescue accessions. II. Pyrrolizidine and ergopeptine alkaloids. Ann. Bot. 1989; 64: 343-349.
- 46. Liu G, Casqueiro J, Banuelos O, Cardoza RE, Gutierrez S, Martin JF. Targeted inactivation of the mecB gene, encoding cystathioninegamma-lyase, shows that the reverse transsulfuration pathway is required for high level cephalosporin biosynthesis in *Acremonium chrysogenum* C10 by not for methionine induction of the cephalosporin genes. Journal of Bacteriology. 2001; 183 :1765-1772.
- Boettcher F, Ober D, Hartmann T. Biosynthesis of pyrrolizidine alkaloids: putrescine and spermidine are essential substrates of enzymatic homospermidine formation. Canadian Journal of Chemistry. 1994; 72:80-5.
- Siegel MR. Bush LP. (eds.).Toxin production in grass/endophyte associations. Springer-Verlag, New York. 1997; Vol.5, pp. 185-207.
- Smith LS. Culvenor CCJ. Plant sources of hepatotoxic pyrrolizidine alkaloids. J. Nat. Prod. 1981; 44: 129-152.
- Kennedy CW, Bush LP, Effect of environmental and management factors on the accumulation of N-acetyl and N-formyl loline alkaloids in tall fescue. Crop Sci. 1983; 23:547-552.
- König J, Fuchs B, Krischke M, Mueller MJ, Krauss J. Hide and seek—Infection rates and alkaloid concentrations of Epichlö festucae var. lolii in Lolium perenne along a land-use gradient in Germany. Grass Forage Sci. 2018; 73: 510–516.
- 52. Fuchs B, Krischke M, Mueller MJ, Krauss J. Plant age and seasonal timing determine endophyte growth and alkaloid biosynthesis. Fungal Ecol. 2017; 29:52–58.
- Jensen AMD. Endophyte persistence and toxin (lolitrem b) production in a Danish seed crop of perennial ryegrass. Eur. J. Agron. 2005; 23:68–78.
- 54. Repussard C, Zbib N, Tardieu D, Guerre P. Ergovaline and lolitrem B concentrations in perennial ryegrass in field culture in southern

France: Distribution in the plant and impact of climatic factors. J. Agric. Food Chem. 2014; 62:12707–12712.

- Bauer JI, Gross M, Cramer B, Humpf H-U, Hamscher G, Usleber E. Immunochemical analysis of paxilline and ergot alkaloid mycotoxins in grass seeds and plants. J. Agric. Food Chem. 2018; 66(1):315–322.
- Hartmann T. Chemical ecology of pyrrolizidine alkaloids. Planta. 1999; 207:483-495.
- 57. Stuedemann JA, Rumsey TS, Bond J, Wilkinson SR, Bush LP, Williams DJ, Caudle AB. Association of blood cholesterol with occurrence of fat necrosis in cows and tall fescue summer toxicosis in steers. Am. J. Vet. Res. 1985; 46:1990-1995.
- Putnam MR, Bransby DI, Schumacher J, Boosinger TR, et al. The effects of the fungal endophyte *Acremonium coenophialum* in fescue on pregnant mares and foal viability. Am. J. Vet. Res. 1991; 52:2071-2074.
- 59. Guglielmone AER, de Sanez AM, Carabelli MA, Guglielmoni MB, Basile EO, Severuga JO. Festucas toxicas e inocuas: diferencias en el contenido de alcaloides ysu relacion con un ensayo preliminar a campo. Rev. Asoc. Argent. Consorcios Regionales Experimentacion Agricola (CREA). 1981; 15:40-47.
- Cakmak M, Mayer P, Trauner D. An efficient synthesis of loline alkaloids. Nature chemistry. 2011; 3(7):543-545.
- Yates SG, Petroski RJ, Powell RG. Analysis of loline alkaloids in endophyte infected tall fescue by capillary gas chromatography. Journal of Agricultural and Food Chemistry. 1990; 38:182-5.
- Dougherty CT, Knapp FW, Bush LP, Maul JE, Van Willigen J. Mortality of horn fly (*Diptera: Muscidae*) larvae in bovine during supplemented with loline alkaloids from tall fescue. Journal of Medical Entomology. 1998; 35:798-803.
- 63. Patterson CG, Potter DA, Fannin FF. Feeding deterrency of alkaloids from endophyte-infected grasses to Japanese beetle grubs. Entomologia Experimentalis et Applicata. 1991; 61:285-9.
- 64. Wilkinson HH, Siegel MR, Blankenship JD, Mallory AC, Bush LP, Schardl CL. Contribution of fungal loline alkaloids to protection from aphids in a grass endophyte mutualism. Molecular Plant-Microbe Interactions. 2000; 13:1027-1033.
- 65. Hincks JR, Kim H-Y, Segall H J, Molyneux RJ, Stermitz FR, Coulombe Jr RA. DNA cross-linking in mammalian cells by pyrrolizidine alkaloids: Structure-activity relationships. Toxicology and Applied Pharmacology.1991; 111:90-98.
- 66. Kim H-Y, Stermitz FR, Li JK-K, Coulombe Jr RA. 1999. Comparative DNA Crosslinking by activated pyrrolizidine alkaloids. Food and Chemical Toxicology. 1999; 37:619-625.
- 67. Jones TA, Buckner R C, Burrus P B. Pyrrolizidine alkaloid levels in tall fescue seed as influenced by seed age, location, and variety. Journal of Seed Technology. 1983; 8: 47-54.
- 68. Hofmeister F. The active constituents of lolium temulentum. Arch. Exp. Pathol. Pharmakol. 1892; 30:203–230.
- Yunusov SY, Akramov ST. Investigation of the alkaloids of the seeds of *Lolium cuneatum* (Nevski). J. Gen. Chem. (Moscow). 1955; 25, 1765–1771.
- Katz, I., Contribution à l'étude de l'ivraie eniurante, Phytopathol. Z. 1949; 15:495.
- 71. Akrarnov S T, Yunusov S Y. Chern Nat Cornpd, Engl Transl. 1965; 1:203.
- Bates RB, Morehead SR. Absolute configurations of pyrrolizidine alkaloids of the loline group. Tetrahedron Lett. 1972; 17:1629-1630.
- Aasen AJ, Culvenor CCJ. Abnormally low vicinal coupling constants for O–CH–CH in a highly strained five-membered-ring ether; the identity of loline and festucine. Aust. J. Chem. 1969; 22:2021–2024.
- Yunusov SY, Akramov ST. Structure of norloline, loline, and lolinine. Doklady Akademii Nauk UzSSR. 1959; 24: 28-31.
- Yunusov SY, Akramov ST. Alkaloids of *Lolium cuneatum*. II. Zhurnal Obshchei Khimii. 1960; 30:677-82.
- Batirov EK, Khamidkhodzhaev SA, Malikov VM, Yunusov SY. Alkaloids of *LoLium cuneatum*. *Khimiya Prirodnykh Soedinenii*. 1976; 1:60-63.
- 77. Batirov EK, Malikov V M, Yunusov S Y. Khirn Prir Soedin. 1976; 1: 120.
- Batirov, E'K, Malikov VM, Yunusov SY, Alkaloids of the seeds of *Lolium cuneatum*. Chem. Nat. Prod. 1977; 12:114–115.
- Batirov EK, Malikov VM, Yunusov SY. Khirn Prir Soedin. 1976; 1: 63.
- Batirov E'K, Malikov VM, Yunusov SY, Lolidine a new chlorinecontaining alkaloid from the seeds of *Lolium cuneatum*. Chem. Nat. Prod. 1977; 12(1):52–54.
- 81. Yunusov SY, Akramov ST. Investigation of alkaloids of *Lolium cuneatum* II. J. Gen. Chem. (Moscow). 1960; 30: 699–704.

- Yunusov SY, Akramov ST, Structure of norloline, loline and lolinine IV. J. Gen. Chem. (Moscow). 1960; 30 :3105–3109.
- Gribble GW. Occurrence of halogenated alkaloids. The Alkaloids: Chemistry and Biology. 2012; 71:1-165.
- Hammouda FM. Rizk AM. El-Missiry MM. Ghaleb HA. Madkour MK. Pohland AE. Wood G. Poisonous plants contaminating edible ones and toxic substances in plant foods iv. phytochemistry and toxicity of Lolium temulentum. International Journal of Crude Drug Research. 1988; 26(4): 240-245.
- Pammel LH. Manual of Poisonous Plants. Iowa, The Torch Press, Cedar Rapids. 1911.
- Bull LB, Culvenor CCJ, Dick AT. The Pyrrolizidine Alkaloids. North Holland Publishing Company. 1969.
- Takeda A, Suzuki E, Kamei K, Nakata H. Detection and identification of loline and its analogues in horse urine. Chem. Pharm. Bull. 1991; 39:964–968.
- Tufariello JJ, Meckler H, Winzenberg K. Synthesis of the lolium alkaloids. *The Journal of Organic Chemistry*; 1986; 51(18):3556-3557.
- Hume DE, Ryan GD, Gibert A, Helander M, Mirlohi A, Sabzalian MR. Epichloë fungal endophytes for grassland ecosystems. In Sustainable Agriculture Reviews; Sustainable Agriculture Reviews; Lichtfouse E, Ed.; Springer International Publishing: Cham, Switzerland, 2016; Volume 19, pp. 233–305.
- Imlach WL, Finch SC, Dunlop J, Meredith AL, Aldrich RW, Dalziel JE. The molecular mechanism of "ryegrass staggers", a neurological disorder of K+ channels. J. Pharmacol. Exp. Ther. 2008; 327:657–664.
- 91. Di Menna M, Finch SC, Popay AJ, Smith BL. A review of the Neotyphodium lolii/Lolium perenne symbiosis and its associated effects on animal and plant health, with particular emphasis on ryegrass staggers. N. Z. Vet. J. 2012; 60:315–328.
- Young CA, Tapper BA, May K, Moon CD, Schardl CL, Scott B. Indole-diterpene biosynthetic capability of Epichloë endophytes as predicted by ltm gene analysis. Appl. Environ. Microbiol. 2009; 75:2200–2211.
- 93. Saikia S, Takemoto D, Tapper BA, Lane GA, Fraser K, Scott B. Functional analysis of an indole-diterpene gene cluster for lolitrem B biosynthesis in the grass endosymbiont. FEBS Lett. 2012; 586:2563–2569.
- 94. Schardl CL, Young CA, Hesse U, Amyotte S, Andreeva K, et al. Plant-symbiotic fungi as chemical engineers: multi-genome analysis of the Clavicipitaceae reveals dynamics of alkaloid loci. PLoS Genet. 2013; 9:1–26.
- Weedon CM, Mantle PG. Paxilline biosynthesis by Acremonium loliae; a step towards defining the origin of lolitrem neurotoxins. Phytochemistry 1987, 26:969–971.
- Adhikari KB, Boelt B, Fomsgaard IS. Identification and quantification of loline-type alkaloids in endophyte-infected grasses by LC-MS/MS. J. Agric. Food Chem. 2016; 64:6212–6218.
- Repussard C, Tardieu D, Alberich M, Guerre P. A new method for the determination of lolitrem B in plant materials. Anim. Feed Sci. Technol. 2014; 193:141–147.
- Cagaš B, Flieger M, Olšowská J. Concentration of ergot alkaloids in Czech ecotypes of Lolium perenne and Festuca pratensis. Grass Forage Sci. 1999; 54:365–370.
- 99. Garthwaite I, Miles CO, Towers NR. Immunological detection of the indole diterpenoid tremorgenic mycotoxins. In Proceedings of the Second International Symposium on Acremonium/ Grass Interactions; Hume DE, Latch GCM, Easton HS, Eds.; AgResearch: Palmerston North, New Zealand, 1993; pp.77–80.
- Davies E, Lane GA, Latch GCM, Tapper BA, et al. Alkaloid concentrations in field-grown synthetic perennial ryegrass endophyte associations. In Proceedings of the Second International Symposium on Acremonium/Grass Interactions; Hume DE, Latch GCM, Easton HS, Eds.; AgResearch: Palmerston North, New Zealand, 1993; pp.72–76.
- Lewis GC, Clements ROA survey of ryegrass endophyte (Acremonium loliae) in the U.K. and its apparent ineffectuality on a seedling pest. J. Agric. Sci. 1986; 107:633–638.
- 102. Oldenburg E. Endophytic fungi and alkaloid production in perennial ryegrass in Germany. Grass Forage Sci. 1997; 52:425–431.
- 103. Dapprich P, Paul VH, Krohn K. Incidence of Acremonium endophytes in selected German pastures and the contents of alkaloids in Lolium perenne. In The 2nd International Conference on Harmful and Beneficial Microorganisms in Grassland, Pastures and Turf, Paderborn, Germany; Krohn K, Paul VH. Eds.; IOBC/WPRS Bulletin: Avignon, France, 1996; Vol 19, pp.103–114.

- 104. Duke JA. Wild lettuce: a bitter herb of biblical proportions. J. Med. Food. 2000; 3: 153–154.
- 105. Strickland JR, Bailey EM, Abney LK, Oliver JW. Assessment of the mitogenic potential of the alkaloids produced by endophyte (*Acremonium coenophialum*)-infected tall fescue (*Festucaarundinacea*) on bovine vascular smooth muscle in vitro. J. Anim. Sci. 1996; 74:1664–1671.
- Larson BT, Samford MD, Camden JM, Piper EL, Kerley MS, Paterson JA, Turner JT. Ergovaline binding and activation of D₂ dopamine receptors in GH4ZR7 cells. J Animal Sci. 1995; 73: 1396-1400.
- 107. Dew RK, Boissonneault GA, Gay N, Boling JA, CrossRJ, Cohen DA. The effect of the endophyte (*Acremonium coenophialum*) and associated toxin(s) of tall fescue on serum titer response to immunization and spleen cell flow cytometry analysis and response to mitogens. Vet Immunnol Immunopathol. 1990; 26:285-295.
- Hornung R, Presek P, Glossmann H. Alpha adrenoceptors in rat brain: Direct identification with prazosin. Naunyn-Schmiedebergs Arch. Pharmakol. 1979; 308(3): 223-230.
- Peroutka SJ. Snyder SH. Multiple serotonin receptors: differential binding of [3H] 5-hydroxytryptamine,[3H] lysergic acid diethylamide and [3H] spiroperidol. *Molecular pharmacology*. 1979; 16(3):687-699.
- Yamamura HI, Snyder SH. Muscarinic cholinergic binding in rat brain. Proceedings of the National Academy of Sciences. 1974; 71(5):1725-1729.
- Lukasiewicz RJ, Bennett EL. α-Bungarotoxin binding properties of a central nervous system nicotinic acetylcholine receptor. *Biochimica et Biophysica Acta (BBA)-General Subjects*. 1978; 544(2):294-308.
- 112. Squires RF. Braestrup C. Benzodiazepine receptors in rat brain. *Nature*. 1977; 266(5604):732-734.
- 113. Strickland JR. Cross DL. Jenkins TC. Petroski RJ. Powell RG. The effect of alkaloids and seed extracts of endophyte-infected tall fescue on prolactin secretion in an in vitro rat pituitary perfusion system. J. Anim. Sci. 1992; 70:2779–2786.
- 114. Larson BT, Harmon DL Piper EL Griffis LM Bush LP. Alkaloid binding and activation of D_2 dopamine receptors in cell culture. J. Anim. Sci. 1999; 77:942–947.
- 115. Jackson JA, Hemken RW, Bush LP, Boling JA, Siegel MR, Zavos PM, Yates SG. Physiological responses in rats fed extracts of endophyte infected tall rescue seed. Drug Chem. Toxicol. 1987; 10:369-379.
- 116. Jackson JA, Yates SG, Powell RG, Hemken RW, Bush LP, Boling JA, Zavos PM, Siegel MR. Physiological responses in rats fed extracts of endophyte-free and endophyte-infected tall rescue seed relative to some known ergot alkaloids. Drug Chem. Toxicol. 1989; 12:147-164.
- 117. Strickland JR, Cross DL, Birrenkott GP, Grimes LW, Effect of ergovaline, loline, and dopamine antagonists on rat pituitary cell prolactin release in vitro. Am. J. Vet. Res. 1994; 55:716–721.
- Strickland J, Cross D, Jenkins T, Petroski R, PowellR, The effect of alkaloids and seed extracts of endophyte infect fescue on prolactin secretion in an in vitro rat pituitary perfusion system. *Journal of Animal Science*. 1992; 70:2779-2786.
- 119. Oliver JW, Strickland JR, Waller JC, Fribourg HA, Linnabary RD, Abney LK, Endophytic fungal toxin effect on adrenergic receptors in lateral saphenous veins (cranial branch) of cattle grazing tall fescue. J. Anim. Sci. 1998; 76:2853–2856.
- Solomons RN, Oliver JW, Linnabary RD, Reactivity of dorsal pedal vein of cattle to selected alkaloids associated with *Acremoniumcoenophialum*-infected fescue grass. Am. J. Vet. Res. 1989; 50:235–238.
- 121. Klotz J, Kirch B, Aiken G, Bush L, Strickland, J. Effects of selected combinations of tall fescue alkaloids on the vasoconstrictive capacity of fescue-naive bovine lateral saphenous veins. *Journal of Animal Science*. 2008; 86:1021-1028.
- Abney L, Oliver J, Reinemeyer C, Vasoconstrictive effects of tall fescue alkaloids on equine vasaculature. *Journal of Equine Veterinary Science*. 1993; 13(6):334-340.
- 123. Oliver J. Physiological manifestations of endophyte toxicosis in ruminant and laboratory species. In: Bacon C, Hill N, *Neotyphodium/Grass* Interactions. 1997; (pp. 311-346). New York: Plenum Press.
- Dougherty CT, Lauriault LM, Bradley NW, Gay N, Cornelius PL. Induction of tall fescue toxicosis in heat-stressed cattle and its alleviation with thiamin. J. Anim. Sci. 1991; 69:1008-1018.
- Edwin EE. Jackman R. Ruminant thiamin requirements in perspective. Vet. Res. Commun. 1982; 5:237-250.
- 126. Karimov VA, Kamilov IK, Pharmacology of the new loline alkaloid and of its derivative. Dok. Akad. Nauk Uzb. SSR. 1961; 12:43-47.

- Petroski RJ, Powell RG, Ratnayake S, McLaughlin JL. Note: Cytotoxic Activities of N-Acyllolines. International journal of pharmacognosy. 1994; 32(4):409-412.
- 128. Ruan J, Liao C, Ye Y, Lin G. Lack of metabolic activation and predominant formation of an excreted metabolite of nontoxic platynecine-type pyrrolizidine alkaloids. *ChemicalResearch in Toxicology*. 2014; 27:7-16.
- Rowan DD. Lolitrems, peramine and paxilline: mycotoxins of the ryegrass/endophyte interaction. Agriculture, Ecosystems & Environment. 1993; 44(1-4):103-122.
- Reddy P, Rochfort S, Read E, Deseo M, Jaehne E, Van Den Buuse M, et al. Tremorgenic effects and functional metabolomics analysis of lolitrem B and its biosynthetic intermediates. *Scientific reports*. 2019; 9(1):1-17.
- Gallagher RT, Hawkes AD. The potent tremorgenic neurotoxins lolitrem B and aflatrem: a comparison of the tremor response in mice. *Experientia*.1986;42:823–825.
- 132. McLeay LM, Mith BL, Munday-Finch SC. Tremorgenic mycotoxins paxilline, penitrem and lolitrem B, the non-tremorgenic 31-epilolitrem B and electromyographic activity of the reticulum and rumen of sheep. *Res. Vet. Sci.* 1999; 66:119–127.
- Combs MD, et al. Development of a model for investigation of perennial ryegrass toxicosis in sheep. N Z Vet J. 2018;66:281–289.
- Sallam AA, *et al.* Bioguided discovery and pharmacophore modeling of the mycotoxic indole diterpene alkaloids penitrems as breast cancer proliferation, migration, and invasion inhibitors. *Med Chem Comm*.2013; 4:1360–1369.
- Sallam AA,*et al.* Indole diterpene alkaloids as novel inhibitors of the Wnt/β-catenin pathway in breast cancer cells. *Eur J Med Chem.* 2013;70:594–606.
- Goda AA, *et al.* Astaxanthin and docosahexaenoic acid reverse the toxicity of the maxi-K (BK) channel antagonist mycotoxin penitrem A. *Mar Drugs.* 2016;14: 208.
- Johnstone LK, Mayhew IG, Fletcher LR. Clinical expression of lolitrem B (perennial ryegrass) intoxication in horses. *Equine Vet. J.* 2012; 44:304–309.
- 138. Knaus H-G. *et al.* Tremorgenic indole alkaloids potently inhibit smooth muscle high-conductance calcium-activated potassium channels. *Biochemistry*. 1994;33:5819–5828.
- Imlach WL, Finch SC, Dunlop J, Dalziel JE. Structural determinants of lolitrems for inhibition of BK large conductance Ca²⁺-activated K⁺ channels. *Eur. J. Pharmacol.* 2009; 605:36–45.
- 140. Miles CO, Wilkins AL, Gallagher RT, Hawkes AD, Munday SC, Towers NR, Synthesis and Tremorgenicity of Paxitriols and Lolitriol: Possible Biosynthetic Precursors of Lolitrem B. J. Agric. Food Chem. 1992; 40:234–238.
- 141. Cole RJ, Cox RH. Handbook of Toxic Fungal Metabolites (Academic Press, New York, NY). 1981; p.355.
- 142. Finch SC, Fletcher LR, Babu JV. The evaluation of endophyte toxin residues in sheep fat. N Z Vet J. 2011;60: 56–60.
- Miyazaki S, Ishizaki I, Ishizaka M, Kanbara T, Ishiguro-Takeda Y. Lolitrem B residue in fat tissues of cattle consuming endophyteinfected perennial ryegrass straw. J. Vet. Diagn. Invest. 2004; 16:340– 342.
- 144. Shimada N, et al. Toxicological evaluation and bioaccumulation potential of lolitrem B, endophyte mycotoxin in Japanese black steers. Food Addit Contam Part A Chem Anal Control Expo Risk Assess. 2013; 30:1402–1406.
- Munday-Finch SC, Garthwaite I. Toxicology of Ryegrass Endophyte in Livestock. Ryegrass Endophyte Essent. N. Z. Symbiosis Grassl. Res. Pract. Ser. 1999; 7:63–67.
- 146. Finch S, Thom E, Babu J, Hawkes A, Waugh C. The Evaluation of Fungal Endophyte Toxin Residues in Milk. N. Z. Vet. J. 2013; 61:11– 17.
- 147. Zbib N, Repussard C, Tardieu D, Priymenko N, Domange C, Guerre P. Toxicity of Endophyte-Infected Ryegrass Hay Containing High Ergovaline Level in Lactating Ewes. J. Anim. Sci. 2015; 93:4098–4109.
- Dalziel JE. Dunstan K E. Finch S C. Combined Effects of Fungal Alkaloids on Intestinal Motility in an in Vitro Rat Model. J. Anim. Sci. 2013; 91:5177–5182.
- Fletcher LR, Easton HS. The Evaluation and Use of Endophytes for Pasture Improvement. In Neotyphodium/Grass Interactions; Bacon CW. Hill NS. Eds.; Springer: New York, NY, USA, 1997; pp. 209– 227.
- 150. Bluett SJ, Thom ER, Clark DA, Macdonald KA, Minneé EMK. Effects of Perennial Ryegrass Infected with Either AR1 or Wild Endophyte on Dairy Production in the Waikato. N. Z. J. Agric. Res. 2005; 48:197–212.

- 151. Thom ER, Waugh CD, Minneé EMK. Dairy Cow Responses to the Wild-Type Endophyte in Perennial Ryegrass. In Proceedings of the 4th Australasian Dairy Science Symposium 2010, Hamilton, New Zealand, 2010; pp. 370–375.
- Ruckebusch Y, Malbert CH, Crichlow EC. Hexamethonium: A Probe to Assess Autonomic Nervous System Involvement in Upper Gastrointestinal Functions in Conscious Sheep. Vet. Res. Commun. 1987; 11:293–303.
- 153. Dalziel JE, Dunlop J, Finch SC, Wong SS. Immune Response Inhibition Using Indole Diterpene Compound. WO2006115423A1, 2006. Available online: https://patents.google.com/patent/WO2006115423A1/ko (accessed on 2 November 2018).
- 154. Betina V. Indole derived tremorgenic toxins. In: Betina V. (Editor), Mycotoxins Production, Isolation, Separation and Purification. Developments in Food Science, 8. Elsevier Scientific Publishing Company, Amsterdam/Oxford/New York, pp. 415-442.
- 155. Munday-Finch SC, Wilkins AL, Miles CO, Tomoda H, O'mura S. Isolation and Structure Elucidation of Lolilline, a Possible Biosynthetic Precursor of the Lolitrem Family of Tremorgenic Mycotoxins. J. Agric. Food Chem. 1997; 45:199–204.
- Wang L, Cross AL, Allen KL, Smith BL, McLeay LM. Tremorgenic Mycotoxins Increase Gastric Smooth Muscle Activity of Sheep Reticulum and Rumen in Vitro. Res. Vet. Sci. 2003; 74:93–100.
- Gallagher RT, White EP, Mortimer PH. Ryegrass Staggers: Isolation of Potent Neurotoxins Lolitrem A and Lolitrem B from Staggers-Producing Pastures. N. Z. Vet. J. 1981; 29:189–190.
- Easton HS, Lane GA, Tapper BA, Keogh RG, Cooper BM, Blackwell M, Fletcher LR. Ryegrass Endophyte-Related Heat Stress in Cattle. Proc. N. Z. Grassland Assoc. 1995; 57:37–41.
- 159. Auldist MJ, Thom ER. Effects of Endophyte Infection of Perennial Ryegrass on Somatic Cell Counts, Mammary Inflammation, and Milk Protein Composition in Grazing Dairy Cattle. N. Z. J. Agric. Res. 2000; 43:345–349.
- Hovermale JT, Craig AM. Correlation of Ergovaline and Lolitrem B Levels in Endophyte-Infected Perennial Ryegrass Lolium Perenne. J. Vet. Diagn. Investig, 2001; 13:323–327.
- 161. Benkhelil A, Grancher D, Giraud N, Bezille P, Bony S. Intoxication Par Des Toxines de Champignons Endophytes Chez Des Taureaux Reproducteurs. Rev. Méd. Vét. 2004; 156:243–247.
- 162. Zbib N, Repussard C, Tardieu D, Priymenko N, Domange C, Guerre P. Ergovaline in Tall Fescue and Its Effect on Health, Milk Quality, Biochemical Parameters, Oxidative Status, and Drug Metabolizing Enzymes of Lactating Ewes. J. Anim. Sci. 2014; 92:5112–5123.
- 163. Gadberry MS, Denard TM, Spiers DE, Piper EL. Effects of Feeding Ergovaline on Lamb Performance in a Heat Stress Environment. J. Anim. Sci. 2003; 81:1538–1545.
- 164. Tor-Agbidye J, Blythe LL, Craig AM, Correlation of Endophyte Toxins ergovaline and Lolitrem B. with Clinical Disease: Fescue Foot and Perennial Ryegrass Staggers. Vet. Hum. Toxicol. 2001; 43:140– 146.
- Guerre P. Ergot Alkaloids Produced by Endophytic Fungi of the Genus Epichloë. Toxins. 2015; 7:773–790.
- Easton HS, Christensen MJ, Eerens JPJ, Fletcher LR, Hume DE, Keogh RG, et al. Ryegrass Endophyte: A New Zealand Grassland Success Story. Proc. N. Z. Grassl. Assoc. 2001; 63:37–46.
- Springer JP, Clardy J, Wells JM, Cole RJ, Kirksey JW. The structure of paxilline, a tremorgenic metabolite of Penicillium paxilli Bainier. Tetrahedron. Lett. 1975; 16:2531–2534.
- Garcia ML, Galvez A, Garcia-Calvo M, King VF, Vazquez J, Kaczorowski GJ. Use of toxins to study potassium channels, J. Biomembr. Bioenerg. 1991; 23:615.
- 169. Giangiacomo KM, Garcia ML, McManus O. Mechanism of the iberiotoxin block of the large conductance calcium-activated potassium channel from bovine aortic smooth muscle. Biochemistry. 1992; 31:6719.
- Selala MI, Laekeman GM, Loendrs B, Musuku A, Herman A, Schenfens P. In vitro effects of tremorgenic mycotoxins. J. Nat. Prod. 1991; 54:207-212.
- 171. Matsui C, Ikeda Y, Jinuma H, Kushida N, Kunisada T, Simizu S, Umezawa K. Isolation of a novel paxilline analog pyrapaxilline from fungus that inhibits LPS-induced NO production. *The Journal of Antibiotics*. 2014; 67(11):787-790.
- Papavlassopoulos M, et al. MaxiK blockade selectively inhibits the lipopolysaccharide induced I kappa B-alpha/NF-kappa B signaling pathway in macrophages. J. Immunol. 2006; 177:4086–4093.
- 173. Fan Y, Wang Y, Liu P, Fu P, Zhu T, Wang W,Zhu W. Indolediterpenoids with anti-H1N1 activity from the aciduric fungus

Penicillium camemberti OUCMDZ-1492. J. Nat. Prod. 2013; 76: 1328–1336.

- 174. DeFarias F P. Carvalho M F. Lee S H. Kaczorowski G J. Suarez-Kurtz G. Effects of the K+ channel blockers paspalitrem-C and paxilline on mammalian smooth muscle. *European journal of pharmacology*. 1996; 314(1-2):123-128.
- 175. Smith BL, McLeay LM, Embling PP. Effect of the Mycotoxins Penitrem, Paxilline and Lolitrem B on the Electromyographic Activity of Skeletal and Gastrointestinal Smooth Muscle of Sheep. Res. Vet. Sci. 1997; 62:111–116.
- 176. McLeay LM, Smith BL. Effects of the Mycotoxins Lolitrem B and Paxilline on Gastrointestinal Smooth Muscle, the Cardiovascular and Respiratory Systems, and Temperature in Sheep. Ryegrass Endophyte: An Essential New Zealand Symbiosis. Grassland Res. Pract. Ser. 1999; 7:69–76.
- 177. Sanchez M, McManus OB. Paxilline inhibition of the alpha-subunit of the high-conductance calcium-activated potassium channel. Neuropharmacology. 1996; 35:963–968.
- Imlach WL, Finch SC, Zhang Y, Dunlop J, Dalziel JE. Mechanism of action of lolitrem B, a fungal endophyte derived toxin that inhibits BK large conductance Ca²⁺-activated K⁺ channels. Toxicon. 2011; 57:686–694.
- Gant DB, Cole RJ, Valdes JJ, Eldefrawi ME, Eldefrawi AT. Action of Tremorgenic Mycotoxins on GABAA Receptor. Life Sci. 1987; 41:2207–2214.
- Norris PJ, Smith CCT, De Belleroche J, Bradford HF, Mantle PG, Thomas AJ, Penny RHC. Actions of Tremorgenic Fungal Toxins on Neurotransmitter Release. J. Neurochem. 1980; 34:33–42.
- 181. Cole RJ, Kirksey JW, Wells JM. A new tremorgenic metabolite from *Penicillium paxilli*. Can. J. Microbial. 1974; 20:1159-1162.
- Gallagher RT, Keogh RG, Latch GCM, Reid CSW. The role of fungal tremorgens in ryegrass staggers. NZ J. Agric. Res. 1977; 20:431-440.
- 183. Lee US, Cui J. BK Channel Activation: Structural and Functional Insights. Trends Neurosci. 2010; 33:415–423.
- 184. Munday-Finch SC, Wilkins AL, Miles CO, Ede, Thomson RA. Structure Elucidation of Lolitrem F, a Naturally Occurring Stereoisomer of the Tremorgenic Mycotoxin Lolitrem B, Isolated from Lolium Perenne Infected with Acremonium Lolii. J. Agric. Food Chem. 1996; 44:2782–2788.
- Dalziel JE, Finch SC, Dunlop J. The Fungal Neurotoxin Lolitrem B Inhibits the Function of Human Large Conductance Calcium-Activated Potassium Channels. Toxicol. Lett. 2005; 155:421–426.
- 186. McMillan LK, Carr RL, Young CA, Astin JW, Lowe RG, et al. Molecular analysis of two cytochrome P450 monooxygenase genes required for paxilline biosynthesis in Penicillium paxilli, and e_ects of paxilline intermediates on mammalian maxi-K ion channels. Mol. Genet. Genomics. 2003; 270: 9–23.
- Aniszewski T. Alkaloids Secrets of Life. Amsterdam: Elsevier Science. 2007.
- Froehlich KA. Metabolism of loline in ruminants and their potential effects on microflora, and gastrointestinal nematodes. In Animal Science; Lincoln University: Lincoln, New Zealand, 2020; p. 120.
- Lai SK, Wang YY, Hanes J. Mucus-penetrating nanoparticles for drug and gene delivery to mucosal tissues. Adv. Drug. Deliv. Rev. 2009; 61:158–171.
- 190. Rudolph W, Remane D, Wissenbach DK, Peters FT. Development and validation of an ultrahigh performance liquid chromatographyhigh resolution tandem mass spectrometry assay for nine toxic alkaloids from endophyte-infected pasture grasses in horse serum. J. Chromatogr. A. 2018; 1560:35–44.
- TePaske MR, Powell RG, Clement SL, Quantitative analyses of bovine urine and blood plasma for loline alkaloids. J. Agric. Food Chem. 1993; 41:231–234.
- Froehlich KA.Metabolism of loline in ruminants and their potential effects on microflora, and gastrointestinal nematodes. PhD thesis submitted to Lincoln University. 2020; pp: i-iii.
- 193. Seawright A, Hrdlicka J, Wright J, Kerr D. The identification of hepatic pyrrolizidine alkaloids in horse by the demonstration of sulfur bound pyrrolic metabolites to their haemoglobin. *Veterinary and Human Toxicology*. 1991; 33(3):286-287.
- 194. Gooneratne SR, Patchett BJ, Wellby M, Fletcher LR. Excretion of loline alkaloids in urine and faeces of sheep dosed with meadow fescue (Festuca pratensis) seed containing high concentrations of loline alkaloids. N. Z. Vet. J. 2012; 60:176–182.
- 195. Westendorf M, Mitchell G, Tucker R, Bush L. In Vitro and In Vivo Ruminal and Physiological Responses to Endophyte-Infected Tall Fescue. *Journal of Dairy Science*. 1993; 76:555-563.
- 196. Rudolph W, Remane D, Wissenbach DK, Peters FT. Liquid chromatography-mass spectrometry-based determination of

ergocristine, ergocryptine, ergotamine, ergovaline, hypoglycin A, lolitrem B, methylene cyclopropyl acetic acid carnitine, N-acetylloline, Nformylloline, paxilline, and peramine in equine hair. *Journal of Chromatography B*,2019; 1117:127-135.

- 197. Grancher D, Durix A, Moulard Y, Bonnaire Y. Carcelen M. Camier Y. Bony S. Distribution Du Lolitrème B et de L'ergovaline Après Un Bolus Intraruminal Chez La Chèvre En Lactation; Rencontres Recherches Ruminants: Paris, France, 2004; Volume 11, p. 37.
- Mantle PG, Burt SJ, Bilton JN. The role of the ryegrass endophyte (Acremonium loliae) in ruminant neurotoxicosis. Proc. Jpn. Assoc. Mycotoxicol. 1988; (Suppl. 1):115-116.
- 199. Cheeke PR. Pyrrolizidine alkaloid toxicity and metabolism in laboratory animals and livestock. In: Cheeke P R. (Editor), Toxicants of Plant Origin. Vol I. Alkaloids. CRC Press, Boca Raton, FL, 1989; pp. 1-22.
- Ruan J, Yang M, Fu P, Yang Y, Lin G. Metabolic activation of pyrrolizidine alkaloids: insights into the structural and enzymatic basis. *Chemical Research in Toxicology*. 2014; 27:1030-1039.
- Stegelmeier BL, Edgar JA, Colegate SM, Gardner DR, Schoch TK, Coulombe RA, Molyneux RJ. Pyrrolizidine alkaloids plants, metabolism and toxicity. *Journal ofNatural Toxins*. 1991; 8(1):95-116.
- 202. Mattocks AR. Chemistry and toxicology of pyrrolizidine alkaloids. Orlando: Academic Press. 1986.
- Dominguez-Bello MG. Detoxification in the rumen. Annales De Zootechnie. 1996; 45(suppl):323-327.
- 204. Mattocks RA, Jukes R. Recovery of the pyrrolic nucleus of pyrrolizidine alkaloid metabolites from sulphur conjugates in tissues and body fluids. *Chemical-BiologicalInteractions*. 1990; 75:225-239.
- 205. Jackson JA, Varney DR, Petroski RJ, Powell RG, Bush LP, Siegel MR, Hemken RW, Zavos PM. Physiological responses of rats fed loline and ergot alkaloids from endophyte-infected tall fescue. Drug and Chemical Toxicology. 1996; 19(1&2):85-96.
- 206. Wang T, Frandsen HL, Christiansson NR, Rosendal SE, Pedersen M, Smedsgaard J. Pyrrolizidine alkaloids in honey: Quantification with and without standards. Food Control. 2019; 98:227–237.
- 207. König J, Fuchs B, Krischke M, Mueller MJ, Krauss J. Hide and seek—Infection rates and alkaloid concentrations of Epichloe festucae var. lolii in Lolium perenne along a land-use gradient in Germany. Grass Forage Sci. 2018; 73:510–516.
- 208. Byam W, Archibald RG. The Practice of Medicine in Tropics. Frowde, Hodder. and Stoughlon, London (1921-1923).
- Watt JM, Breyer-Brandwijk MG. The Medicinal and Poisonous Plants of Southern and Eastern Africa. E. & S. Livingstone, London. 1962.
 2nd edition.
- 210. Katz I. Contribution à l'étude de l'ivraie eniurante. Phytopathol. Z. 1949;15: 495.
- 211. Casabuono AC, Pomilio AB. Alkaloids from endophyte-infected *Festuca argentina*. Journal of Ethnopharmacology. 1997; 57:1-9.
- Finch S, Munday J, Munday R. Kerby J. Short-term toxcity studies of loline alkaloids in mice. Food and Chemical Toxicology. 2016; 94:243-249.
- Vikuk V, Young CA, Lee ST, Nagabhyru P, Krischke M, Mueller MJ, Krauss J. Infection rates and alkaloid patterns of different grass species with systemic Epichloë endophytes. Appl. Environ. Microbiol. 2019; 85:e00465-19.
- Gallagher RT, Campbell AG, Hawkes AD, Holland PT, McGaveston DA, Pansier EA, Ryegrass staggers: The presence of lolitrem neurotoxins in perennial ryegrass seed. N. Z. Vet. J. 1982; 30:183– 184.
- 215. Philippe G. Lolitrem B and indole diterpene alkaloids produced by endophytic fungi of the genus Epichloë and their toxic effects in livestock. *Toxins*. 2016; 8(2):47.
- 216. Craig AM, Blythe LL, Duringer JM. The role of the Oregon State University Endophyte Service Laboratory in diagnosing clinical cases of endophyte toxicoses. J. Agric. Food Chem. 2014; 62:7376–7381.
- 217. Miles CO, Lane GA, Di Menna ME, Garthwaite I, et al. High levels of ergonovine and lysergic acid amide in toxic Achnatherum inebrians accompany infection by an Acremonium-like endophytic fungus. J. Agric. Food Chem. 1996; 44:1285–1290.
- Rowan DD, Shaw GJ. Detection of ergopeptine alkaloids in endophyte-infected perennial ryegrass by tandem mass spectrometry. N. Z. Vet. J. 1987; 35:197–198.
- 219. Fletcher LR, Garthwaite I, Towers NR. Ryegrass staggers in the absence of lolitrem B. In Proceedings of the Second International Symposium on Acremonium/Grass Interactions; Hume D E, Latch G C M, Easton H S, Eds.; AgResearch: Palmerston North, New Zealand, 1993; pp.119–121.

- Saikkonen K, Lehtonen P, Helander M, Koricheva J, Faeth SH. Model systems in ecology: Dissecting the endophyte-grass literature. Trends Plant Sci. 2006; 11:428–433.
- 221. Rowan DD, Dymock JJ, Brimble MA. Effect of fungal metabolite peramine and analogs on feeding and development of Argentine stem weevil (*Listronotus bonariensis*). J. Chem. Ecol. 1990; 16:1683-1695
- Vamey DR, Prestidge RA, Jones DD, Varney LA, Zavos PM, Siegel MR. Effect of endophyte-infected perennial ryegrass seed diets on growth and reproduction in mice. NZ J. Agric. Res. 1989; 32:547-554.
- Cunningham IJ, Hartley WJ. Ryegrass staggers. NZ Vet. J. 1959; 7:1-7.
- 224. Aboling S, Drotleff AM, Cappai MG, Kamphues J. Contamination with ergot bodies (Claviceps purpurea sensu lato) of two horse pastures in Northern Germany. Mycotoxin Res. 2016; 32:207–219.
- Lorenzo-Velazquez B, Dominguez L A. Estudio experimental sobre la ototoxicidad de los fármacos: y sal sódica del acido fusídico. Arch. Inst. Farmacol. Exptl. (Madrid); 1965; 17(1):83.
- Page MG, Ankona-Sey V, Coulson WF, Bender DA. Brain glutamate and gamma-aminobutyrate (GABA) metabolism in thiamin-deficient rats. Br. J. Nutr. 1989; 62:245-253.
- Porter JK, Stuedemann JA, Thompson Jr. FN, Lipham LB. Neuroendocrine measurements in steers grazed on endophyte-infected fescue. J. Anim. Sci. 1990; 68:3285-3292

